ASSESSMENT OF MOLECULARLY IMPRINTED POLYMERS AS SUSTAINABLE PHOSPHATE

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ABSTRACT

Wastewater effluents and agricultural runoff are major sources of phosphorus overloading in surface waters. Phosphorus overloading ignites eutrophication, which devastates aquatic ecosystems. On the other hand, phosphorus, which is currently produced from phosphate rock, is a critical component of fertilizer mixes. However, the world is predicted to face a shortage of phosphate supply beyond 2033 due to unsustainable mining. This research aims to develop a polymeric sorbent that recovers low-concentration phosphorus for eutrophication prevention and fertilizer reuse. Available polymer-based products have underwhelmed expectations by having poor selectivity or lacking appropriate biodegradation rates. This research identified molecularly imprinted polymers (MIPs) as possible sorbents for overcoming the deficiencies of reported technologies. Screening of several MIPs resulted in one potentially feasible MIP for phosphate sorption. Further studies showed a sorption capacity of $\sim$28 mg PO_4^{3-}-P/g and partial phosphate-selectivity. Potential phosphate removal mechanisms were identified, providing foresight into MIPs' viability as phosphorus sorbents.

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DEDICATION

To the Baker Near-Wilderness Settlement for sparking my inspiration in environmental

stewardship, and for continuing to provide the same opportunities to children across the

state of Minnesota.

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CHAPTER 1. LITERATURE REVIEW

1.1.   Introduction

Phosphorus (P) is an element that has raised a lot of attention in research with its

contradictory influences in the environment and agriculture. P is a nonmetal of the

nitrogen group and is an essential macronutrient for all life on earth (Desmidt et al., 2014,

You et al., 2015). P is primarily found as phosphate (PO_4^{3-}) instead as a free element due to

its high reactivity (Desmidt et al., 2014). The role P plays as an essential macronutrient

gives it an extremely important role in agriculture. However, the supply of mineral P is

finite and is concentrated in only a few countries (Rittmann et al., 2011). The varying

control of P between countries could result in political strife as Morocco possesses

dominant control over the Western Saharan reserves, being the largest in the world. China

is already drastically reducing their exports of P to try and secure future domestic supply,

the United States is estimated to have less than 30 years of domestic supply left, and

Western Europe and India are already completely dependent upon P imports (Cordell et al.,

2009). This mineral P is mined as a phosphate rock, and it is predicted that deposits of this

mineral will only last for another 60 – 240 years (Cornel and Schaum, 2009, USGS, 2012).

The vast majority of mined phosphate rock around the globe is used in agricultural

products or combined with sulphuric acid, nitrogen and potassium for fertilizer-based

applications (Desmidt et al., 2014, Cordell et al., 2009). Currently, the world mines about

180 – 190 million tons of phosphate rock annually. Initial observations of trends in

phosphate mining indicated that the peak amount of mined phosphate rock was reached in

2009. An upward trend has been expected to increase by 50 – 100% by 2050 with an

increase in global food demand, changing population diets, and increased usage of plant

derived biofuels (Desmidt et al., 2014, Cordell et al., 2009). At the current rate of mining

alone, existing phosphate rock reserves would be fully depleted in approximately 369 years

(time approximation based on adjusted value reported (Desmidt et al., 2014)). As

production and demand continue to increase, the estimated period will likely be less than

369 years. With over 800 million people without sufficient access to food, ensuring food

security is considered a global priority (UN, 2000, 2005). Reduction in fertilizer supply can

result in reduced crop yields, which will make it impossible to meet the world's increasing

food demands (Desmidt et al., 2014). Conservative analysis done by Cordell et al. (2009)

predicts that the peak in global P production could occur by 2033. A peak in P production

does not mean complete depletion of supply, but it is indicative of the depletion of high

quality and easily accessible reserves (Cordell et al., 2009). After this point, the quality of

the remaining reserves becomes lower with harder accessibility, resulting in uneconomical

mining and processing (Cordell et al., 2009). Along with an increasing demand for

phosphate rock, the cost of mining phosphate rock is increasing due to the decline in the

quality of phosphate rock present in easily accessible mines and extracting higher quality

phosphate rock from deeper layers is relatively expensive (Desmidt et al., 2014).

 While short supply of P can cause reduced crop productivity, excessive loading of P

into surface waters can promote eutrophication of lakes, rivers, reservoirs, estuaries, and

parts of the oceans (USEPA, 1983, 1998). Eutrophication is the process in which aquatic

ecosystems experience an increase in the abundance of algae and aquatic plants. These

organisms die after their life cycle is complete, allowing for microorganisms to degrade

their remnants. This biodegradation results in the microorganisms consuming oxygen from

the aquatic ecosystem and thus depleting oxygen from the rest of the water body. A lack in

adequate levels of oxygen often leads to stress and death of important ecosystem

components. Altered nutrient regimes are one of the many stressors leading to species

endangerment and P is the key growth-limiting nutrient for algae in this eutrophication

process, which leads to an overall degradation of precious water resources (Smith, 2003,

Dodds et al., 2009). In fact (USEPA, 1996) reported that eutrophication accounted for

nearly half of impaired lakes and 60% of impaired river reaches within the United States.

The intensive nature of agriculture to meet global food demands has led to an increase in

fertilizer application on croplands to improve profitability. This intensification has been a

major influence in the eutrophication rates of aquatic ecosystems as more than half a

century of generous applications of P and nitrogen (N) fertilizers has resulted in North

American agricultural soils having surpassed 'critical' P levels (Smith, 2003, Cordell et al.,

2009, McDowell et al., 2016). This overly nutrient-rich cannot contain much of the

additional P which is applied annually (McDowell et al., 2016). Approximately 80% of

mined P is used in agricultural fields, but only about 40% of what is applied is actually

available for crops to uptake. The rest of the P, that is not immobilized in soil, results in P-

rich runoff and leads to impairment of surface water quality (Rittmann et al., 2011, USGS,

2010).

Eutrophication is not only extremely detrimental to aquatic ecosystems, but also to

the economic well-being of communities. Lakes and rivers provide drinking water,

recreation, and aesthetic benefits, all of which can be negatively influenced by

eutrophication. Lakes contain up to 68% of the global liquid surface fresh water are

essential for economic vitality and quality of life across the globe (Beeton, 2002). These

water sources, which often support municipal water supplies, can exhibit an increase in the frequency and severity of potentially toxic cyanobacterial blooms under eutrophic conditions (Dodds et al., 2009). Toxic algal growth can lead to human health problems and is not aesthetically pleasing. Additionally, the detrimental effect of algal growth toward the surface water's viability as drinking water supply results in higher water treatment costs (Smith, 2003). Recreational use of such water bodies is reduced as angling and boating activities can be physically impeded while other water users are less likely to swim during heavy algal blooms due to health risks, unfavorable appearance, or unpleasant odors. Reduction in these recreational and aesthetic characteristics of the water body can also decrease shoreline property values. Combined losses due to freshwater eutrophication are approximately \$2.2 billion annually in the United States (Dodds et al., 2009). USGS (2010) found that of 190 streams studied in the United States, which are in agricultural and urban watersheds, 90 percent of them exceeded natural background levels (34 µg/L) of orthophosphate and total phosphorus (TP). The streams in agricultural and urban areas were found to have a median TP concentration of 0.25 mg/L, which is about 6 times greater than background levels. Two agricultural practices were found to significantly contribute to the transport of nutrients in streams within the United States. First, the prevalent use of tile drains and ditches in the Midwest, which help to drain clay-rich soils, facilitates the immediate transport of nutrients to streams. Secondly, the large amount of irrigation-return flow contributes to the transport of nutrients. Most P entering surface waters comes via erosion and runoff from non-point sources (such as agricultural croplands and urban areas), and has been identified as the primary reason that approximately 52% of monitored rivers, lakes, and estuaries are not clean enough for recreational uses (USEPA,

2015). However, point sources, such as wastewater effluent, have also been identified as significant contributors to excessive P loading and eutrophication in surface waters as approximately 8% of globally mined P is discharged from wastewater treatment plants (Rittmann et al., 2011). The P concentration discharged from wastewater effluent generally ranges from 5 – 10 mg/L, which is far higher than the allowable level in surface waters. Concern about wastewater effluent causing rapid rates of eutrophication has resulted in the United States establishing a goal of regulating P limits in treated wastewater effluent to 10 μg/L by 2027 (Hansen, 2006). Reaching this goal will reduce point source pollution below excessive loading rates, allowing natural water bodies to self-regulate and maintain healthy ecosystems.

1.2. Established P Reduction Plans and Technologies for Non-point and Point Sources, and *in-situ* Lake Remediation

The problems associated with phosphate pollution have been long recognized by the United States and other countries, and there are established plans and technologies to mitigate them.

1.2.1. Non-point Sources

Non-point sources of P pollution are difficult to treat as they come from extended areas, and sometimes their sources are not clearly identified. Best Management Practices (BMPs) such as infiltration basins, rain gardens, constructed wetlands, and soil amendments are accepted methods of water pollution control for non-point sources. BMPs can be effective for non-point sources but cannot remove all of the P transported from an area. Additionally, some BMPs suffer from site-specific design restraints, low P capacity, and desorption/leaching of P (Perry et al., 2009, USDA, 2006, Robalds et al., 2015).

Many states in the United States have begun to develop proactive means to reducing

P loading by adopting Total Maximum Daily Loads (TMDLs) and the National Pollution

Discharge Elimination System (NPDES) regulations. However, non-point sources such as

agricultural runoff are exempt from NPDES programs and TMDL regulations. Policies and

guidelines have been set by the United States Department of Agriculture (USDA) for

nutrient management in agricultural areas. States generally customize their own

approaches and procedures that vary from purely voluntary measures (nutrient

management planning, soil testing, and conservation adoption) to more restrictive

requirements. Voluntary efforts and fee paying schemes have been pushed by some states.

Nevertheless, land use practice that leads to substantial losses of P into surface runoff still

occur (McDowell, 2016). This wide range of flexibility makes non-point source P mitigation

highly variable from state to state and an unreliable system for P removal in the United

States.

1.2.2. Point Sources

On a regulatory level, TMDLs can be established for all point sources and may be

enforceable. Specifically, a TMDL identifies the pollutant load reduction needed so that a

waterbody can maintain water quality standards. NPDES is also viable for point source

regulation with permits to limit P discharges from the sources (McDowell, 2016).

The primary technologies that are currently used for removing P from point sources, such

as wastewater, are chemical precipitation and biological remediation (Rittmann, 2011,

Jenkins and Hermanowicz, 1991, Stensel, 1991, Pan et al., 2009). Chemical precipitation of

P can be achieved through the addition of cations such as iron, aluminum, calcium,

magnesium, or ammonium to produce a chemical sludge which is then removed from

wastewater effluent and disposed in landfills (Rittmann et al., 2011). Biological remediation of P is an ecological approach which produces P-containing biosolids as polyphosphate accumulating organisms (PAOs). These biosolids are removed from wastewater effluent and can be applied as a soil amendment for cropland or disposed in landfills (Oehmen et al., 2007, Rittmann et al., 2011, Ekama, 2015). These conventional methods tend to be technically difficult or economically unsustainable to meet increasingly stringent P discharge limits. Precipitation and biological methods for P removal are inadequate at reducing P concentrations below environmentally significant levels (100 µg/L) and only well-suited for high P concentration waste streams (Pan et al., 2009, Chouyyok et al., 2010, You et al., 2015). Other issues with these conventional methods involve sludge disposal/recovery problems, and sensitivity of the processes to P loading rates and temperature (Jenkins and Hermanowicz, 1991, Stensel, 1991).

1.2.3. *In-situ* Lake Remediation

Several methods for *in-situ* removal of P from lakes have been developed over the years. Some of the physico-chemical methods include sediment dredging, hypolimnetic withdrawal, hypolimnetic aeration, and phosphorus inactivation. Sediment dredging can remove internal nutrient loadings from lake systems, but it also destroys the sediment-water interface which is known to play a key role in the stabilization of lake systems. This dredging acts as only a brief moment of relief as the lake can be reloaded with P over time from pre-existing external sources. Therefore, mitigation of pre-existing external P sources is necessary to achieve long-term desired effects with dredging (Jing et al., 2015). The hypolimnion is the lowest layer of water to come in contact with the sediment after lake stratification and it often contains the highest P concentration. Thus, treating or removing

this layer of P-rich water through physical methods has been identified as a viable strategy for lake remediation. Hypolimnetic withdrawal and aeration have been practiced, however these hypolimnetic altering methods can be inefficient, cost prohibitive, or cause excessive disturbance to the ecosystem. Phosphorus inactivation is a common chemical restoration method. Phosphorus inactivation caps the P-rich sediment with a stable material that can bind with P. Alum is one of the most commonly used materials for this application. Issues with capping materials such as alum are its inefficiency over a wide range of pH and release of toxic materials via hydrolysis after long residence in water (Goldyn et al., 2014, Lewtas et al., 2016, Singh, 1982).

1.3.   Developing P Reduction Technologies

1.3.1. Waste By-products

Waste by-products have been extensively investigated for P removal due to their great economic feasibility. They are low-cost materials which are typically easily acquirable and renewable. Their use can also reduce imminent disposal costs (Boyer et al., 2011). Examples of waste by-products used are cationized wood residues (Unnithan et al., 2002, Tshabalala et al., 2004, Karthikeyan et al., 2004), iron oxide tailings (Zeng et al., 2004), alum and ferric sludge (Babatunde and Zhao, 2010, Song et al., 2011), alkaline fly ash (Cheung and Venkitachalam, 2000), egg-shell waste (Yeddou Mezenner and Bensmaili, 2009), red mud (Huang et al., 2008), blast furnace slag (Oguz, 2004), and waste lime (Siobhan Dunets and Zheng, 2014, Ahn and Speece, 2010). Some concerns with these low-cost materials are that they are typically not homogenous, have a wide range of P removal efficiencies, and may cause undesirable secondary changes in water chemistry.

1.3.2. Ion and Ligand Exchangers

Ion exchangers have been found to have extremely high P recovery potentials. Ion exchangers allow P to be recovered as calcium phosphate or struvite upon the addition of calcium nitrate or ammonium chloride and magnesium sulfate (Sengupta and Pandit, 2011). This recovered material has potential for reuse as fertilizer. Many researchers looked into developing fixed-bed processes for P removal due to their technical simplicity and adaptability to fluctuations in the composition and flow rates of effluent wastewater (Blaney et al., 2007). Polymeric materials were viewed as the optimal material for developing fixed-bed processes as they offer the desired durability and mechanical strength. However, commercially available anion exchangers still suffer from a lack of P selectivity, poor mechanical strength, low regeneration efficiency, and high cost. Some studies have reported the development of polymeric ligand exchangers (PLEs) which have shown very high P selectivity. Nevertheless, these PLEs require a chelating polymer which is cost prohibitive (Blaney et al., 2007, Choi et al., 2012). Research has also branched into incorporating nanoparticles (NPs) in anion exchangers for P removal due to the extremely high surface area to volume ratio that NPs offer. These characteristics can produce fast kinetics and enhanced sorption capacity. While these nano-based anion exchangers were shown to be highly regenerable, they suffered from poor P selectivity. NPs are not suitable for direct application in fixed-bed columns due to their poor durability, agglomeration, and contribution toward excessive pressure drop across the column (Sarkar et al., 2011).

1.3.3. Membranes

Membrane technology has also been studied for P removal from wastewater. P removal using membrane bioreactors (MBRs) and reverse osmosis/nanofiltration (RO/NF)

membrane systems are the predominant methods so far studied (Park et al., 2017, Luo et al., 2016, Dolar et al., 2011). MBRs are a promising technology as they have a small physical footprint and high biomass retention (Park et al., 2017). MBRs showed to recover P from permeate as amorphous calcium phosphates under high pH. However, a common issue with membrane use in wastewater is its tendency to foul with an excessive build-up of organic matter and ammonia in the draw solution. This fouling extremely limited the MBR's efficiency when removing P (Luo et al., 2016). Additionally, MBRs long solids retention time with large quantities of biomass reduces effective P control as P removal primarily relies on sludge waste (Park et al., 2017). RO/NF treatment also showed high retention of fluoride and phosphate ions, but also faced the same complications as previously discussed with MBRs (Dolar et al., 2011).

1.3.4. Adsorbents

Adsorption-based processes have been among the most heavily studied for P removal. Adsorption refers to the transfer of liquid-phase solutes to solid-phase adsorbents (Crittenden, 2005). P separation by adsorption during coagulation, followed by removal through sedimentation and filtration was first used in the 1950s and has since been applied to treat water and wastewater using a variety of sorbents (Rittmann et al., 2011). Adsorption processes have advantages over other techniques in terms of infrastructure cost, modular design, simplified technological design, higher P removal rates, and low-cost operation (Sowmya and Meenakshi, 2014, Pan et al., 2009, Chouyyok et al., 2010). Though there are many characteristics that make adsorption processes highly beneficial, cost effectiveness is identified as the primary criterion in the selection of an adsorption technology for P removal (Mishra, 2010). Adsorption processes regulate the mobility of

aqueous anions by competing with coexisting anions for the adsorbent surface binding sites. The capacity of adsorbents depends on the number active binding sites, surface area-to-volume ratio and porosity (Nthumbi et al., 2012). Many adsorbents, both polymeric and non-polymeric, incorporate immobilized metals because they can develop inner-sphere complexes with phosphate. In fact, formation of inner-sphere complexes between transition metals and anions is more preferential toward phosphate than other inorganic anions (Zhu and Jyo, 2005). While this does not give transition metals explicit selectivity toward P removal, it does allow them to overcome some interference issues with coexisting anions.

1.3.4.1. Non-polymeric Adsorbents

There is a wide sampling range of non-polymeric adsorbents. Hydrated ferric oxides (Pan et al., 2009, You et al., 2015, You et al., 2016), magnetic-based adsorbents (Janos et al., 2013, Lai et al., 2016, de Vicente et al., 2011), layered double hydroxides (Das et al., 2006, Cheng et al., 2009), synthesized minerals (Peleka and Deliyanni, 2009, Chitrakar et al., 2006), hydroxides (Guan et al., 2006), modified clays (Copetti et al., 2016, Reitzel et al., 2013), modified biochars (Jung and Ahn, 2016, Li et al., 2016), oxides (Xie et al., 2015, Park et al., 2017), immobilized transition metals (Mahaninia and Wilson, 2015, An et al., 2014, Ogata et al., 2011, Liu and Zhang, 2015), and nanoparticles (Zheng et al., 2016, Chouyyok et al., 2010, You et al., 2016, Almeelbi and Bezbaruah, 2012, Lai et al., 2016) have been extensively studied for P removal. Many of these materials have shown high P selectivity, high P sorption capacity, environmentally benign, stable under a wide pH range, low cost, fast removal rate even at low P concentrations, or P recovery capabilities.

1.3.4.2.   Polymeric Adsorbents

In this section, polymeric adsorbents are defined as adsorbents where the polymer(s) are the responsible component for P removal. Polymeric adsorbents have garnered a lot of attention due to their advantageous properties. Polymeric adsorbents can remove low concentration P and are cost effective in terms of regeneration and disposal. Additionally, they can be easily modified to better fit varying applications (Kioussis and Kofinas, 2005a, 2005b, Mahaninia and Wilson, 2016). Polymeric adsorbents also tend to be non-toxic, environmentally stable, and require minimal effort or capital to prepare (Hammud et al., 2015).

Several studies developed hydrogels for P remediation from wastewater. Kioussis and Kofinas (2005a) developed a cross-linked cationic hydrogel which could bind to anions in contaminated slurries. Ionic polyamine hydrogels have also been developed to remove anions from wastewater and were found to have high sorption capacities. Gel morphology and formation were found to influence the cross-linking density/uniformity as well as the swelling response, which results in changes in their anion binding capacity (Kioussis and Kofinas, 2005b, Kofinas and Kioussis, 2003). Chitosan, a widely studied and well-known polymer, was shown to have a capacity for P adsorption after modification. A study conducted by Mahaninia and Wilson (2016) found that cross-linking chitosan with glutaraldehyde and epichlorohydrin at varying compositions introduced new functional groups to chitosan's surface which allowed it to remove P. This modified chitosan polymer was also found to remove P effectively in highly alkaline pH conditions, a feat which is often difficult to be found in other polymer adsorbents. Another study reported the copolymerization of chitosan with polyacrylamide to remove P. However, the resulting

sorption capacity was too low to be considered feasible (Nthumbi et al., 2012). Electrostatic interactions and ion exchange are the believed modes of interaction between polymers such as chitosan and phosphate that contribute to adsorption. Another potential mechanism could be hydrogen bonding between the polar functional groups of chitosan with the donor-acceptor groups of phosphate (Mahaninia and Wilson, 2016). Polypyrrole is a polymer shown to have very high P sorption capacity, but its removal at concentrations below 10 mg/L were unproven (Hammud et al., 2015). Aminopolystyrene copolymerized with 4-amino-N-azobenzenesulfamide was another polymer shown to have an affinity toward P (Basargin et al., 2007).

1.3.5. Limitations

Many of the developing technologies have an abundance of potential for being viable options for P removal. However, none of these technologies currently succeed in meeting all of the desired criteria for an optimal P adsorbent such as:

- P selectivity
- P removal from concentrations below 100 μg/L in solution
- Cost effective
- Environmentally stable and benign
- High P sorption capacity
- Stable and effective over a wide range of pH
- Fast removal rate
- P recoverability and reusability

The most common setback holding these technologies from becoming industrially and commercially effective is P selectivity (Chitraka, 2006, Saha, 2009). Many of these technologies, such as ones that utilize transition metals, witness high P sorption capacities, but are not truly selective toward P as they also bind with many other anions. This lack of selectivity can lead to a reduction in P sorption capacity and overall interference with P

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removal from coexisting anions. Technologies that were selective toward P, such as PLEs, were far too expensive to be economically viable options (Blaney et al., 2007, Choi et al., 2012). Many technologies cannot remove P below 100 µg/L, which is the typical level of P in eutrophic lakes. This is a significant issue as 50 µg/L has been identified as the level of P needed for which eutrophication can ignite (Smith, 2003). While these technologies can remove P, many of them are not able to remove it in a recoverable form for reuse in fertilizer. The importance and current trend toward recoverability will be discussed further in the following section.

1.4.   Trend toward Sustainability

As discussed in Section 1.1, P supplies are dwindling while demand is trending upward. Conventional approaches to finding new P supplies are to continue exploration for new phosphate rock mines and further intensify the exploitation of existing mines. This also includes expanding more into off-shore and low grade deposits. However, this approach fails to address the issue of the finite nature of phosphate rock reserves in the long term (Cordell et al., 2009). A primary goal toward solving this issue was identified as generating non-conventional P sources. Capturing P from municipal effluent, surface runoff, and other waste products have been recognized as the most viable means for reaching this goal (Rittmann et al., 2011, Cordell et al., 2009). In order to accomplish this, P removal technologies can no longer be optimized for just pollution prevention. The resultant product will need to be suitable for use in agriculture and other applications (Cordell et al., 2011). P removal only aims at obtaining a P-free effluent by transferring P to sludge or some other media for disposal. P recovery aims at obtaining a P-free effluent while producing a P-containing product that can be reused either in agriculture or in the P

industry (Desmidt et al., 2014). Of late, P recovery from wastewater has been the P source to receive the most attention due to its higher relative concentration and ease of implementing technology changes/upgrades. However, eutrophic lakes are also being looked into as veritable sources for P recovery.

For P to be reusable, recovery methods must produce products that have P which is biochemically available to plants and preferably contains the appropriate nitrogen:phosphorus:potassium (N:P:K) ratio that is found in common fertilizers. Other parameters for successful recovery include the P-containing product to be non-odorous, socially acceptable, and logistically feasible for field application (Cordell et al., 2011). Metal phosphate salts, such as iron or aluminum, have been reported to be implausible for reuse in agriculture by virtue of being biochemically unavailable for plants under normal pH conditions (Desmidt et al., 2014). On the other hand, it has been reported by Almeelbi and Bezbaruah (2012) that iron phosphate salts formed from nanoscale zero-valent iron (NZVI) were in fact bioavailable to plants and even helped fortify plants that were deficient of iron. Therefore, it is still unclear as to whether or not recovery technologies which result in metal phosphate salts will be feasible for fertilizer reuse. Additionally, heavy metals and toxic organics which can be removed by non-selective technologies, such as transition metals, can be detrimental to plant health. This makes wastewater, which often contains these harmful contaminants, a questionable source for P recovery and reuse (Desmidt et al., 2014). Only technologies that selectively bind P, and none of the potentially harmful contaminants, would be appropriate for use in wastewater. This selectivity is not as essential for removal from eutrophic lake water as they typically do not contain the same harmful contaminants.

Further research is still required to determine the most sustainable means for recovering P and reusing the final product as a fertilizer. Such research needs to take into account life cycle costs, energy consumption, bioavailability, farmer accessibility, resource use, and potential pollution from the product (Cordell et al., 2011).

1.5. Need Statement

The phosphorus cycle in today's society (**Scheme 1**) is unsustainable. To develop a more sustainable cycle, P needs to be reclaimed from non-conventional sources and reused for agricultural and industrial applications (**Scheme 2**). There are other factors associated with the phosphorus cycle which need to be improved to develop a sustainable system, but recovery and reuse from surface waters has been identified as one of the methods to reduce human dependence on phosphate rock (Desmidt et al., 2014). In order to reach this goal, a technology must be developed that can successfully recover P from natural water bodies that will make P bioavailable for plants as fertilizer. Selective removal of P is necessary in order to make wastewater a usable source of P recovery.

Without selective removal, P recovered from wastewater runs the risk of having high associations with toxic heavy metals which tend to coexist in wastewater sources. The presence of those toxic metals would reduce the applicability of the recovered P to fertilizer application as they can have severely detrimental effects on overall plant growth and health. There are many coexisting constituents other than heavy metals found in wastewater and eutrophic lakes which, in large quantities, could also have detrimental effects on crops. Therefore, a technology with selective removal of P is ideal for producing a usable recovery product. Many of the technologies available (i.e. bioremediation, chemical precipitation, etc.) cannot remove P below levels which are commonly found in eutrophic

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lakes (100 μg/L). The lack of this capability not only makes it extremely difficult to restore the health of important surface waters, but it also eliminates another major source of recoverable P for reuse in fertilizer applications. In addition to these two innately important criteria for successful P recovery and reuse, the technology must have sorption capacities that are competitive enough to be considered economically justifiable for use. A sorption capacity of approximately 10 – 26 mg P/g can be considered as competitive with many of the technologies currently available.
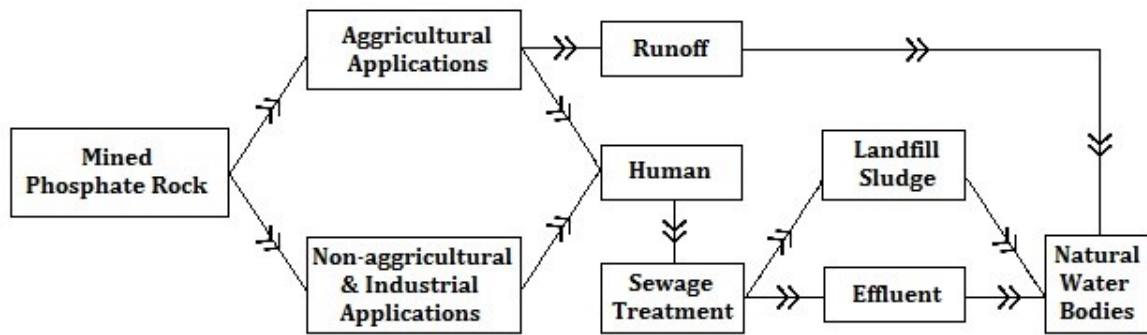
Scheme 1. Current non-sustainable phosphorus cycle in modern society (modified after (Desmidt et al., 2014))
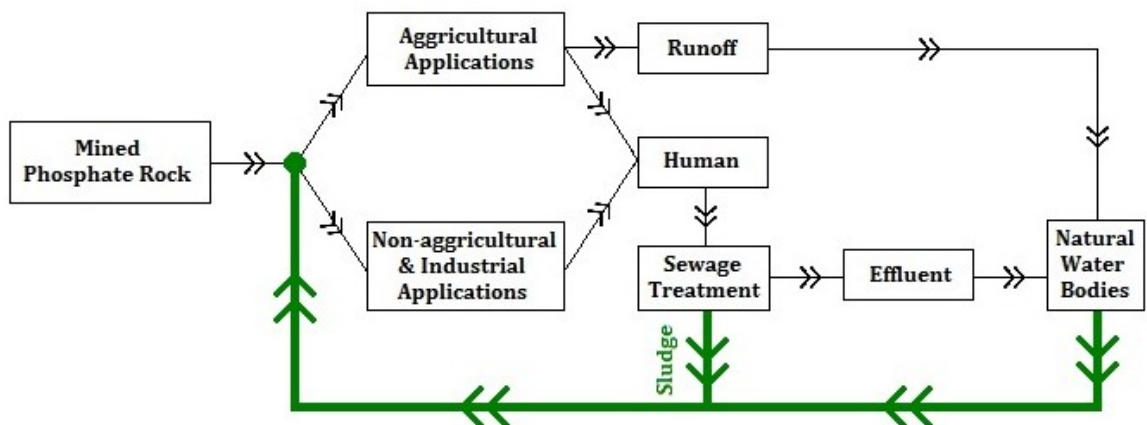
Scheme 2. Desired and sustainable phosphorus cycle for future societies (modified after (Desmidt et al., 2014))

The goal of this research was to assess the feasibility of using molecularly imprinted polymers as recoverable and reusable P sorbents. These polymers show potential to provide selective removal of P at environmentally significant concentrations. Further discussion on molecularly imprinted polymers can be seen in Section 1.6.

1.6.   Molecularly Imprinted Polymers

Molecular imprinting technology (MIT) has been developed as a viable synthetic approach to design robust molecular recognition materials. The design of synthetic materials, which can mimic recognition processes found in nature, is a well-studied area of research in recent years. Molecularly imprinted polymers (MIPs) are versatile materials which have shown to be able to recognize both biological and chemical molecules. Some molecules that have been thoroughly studied for recognition with this technology are proteins, nucleotide derivatives, pollutants, drugs, and food. This has led to MIPs having promising applications in separation sciences and purification (Vasapollo et al., 2011, Khorami and Edrisi, 2010, Amut et al., 2010, Lee et al., 2010), solid phase extraction (Vasapollo et al., 2011, Bravo et al., 2007, Al-Degs et al., 2009, Alexiadou et al., 2008, Tse Sum Bui and Haupt, 2010, Ensing et al., 2002), chemical sensors (Vasapollo et al., 2011, Greene and Shimizu, 2005, Lakshmi et al., 2009), catalysis (Vasapollo et al., 2011), drug delivery (Vasapollo et al., 2011, Alvarez-Lorenzo and Concheiro, 2004), biological antibodies (Vasapollo et al., 2011), receptor systems (Vasapollo et al., 2011), and recently as adsorbents (Lee and Doong, 2012).

The main advantages of MIPs are their high selectivity and affinity for the target molecules, excellent environmental stability, low cost, and relative ease to design, synthesize, and modify (Vasapollo et al., 2011, Wu, 2012). Properties of MIPs that

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contribute to their excellent environmental stability are their high physical robustness, strength, and inertness towards acids, bases, metal ions, and organic solvents (Vasapollo et al., 2011). These properties are likely contributions from their tendencies to have much higher cross-link densities than most other polymers. For adsorption processes, selectivity has been identified as the primary means toward effectiveness of adsorbent products (Chitraka, 2006, Saha, 2009). However, many current technologies lack the necessary selective removal at low concentrations. This is likely due to the fact that as concentrations of P decrease, the number of matrix components that can potentially interfere with removal increase (Ensing et al., 2002). Thus, MIPs' selective recognition of molecules at extremely low concentrations is the foundation of their potential use as adsorbents for P removal.

1.6.1. *Fundamentals*

Creating a MIP is based on the formation of a complex between an analyte (commonly referred to as the template) and a functional monomer. In the presence of a large excess of cross-linking agent, a three-dimensional polymer network will form around the complex (Ramstrom and Mosbach, 1999). Removal of these imprinted molecules leaves cavities with a size, shape, and chemical functionality complimentary to those of the template (Tse Sum Bui and Haupt, 2010). The development of this synthetic receptor allows for MIPs to recognize the analyte of interest in solution and selectively remove it (Vasapollo et al., 2011). In this work, the template will be used as reference to the molecule used during the imprinting procedure. The term 'analyte' will be used in reference to the molecule targeted for removal (i.e. PO_4^{3-}) from aqueous solution.

1.6.1.1.   Synthesis and Characteristics of MIPs

There are two primary techniques for synthesizing MIPs (**Scheme 3**). The selection

of the techniques depends on the nature of the pre-polymerization interactions between

the template and the monomer. These techniques are self-assembly and preorganization.

The self-assembly approach, which was utilized in this research, uses non-covalent

forces such as hydrogen bonds, Van der Waals forces, ion/hydrophobic interaction, and

metal coordinations (Vasapollo et al., 2011, Beltran et al., 2010, Tom et al., 2012, Wu,

2012). The geometry and recognition of the cavities in the polymer matrix for this

approach is determined by the size and shape of the anionic target molecule. The functional

groups from the monomers that are attached to the polymer network provide the driving

force to pull analytes into the imprinted cavities (Beltran et al., 2010). This non-covalent

method allows for easy removal of the template/analyte and reversible binding during

later use (Tom et al., 2012). The non-covalent self-assembly approach is the most

frequently utilized technique for the preparation of MIPs due to the simplicity of complex

formation and availability of functional monomers that can be used to interact with a wide

variety of templates. The drawback of the non-covalent self-assembly technique is that

tends to produce a lower binding affinity, kinetics, and capacity for the target analyte than

MIPs synthesized using the pre-organized covalent approach (Vasapollo et al., 2011, Wu,

2012). Additionally, the monomer-template complexes in non-covalent MIPs are unstable.

Thus, a heterogeneous distribution of the imprinted templates is found (Wu, 2012).
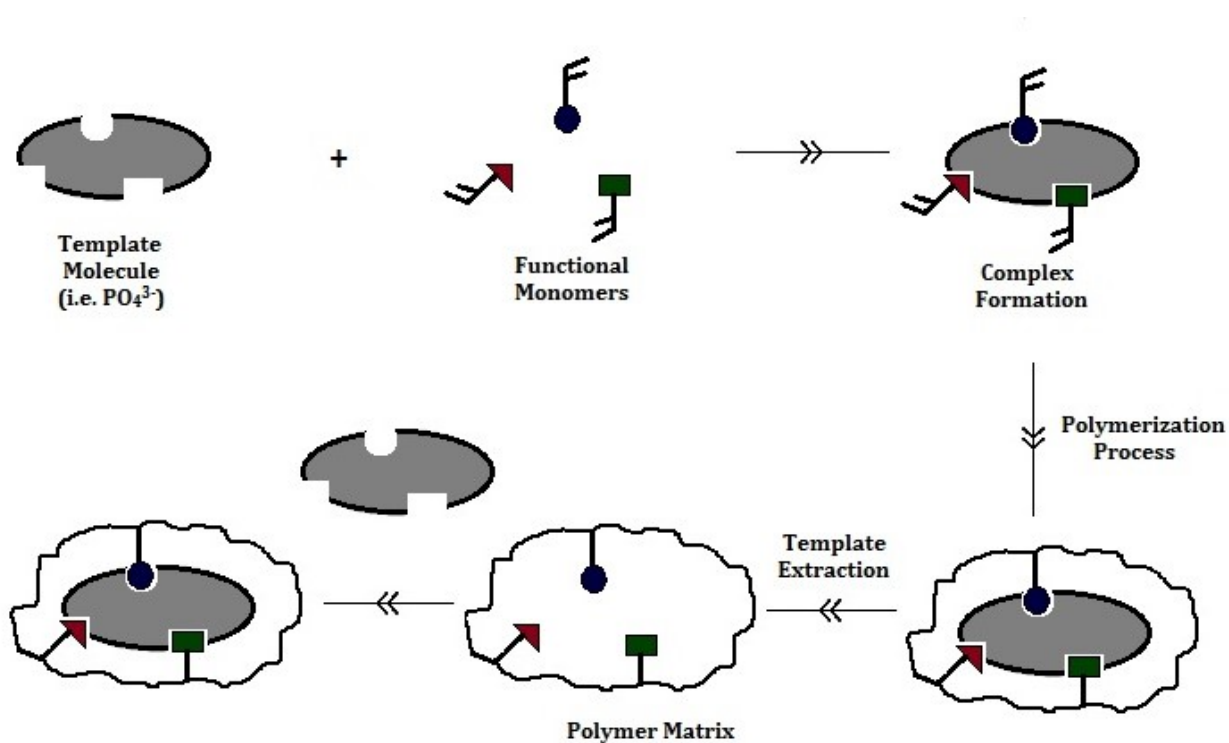
Scheme 3. Synthesis procedure for producing molecularly imprinted polymers (modified after (Wu, 2012))

Free radical polymerization is employed as the most popular synthetic method in the preparation of MIPs. Synthesis is typically conducted under mild conditions (at atmospheric pressure with temperatures lower than 80 °C). Free radical polymerization is typically very rapid and tends to have great compatibility for a wide range of functional monomers and templates (Vasapollo et al., 2012, Wu, 2012). The synthesis conditions can play a major role on MIP properties. It has been reported that non-covalent MIPs synthesized with the free radical polymerization method have shown better selectivity and higher binding capacity when synthesized under lower temperatures (Wu, 2012).

Typically, researchers polymerize MIPs in bulk and then grind the resulting polymer. The polymer particles are then sieved into desired ranges according to their intended application. This method is so widely used because it is simplistic. However,

grinding the particles is a tedious and time-consuming procedure which commonly results in particles of irregular shape and size (Vasapollo et al., 2011). Moreover, the grinding of these polymers results in the destruction of some recognition sites. The destruction of these recognition sites can lead to reduced analyte loading capacity (Vasapollo et al., 2011, Wu, 2012). Alternative synthesis methods can prepare different MIP formats such as beads, membranes, *in situ* prepared monoliths, surface imprinting, and monolayers (Poma et al., 2010).

After synthesizing and grinding the MIPs, extraction of the template needs to be carried out to generate the cavities for analyte recognition. Extraction is typically performed by exhaustive washing through rinsing the MIP continuously with a solvent. Soxhlet extractors are commonly employed to achieve this continuous rinsing over periods of 16 – 48 hours. Although, it is well known that even after exhaustive washing steps, much of the original template will still remain in the polymer matrix (Vasapollo et al., 2011). This inefficient template extraction is due, in large, to the high cross-link nature of the polymer. Many of the template molecules become trapped in the polymer matrix, making them unreachable for solvent extraction. Without extraction, these recognition sites remain inaccessible for rebinding with the target analyte. Additionally, the relaxation of polymer chains and subsequent swelling of MIPs by solvent can also lead to the inaccessibility of some recognition sites (Wu, 2012). Furthermore, not all of the recognition sites are selective. Some recognition sites in non-covalent MIPs are located outside of the cavities formed after extraction. These outside recognition sites tend to have non-specific binding (Rotello and Thayumanavan, 2008). This inefficiency in template extraction results in only

10 – 15% of the recognition sites being available for analyte rebinding (Sellegren and Shea, 1993).

1.6.1.2. Role of MIP Components

Some works have demonstrated that all components of the MIP (functional monomer, cross-linker, template, solvent, and initiator) play a significant role in affecting the template complexation (Karlsson et al., 2009, O'Mahony et al., 2007). The important role that each component specifically plays will be discussed further herein.

1.6.1.2.1. Functional Monomers

The functional monomer is the most important and influential component of the MIP. To synthesize a MIP with stable complexes and selective recognition capabilities, the monomer must first be found to have good interactions with the template (Vasapollo et al., 2011, Beltran et al., 2010). In non-covalent MIPs, monomer-template complexes are governed by equilibrium, and monomers are usually applied in excess (compared to the template) to favor the formation of these complexes. The monomer:template molar ratio to generate optimal complex formation has generally been seen as 4:1. However, Tom et al., (2012) showed that the imprinting factor (IF), a measure of the amount of successfully generated recognition sites, for MIPs increased as the monomer:template molar ratio was increased to 6:1. In spite of this trend, a significantly higher molar ratio of 15:1 was shown to essentially eliminate the imprinting effect. Consequently, this lopsided ratio has led to varying complex configurations and affinities. Also, it is seen as the reason for heterogeneous distribution of binding sites (Vasapollo et al., 2011). One option to increase the selectivity of MIPs is to synthesize the functional monomer used according to the

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functionalities present in the target molecule. Doing so may provide extra interaction

points between the monomer and template (Beltran et al., 2010).

<u>1.6.1.2.2. Cross-linking Agent</u>

In MIPs, the cross-linker also assumes important roles. The cross-linker is important

in controlling the morphology of the polymer matrix, stabilizing the recognition sites, and

imparting mechanical stability to the polymer. This allows the MIP to maintain its robust

nature and recognition capability (Sellergren, 1999). High cross-link ratios have generally

been used to generate macroporous materials with adequate mechanical stability.

Ethyleneglycol dimethacrylate (EGDMA) and trimethylolpropane trimethacrylate (TRIM)

tend to be among the most commonly utilized cross-linkers (Vasapollo et al., 2011).

Previous researches have distinguished that cross-linkers largely influence physical

characteristics of MIPs, but have little effect on the specific interactions between the

monomer and template (Molinelli et al., 2005, Navarro-Villoslada et al., 2004, Shi et al.,

2007). Contrarily, Tom et al. (2012) found that in the case of a sulfadimethoxine targeting

MIP, reducing the amount of cross-linker in the pre-polymerization mix improved the IF.

<u>1.6.1.2.3. Template Molecule</u>

The template used for MIP synthesis is typically dictated by the analyte molecule

being targeted for removal as they are generally one and the same. But, that is not always

the case. In some cases, the target analyte is not compatible with one or more of the other

components in the pre-polymerization mixture. Therefore, using the target analyte itself as

the template molecule is futile as it will not result in the development of monomer-

template complexes. Nonetheless, MIPs can still be synthesized by using a close structural

analogue of the analyte as the template. These templates are commonly referred to as

'dummy templates.' These dummy templates have a close enough resemblance to the target

analyte in shape, size, and chemical composition that it can generate very similar

recognition cavities. These comparable recognition sites are also able to capture the target

analyte, although it is not always as efficient (Maier et al., 2004, Rayane et al., 2008).

1.6.1.2.4. Solvent

The nature and volume of solvent also have a key role in the imprinting process. The

solvent serves to bring all of the components together into one phase. Additionally, it is

responsible for creating the pores throughout the polymer. Solvents commonly used for

synthesizing MIPs are toluene, chloroform, dichloromethane, or acetonitrile. To be a

practical solvent for MIP synthesis, the solvent must produce large pores to assure

adequate flow-through properties. Increasing the volume of the solvent used in pre-

polymerization mixes increases the pore volume throughout the polymer. Solvents are

commonly referred to as 'porogens' due to its influence on these properties (Vasapollo et

al., 2011). In non-covalent MIPs, the solvent can play a major role in monomer-template

interactions. Less polar solvents increase the complex formation, which facilitates polar

interactions between the monomer and template such as hydrogen bonding. More polar

solvents tend to dissociate the non-covalent interactions as they form hydrogen bonds with

template and/or receptors, prohibiting the formation of monomer-template complexes

(Vasapollo et al., 2011, Wu, 2012). However, it has been observed in several papers that

successful imprinted polymers have been developed in polar solvents. These solvents are

generally a combination of water and a less polar solvent like methanol or acetonitrile

(Scorrano and Vasapollo, 2010). It has also been found that the rebinding process of MIPs

functions optimally when the MIP is rebinding with target analytes in the same solvent that it was synthesized in (Vasapollo et al., 2011).

<u>1.6.1.2.5. Initiatiator</u>

The initiator does not play a very influential role in the polymerization process, but it is just as crucial a component as the rest. Small amounts of initiator (0.2 – 5 wt%) kick-starts the polymerization reaction (Wu, 2012). Azo-initiators, such as N-N'-bis isobutyronitrile (AIBN), are commonly used to perform this role. The reactions is typically very rapid and can be initiated by thermal or photochemical sources (Vasapollo et al., 2011). Photo-initiation at low temperatures decreases the kinetic energy of the pre-polymerization mixes, which has shown to increase complex stability and allow for better binding capacity and selectivity than thermal initiation. This is due, in large, to thermal initiation raising the kinetic energy of the pre-polymerization mix as it requires temperatures higher than 40 °C (Puoci et al., 2007, Athikomrattanakul et al., 2009, Spivak et al., 1997).

1.6.1.3.   Optimization of Components

Design of new MIPs requires a lot of time and effort for synthesis, washing, and testing. As can be seen from the discussion in Section 1.6.1.2, many of the components can generate different characteristics in MIPs after just being altered slightly. This requires many attempts to be made, by changing various parameters, to achieve optimum conditions (Vasapollo et al., 2011). In order to expedite this process, combinatorial chemistry can be applied (Lanza and Sellergren, 1999, Martin-Esteban and Tadeo, 2006, Dirion et al., 2003). Dirion et al. (2003) developed a high-throughput synthesis and screening (HTS) system in a 96-well plate format. This HTS system allows researchers to

optimize recognition properties of MIPs in a short amount of time by the use of filter plates

for rapid template removal and a multifunctional plater reader for parallel analysis of

supernatant fractions (Vasapollo et al., 2011).

1.6.1.4. Characterization of MIPs

To begin developing MIPs for removal of specific analytes, knowledge on the

formation of monomer-template complexes is often desired. UV-Vis, NMR, and theoretical

models are often employed to investigate the potential formation of these complexes

(Molinelli et al., 2005, Wei et al., 2007, Karlsson et al., 2009, Del Sole et al., 2009, Yañez et

al., 2010, Pietrzyk et al., 2009, O'Mahony et al., 2007). Job's method, titration curves, and

binding isotherms are methods currently being used to determine the nature of

interactions, the complex coordination number, and the association constant (Vasapollo et

al., 2011).

MIPs, themselves, are very difficult to characterize. Much of this difficulty arises

from their insoluble nature. To determine their chemical composition, analytical methods

such as solid-state NMR, elemental micro-analysis, and FTIR spectroscopy can be applied.

In order to obtain their morphological characteristics, light microscopy and scanning

electron microscopy (SEM) are commonly used (Del Sole et al., 2007, Cacho et al., 2004).

1.6.2. Anionic Imprinting

Cationic and neutral templates have been more extensively studied than the anionic

templates (Bianchi and Bowman-James, 1997, Beer and Gale, 2001). Anionic molecules are

larger in size than their cationic counterparts (Shannon, 1976), giving them a lower charge

to radius ratio. This smaller ratio has limited the usefulness of anionic imprinting as the

electrostatic interactions are far weaker than in cationic imprinting (Wu, 2012). Additionally, the application of anionic imprinted polymers has been strained due to the pH sensitivity of anions. At low pH, anions become protonated, resulting in a loss of their negative charge (Wu, 2012). A change in the charge of the analyte would make recognition by the MIP very difficult, or even impossible. Therefore, anionic imprinted polymers must be synthesized with the expected environmental pH range in mind. Generally the pH range cannot be large, therefore limiting the usefulness of anionic imprinted polymers. Synthesis of imprinted polymers with ionic moieties is termed ion-imprinted polymers (IIP). However, most works can be seen defining their polymers as MIPs due to the tendencies of many works to revert to molecular analogues. While these analogues may not show the same selectivity, they are generally easier to imprint.

1.6.2.1. Phosphate Imprinting

PO_4^{3-} is a tetrahedral anion which currently has not been studied extensively for imprinting technologies. Most examples involving phosphate are for organophosphorylated/organophosphonylated molecules, not phosphate itself (Wu, 2012, Wu et al., 2008). Furthermore, almost all of the work done on phosphate recognition with imprinting technologies has had sensor application in mind, not recoverable adsorbents.

Some of the complexities involved in developing MIPs with effective recognition of phosphate, which may explain a general want of available research, may arise from the physical and chemical nature of the molecule. Phosphate is a relatively large oxyanion. While this may cause some issues with the charge to radius ratio, as discussed before, there are more issues with selective recognition of PO_4^{3-} over other oxyanions similar in shape and size, such as sulfate (SO_4^{2-}) (Katayev, 2006). Additionally, phosphate is not soluble in

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non-polar organic solvents. This lack of solubility in organic solvents greatly limits the range of components that can be used to directly imprint phosphate. However, some works have managed around these traits to develop imprinted polymers for phosphate recognition.

(Özkütük et al., 2008) was one of few reported works to develop a phosphate IIP and look into its application as an adsorbent. To achieve this feat, Özkütük et al. (2008) first synthesized a monomer, Fe(III)-chitosan-succinate, which could dissolve in water while also maintaining groups that could develop a complex with the phosphate anion. After phosphate anions were mixed and allowed to form complexes with Fe(III), the metal-chelate polymer was then cross-linked with epichlorohydrin to form the polymer matrix. After removal of the template from the polymer matrix, this polymer showed a relatively low sorption capacity of approximately 11 mg P/g polymer. The non-imprinted polymer (NIP), acting as the control, had the same sorption capacity and characteristics as the IIP. This leads to the belief that the mechanism of removal was not from selective recognition sites, but more likely from the Fe(III)'s electronegativity. Preferential removal of phosphate over other anions was observed; this is likely attributed to phosphates stronger affinity to form inner-sphere complexes with transition metals than other anions. When rinsed with a high pH solution (KOH), the bound phosphate anions desorbed efficiently resulting in a sorption capacity decrease of only 1% after seven repeated adsorption-desorption cycles. Instead of investigating the potential to recover P by directly applying the metal-chelate polymer into fertilizer, this work was more concerned with the reusability of the polymer sorbent.

The Kugimiya group has been the most prominent group in the development of imprinted polymers for phosphate recognition. As opposed to the last discussed work, this group has been developing MIPs for recognition of phosphate with the use of structurally similar analogue molecules for templates. 1-Allyl-2-thiourea (AT) was the group's primary monomer studied for the selective removal of phosphate as it has been seen that thiourea groups have a strong binding affinity to phosphate (Nishizawa et al., 1999, Aoki et al., 2003). Their studies reported that when 40 mg of polymer was applied, the amount of phosphate bound to the imprinted polymer was reported as nearly 90% compared to the non-imprinted polymer which showed a phosphate binding of only 4% (at 25 µM PO_4^{3-}). However, when studying the interaction of the monomer with other anions, they also found strong interactions. While an explicit interference study with competing anions was not conducted, it can be assumed that interference would be present (Kugimiya and Takei, 2006, 2007, 2008, Kugimiya and Babe, 2011). The Kugimiya group also attempted to develop their own monomer for MIP phosphate recognition. In their work, the group also tried applying methacrylic acid and N-methyl-N'-(4-vinylphenyl)-thiourea as functional monomers for MIP phosphate recognition. However, both of these monomers showed little to no phosphate binding activity (Kugimiya and Takei, 2006). Additionally, one work investigated the effectiveness of using different structural analogues for phosphate templating (phenylphosphonic acid and diphenylphosphate). This work showed that phenylphosphonic acid imprinting resulted in better phosphate recognition (Kugimiya and Takei, 2006). This superior recognition is likely due to the fact that phenylphosphonic acid is more closely related to phosphate than is diphenyl phosphate. Phenylphosphonic acid maintains essentially the same structure as phosphate except with one oxygen atom

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replaced with a benzene ring. Diphenyl phosphate is the same way except it replaces two oxygen atoms with benzene rings.

Work done by Warwick et al. introduced some new potential monomers for MIP phosphate recognition with the goal of sensor development in mind. In this work, computational modeling was performed using the molecular modelling software program CentOS Linux-based hardware, SYBYL, version 7.3 (Tripos International, 2006). Over 30 functional monomers were screened using this computational modeling. Four monomers, allylamine, 2-(diethylamino)ethylmethacrylate (DEAEM), 2-methacryloyloxyethyltrimethylammonium chloride (METAC), and N-phenylethylene diamine methacrylamide (NPEDMA) were selected from the screening for further study. Warwick et al. selected thiourea as a control, citing its thorough study from the Kugimiya group. However, this appears to be a clear misunderstanding as the Kugimiya group thoroughly studied 1-Allyl-2-thiourea (AT), which simply contains a thiourea group. Therefore, the report from this study stating thiourea as a control for comparison from previous works is erroneous. Accordingly, the analytical comparison between the sorption capacities of the thiourea MIP and AT MIP, reported by Warwick et al., is a fallacy. Nevertheless, the data acquired on the reported polymers appears to be reliable and valid. Allylamine, DEAEM, and NPEDMA all had removal efficiencies that were nearly identical between their imprinted and non-imprinted versions. This showed that removal did not result from imprinting, but merely from electrostatic interactions of the polymers with phosphate. The imprinted and non-imprinted forms of METAC showed to each remove 100% of P from solution (0.78 mg P/L). Thus, it is unknown from this report as to whether imprinting occurred or not. Lastly, the thiourea imprinted polymer showed to bind 22% of

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the phosphate, while the non-imprinted polymer (NIP) only bound 1%. This was, however, not entirely convincing of thiourea's ability to be effectively imprinted as the thiourea MIP deviated largely from the mean (Warwick et al., 2014a).

A later work conducted by Warwick et al. examined the effectiveness of varying cross-linkers on the successful imprinting phosphate analogues and recognition of phosphate anions for sensor development. Commonly used cross-linkers diethyleneglycol diacrylate (DEGDA), ethylene glycol dimethacrylate (EGDMA), tetra(ethylene glycol) diacrylate (TTEGDA), N,N'-methylenebis(acrylamide) (Bisacrylamide), and trimethylolpropane trimethacrylate (TRIM) were compared. Of these cross-linking reagents, EGDMA was found to be the most effective cross-linker for developing phosphate recognition sites while bisacrylamide was a close second. This was described to be a likely result of the physical characteristics of EGDMA and phosphate. Phosphate and its structural analogues are relatively small templates for imprinting. Imprinting such a template will thus result in small cavities. Therefore, it was deemed that shortest cross-linkers, such as EGDMA and bisacrylamide, would likely be more appropriate to enhance the rigidity of the polymeric matrix and uphold the integrity of the recognition sites (Warwick et al., 2014b). It seems reasonable that maintaining more active recognition sites would lead to higher P removal, and the data backing this explanation appears to act as validation.

Some other studies have worked on imprinting phosphate containing molecules. While this is not directly impacting the area of research for phosphate sensing/removal with MIPs, it has provided some valuable information which may influence future MIP development. For instance, it was thought that the use of multifunctional monomers in MIPs may lead to some problems for selective removal of analytes. Using multifunctional

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monomers creates more interaction points around the monomer:template complex, which is good for selective recognition. However, having these multifunctionalities may also increase the number and the binding affinity of untemplated binding sites such as those formed in NIPs due to the preorganization of the functional groups. These background sites would typically have lower selectivity and would compete with more selective recognition sites, thus diminishing the magnitude of the imprinting effect and the overall selectivity of the polymer (Alexandratos, 2004, Filby, 2006, Weck, 2007). This potential problem for phosphate binding MIPs was studied by (Wu et al., 2008). In contrary to the potential problem at hand, Wu et al. (2008) showed that phosphate binding MIPs prepared with multifunctional monomers actually exhibited better binding capacity and selectivity than MIPs prepared with monofunctional monomers. While this work was focused on organophosphates, it still provides evidence to suggest that MIPs developed for specifically targeting orthophosphates will exhibit increased efficiency with multifunctional monomers.

1.7. Research Objectives

With society trending toward more sustainable methods to utilize crucial resources, more advanced technologies are needed to achieve these goals. For P specifically, new sources need to be exploited to reduce the pressure on natural and finite sources of phosphate rock. Surface waters, which serve as a typical landing point for P from wastewater effluent or that which is not uptaken by agricultural plants, serve to act as a reasonable alternative. In tandem to generating new recoverable P sources from surface waters, the health of economically significant lakes could be restored; thus, saving billions for the United States and other countries annually. Currently established technologies do

not contain all of the essential characteristics needed for enlisting a completely effective P adsorbent. Technologies most commonly lack one or more of the following characteristics: (1) effective removal at environmentally relevant concentrations of 100 μg P/L or lower, (2) selective removal of P amongst coexisting constituents, (3) cost-efficiency, and (4) recoverability of a useful P product for reuse in agriculture. Molecularly imprinted polymers (MIPs) serve as a potential vehicle for meeting all of these desired traits. MIPs developed for phosphate recognition (sensors) have shown to work for concentrations below 100 μg P/L. These polymers are also known for high analyte selectivity due to their specific recognition sites. Additionally, MIPs are generally biodegradable polymers. While they can typically be biodegraded, their robust physical nature is expected to maintain their physical structure and recognition site integrity while in the field. This study focused on three MIPs found in literature that were previously studied only for P sensing. The selection of these MIPs was based on the presence of potential imprinting effect. The three MIPs were then assessed to see if MIT could be a feasible technology for P removal and recovery. Sorption capacities were studied to determine whether or not these MIPs could be competitive when compared to currently available technologies. Several experiments were conducted to understand the selective nature of the MIPs and the mechanisms of P release/removal. The goal was to understand the MIPs' behavior under typical environment conditions so that predictions could be made as to how these polymers function as sorbents and how to overcome potential limitations in the future.

CHAPTER 2. ASSESSMENT OF MOLECULARLY IMPRINTED POLYMERS AS SUSTAINABLE PHOSPHATE SORBENTS

2.1. Introduction

The role phosphorus (P) plays as an essential macronutrient in plants is extremely important. P is also extremely important for microorganisms. However, the supply of mineral P is finite and it is predicted that deposits of this mineral will only last for another 60 – 240 years (Cornel and Schaum, 2009, USGS, 2012). The vast majority of mined phosphate rock around the globe is used in agricultural products such as fertilizers to increase crop production and meet food demands of the global population (Desmidt et al., 2014, Cordell et al., 2009). As the global population continues to grow, demand for P-containing fertilizer will increase to maintain sufficient crop yields and sustain global food security (UN, 2000, 2005). Along with an increasing demand for P, the cost of mining phosphate rock is increasing due to the decline in the quality of deposits (Desmidt et al., 2014).

While being innately important in agriculture and global food sustainability, P has more complications than just being a nonrenewable resource; excessive loading of P into surface waters can promote eutrophication of lakes, rivers, reservoirs, estuaries, and parts of the oceans (USEPA, 1983, 1998). P is the key growth-limiting nutrient for algae in the eutrophication process, which leads to an overall degradation of precious water resources (Smith, 2003, Dodds et al., 2009). Eutrophication is not only extremely detrimental to aquatic ecosystems, but also to the economic well-being of communities. Lakes and rivers provide drinking water, recreation, and aesthetic benefits, and all of these can be negatively

influenced by eutrophication (Beeton, 2002). Combined costs of freshwater eutrophication due to these effects were estimated at approximately $2.2 billion annually in the United States (Smith, 2003). Recovering P from surface water for its reuse in agricultural applications has been identified as a means to establish renewable P sources while also restoring surface waters to acceptable (non-eutrophic, <50 μg/L) P limits (Rittmann et al., 2011, Cordell et al., 2009, 2011).

Methods such as sediment dredging, hypolimnetic withdrawal, hypolimnetic aeration, and P inactivation are a few techniques currently used for P remediation from surface water. Dredging and hypolimnetic withdrawal/aeration techniques typically only provide temporary relief and can cause excessive disturbance to lake ecosystems. Additionally, they are often cost inefficient. P inactivation is a commonly applied chemical restoration method. Alum is one of the most commonly used materials for this application, but it is ineffective over a wide pH range and thus needing pH adjustments; furthermore, it can release toxic materials after long exposure to hydrolysis (Goldyn et al., 2014, Lewtas et al., 2016, Singh, 1982). The technologies that are the most well-established and utilized for P removal from aqueous media are chemical precipitation and biological remediation (Rittmann et al., 2011, Jenkins and Hermanowicz, 1991, Stensel, 1991, Pan et al., 2009). These technologies are chiefly designated for removal of P from wastewater but are technically challenging and not cost effective to meet the increasingly stringent P discharge goals. These techniques are also inadequate at reducing P concentration when present in environmentally significant levels of 100 μg/L or less (Pan et al., 2009, Chouyyok et al., 2010, You et al., 2016). Other technologies which have been developed for P removal from aqueous media include waste by-products (i.e. cationized wood residues (Unnithan et al.,

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2002, Tshabalala et al., 2004, Karthikeyan et al., 2004), iron oxide tailings (Zeng et al., 2004), alum and ferric sludges (Babatunde and Zhao, 2010, Song et al., 2011), alkaline fly ash (Cheung and Venkitachalam, 2000), egg-shell waste (Yeddou Mezenner and Bensmaili, 2009), red mud (Huang et al., 2008), blast furnace slag (Oguz, 2004), and waste lime (Siobhan Dunets and Zheng, 2014, Ahn and Speece, 2010)), ion and ligand exchangers (Blaney et al., 2007, Sarkar et al., 2011, Sengupta and Pandit, 2011, Choi et al., 2012), membranes (Park et al., 2017, Luo et al., 2016, Dolar et al., 2011), and adsorbents (i.e. hydrated ferric oxides (Pan et al., 2009, You et al., 2015, You et al., 2016), magnetic-based adsorbents (Janos et al., 2013, Lai et al., 2016, de Vicente et al., 2011), layered double hydroxides (Das et al., 2006, Cheng et al., 2009), synthesized minerals (Peleka and Deliyanni, 2009, Chitrakar et al., 2006), modified clays (Copetti et al., 2016, Reitzel et al., 2013), modified biochars (Jung and Ahn, 2016, Li et al., 2016), immobilized transition metals (Mahaninia and Wilson, 2015, An et al., 2014, Ogata et al., 2011, Liu and Zhang, 2015), polymers (Dimitri et al., 2005, Mahaninia and Wilson, 2016, Hammud et al., 2015), and nanoparticles (Zheng et al., 2016, Chouyyok et al., 2010, You et al., 2016, Almeelbi and Bezbaruah, 2012, Lai et al., 2016)). Many of these technologies have the potential for being viable options for P removal; however, none of these technologies currently succeed in meeting the desired criteria for an optimal P removal which include: (1) P selectivity, (2) high P removal capacity, (3) environmental stability and harmlessness, (4) effective removal over a wide pH range, (5) low cost, (6) fast removal rate, (7) removal from concentrations below 100 µg/L, and (8) recoverability and reusability.

Table 1. Summary of P Management for Eutrophic Lakes

Phosphorus Management Method	Technology Advantages	Technology Disadvantages
Dredging	Remove nutrient-rich soil, deepen lake, immediately improve water quality	High cost, ecological risks
Hypolimnetic Aeration	Limit P release sediments, no resuspension	Ineffective control of P
Hypolimnetic Withdrawal	Reduces nutrients, eliminates low oxygen water	Detrimental to downstream waters
Nutrient Inactivation	Removes P and algae from water column, acts as barrier on sediment	Ecological risks
Dilution	Flushes out algae and high nutrient water	High cost, inefficient
In-situ Sediment Capping	Highly effective and environmentally friendly	Site dependent
Thermally Treated Attapulgite	Transforms mobile P into inert P	Energy intensive

Table 2. Summary of Available P Removal Technologies

Phosphorus Removal Technology	Removal Capacity	Recovery Value	Technology Advantages	Technology Disadvantages
Chemical Precipitation	(48 - 230 mg P/g precipitant)	Low	Well-established technology	Low recyclability, can't remove P <100 µg/L
Biological Remediation	(50 - 300 mg P/g PAOs)	Moderate	Well-established technology, no chemical usage	Highly variable P removal, extensive sludge handling, difficulty removing P <100 µg/L
Waste By-product (WBP) Removal	(0.6 - 26 mg/g WBP)	Low	Low-cost, efficient use of renewable waste	Low selectivity, low reusability
Ion and Ligand Exchange	(2.5 - 26 mg/g resin)	High	High selectivity, high P recoverability, high exchanger regenerability	High-cost
Filtration/Membrane Rejection	(65 - 98% rejection)	None	Easy to use, small physical footprint	High-cost, no useful product
Adsorption	(6 - 200 mg/g adsorbent)	Low	Effective at low concentrations	Regeneration issues, low selectivity

Molecularly imprinted polymers (MIPs) are synthetic recognition materials capable of being tailored to selectively recognize a wide range of molecules. Some molecules that have been thoroughly studied for recognition with this technology are proteins, nucleotide derivatives, pollutants, drugs, and food. This has led to MIPs having promising applications in separation sciences and purification (Vasapollo et al., 2011, Khorami and Edrisi, 2010, Amut et al., 2010, Lee et al., 2010), solid phase extraction (Vasapollo et al., 2011, Bravo et al., 2007, Al-Degs et al., 2009, Alexiadou et al., 2008, Tse Sum Bui and Haupt, 2010, Ensing et al., 2002), chemical sensors (Vasapollo et al., 2011, Greene and Shimizu, 2005, Lakshmi

et al., 2009), catalysis (Vasapollo et al., 2011), drug delivery (Vasapollo et al., 2011, Alvarez-

Lorenzo and Concheiro, 2004), biological antibodies (Vasapollo et al., 2011), and as

adsorbents (Lee and Doong, 2012). The main advantages of MIPs are their (1) high

selectivity and affinity for the target molecule, (2) excellent environmental stability, (3) low

cost, and (4) relative ease to design, synthesize, and modify (Vasapollo et al., 2011, Wu,

2012). Selectivity has been identified as the primary means toward commercial

effectiveness of adsorbent products (Chitraka, 2006, Saha, 2009). The demonstrated nature

of MIPs to selectively recognize molecules at extremely low concentrations is the

foundation of their potential use as an adsorbent for P removal.

Phosphate (PO_4^{3-}) has been studied in a very limited fashion for imprinting

technologies. Most examples involving phosphate are for

organophosphorylated/organophosphonylated molecules, not PO_4^{3-} itself (Wu, 2012, Wu et

al., 2008). Furthermore, among the limited work done on phosphate recognition with

imprinting technologies, almost all had sensing applications in mind, not phosphate

adsorption. Work done by Ozkutuk et al. (2008) was one of the only to produce an

ionically-imprinted polymer (IIP) for phosphate adsorption with reusability in mind. While

this product exhibited some sorption capacity, it was shown to have no actual selective

removal of phosphate. Preferential removal of phosphate over other anions was observed,

though this is likely attributed to phosphate's stronger affinity to form inner-sphere

complexes with transition metals than other anions. The Kugimiya group has been the most

prominent group in the development of imprinted polymers for phosphate recognition. As

opposed to the last discussed work, this group has developed MIPs for recognition of

phosphate with the use of structurally similar analogue molecules for templates. 1-Allyl-2-

thiourea was the group's primary monomer studied for the selective sensing of phosphate due to thiourea groups having a strong binding affinity to phosphate (Nishizawa et al., 1999, Aoki et al., 2003, Kugimiya and Takei, 2006, 2007, 2008, Kugimiya and Babe, 2011). Work done by Warwick et al. introduced other monomers for MIP phosphate recognition with the use of computational modeling, some of which showed promising P removal traits (Warwick et al., 2014).

As mentioned before, almost the entirety of the work completed for P recognition by MIPs has been aimed toward the development of sensor technology. MIPs show potential to be a viable solution to solving the critical P problem, yet none of the work on MIPs has looked into their feasibility as sorbents.

In this work, a survey of all MIPs developed for phosphate removal was conducted. Three monomers which showed the greatest traits that could potentially lead to them being viable MIP sorbents were selected for further study. The three monomers selected were (1) 1-Allyl-2-thiourea (AT), (2) thiourea (TU), and (3) 2-methacryloyloxyethyltrimethylammonium chloride (METAC). The feasibility of the resultant MIPs from these monomers for sorbent applications was assessed by studying their sorption capacities, resistance to interference by coexisting anions, and their potential for P recovery. The influence of some design parameters were studied to analyze how MIPs can be optimized for sorption applications. Additionally, the mechanism of phosphate removal was studied to gain insight into how the MIPs function, where some limitations have originated from, and how to potentially overcome those limitations in the future.

2.2.   Materials and Methods

2.2.1. Chemicals

The chemicals used in this study include: potassium dihydrogen phosphate

(KH_2PO_4, 99% pure, EMD), 1-Allyl-2-thiourea (>98%, TCI America), thiourea (99%, Alfa

Aesar), 2-methacryloyloxyethyltrimethylammonium chloride (80 wt. % in H_2O, Sigma

Aldrich), ethylene glycol dimethacrylate (EGDMA) (98%, stabilized with 100ppm 4-

methoxyphenol, Alfa Aesar), phenylphosphonic acid (≥98%, TCI America), acetonitrile

(ACS Grade, BDH), and 2,2'-Azobis(2-methylpropionitrile) (AIBN) (98%, Sigma Aldrich),

methanol (ACS Grade, BDH), potassium nitrate (KNO_3, 99% pure, Alfa Aesar), potassium

hydroxide (KOH, ACS Grade, BDH), potassium sulfate (K_2SO_4, ACS Grade, HACH), potassium

chloride (KCl, ACS Grade, BDH), potassium bicarbonate ($KHCO_3$, ACS Grade, Alfa Aesar),

natural organic matter (Suwannee River NOM, RO isolation, IHSS), humic acid (H1452,

Spectrum), sulfuric acid (Solution 6.0 N, BDH), antimony potassium tartrate (≥99%,

Spectrum), ammonium molybdate tetrahydrate (ACS Grade, BDH), ascorbic acid (≥98%,

Alfa Aesar), NitraVer X Reagent Set (Test 'N Tube, Hach), hydrochloric acid (Solution 6.0 N,

BDH), ethanol (Denatured, 95%, BDH), glycerol (Laboratory Reagent Grade, BDH), barium

chloride dihydrate (ACS Grade, BDH), and sodium carbonate (Anhydrous, ACS Grade, BDH)

. All the chemicals were used as received unless otherwise specified. The solutions were

prepared with deionized (DI) water, which was previously passed through E-pure system

(Series 1090, Barnstead).

2.2.2. MIP Synthesis

MIP synthesis requires chemical reactions between the monomer, template, and cross-linker during polymerization (**Schemes 4 – 6**). The overall polymerization process involved the combination of reagents (functional monomer, cross-linker, solvent, and template) following the addition of the initiator and nitrogen flushing to remove oxygen, which is a reaction inhibitor. After flushing, the combined reagent mixture was then polymerized by free radical polymerization in a water bath at 65 °C for 24 hours. The phosphate anion analogue, phenylphosphonic acid, was selected as a template to overcome solubility issues associated with phosphate anions in organic media. In addition to preparing MIPs, negative controls were prepared, non-imprinted polymers (NIPs), which followed the same procedure but omitted the phosphate template. After the polymerization process was complete, the polymers were blended and then purified using Soxhlet extraction with methanol overnight and then dried at 30 °C to remove remaining methanol from the surface (**Scheme 7**).

Table 3. Combination of Reagents for Synthesis of MIPs

Name	Monomer Qty.	Cross-linker[1]	Solvent[2]	Template
METAC	830 mg (4 mmol)	3.8 mL (20 mmol)	5 mL	158 mg (1 mmol)
TU	305 mg (4 mmol)	3.8 mL (20 mmol)	6 mL	158 mg (1 mmol)
AT	465 mg (4 mmol)	3.8 mL (20 mmol)	6 mL	158 mg (1 mmol)

[1]All polymers used ethylene glycol dimethacrylate (EGDMA) as the cross-linker. [2]Acetonitrile was used as the solvent for all polymers, however varying amounts were used to meet solubility requirements. <u>Note</u>: 0.02% wt. of 2,2'-Azobis(2-methylpropionitrile) (AIBN) was used as the initiator for each polymer.
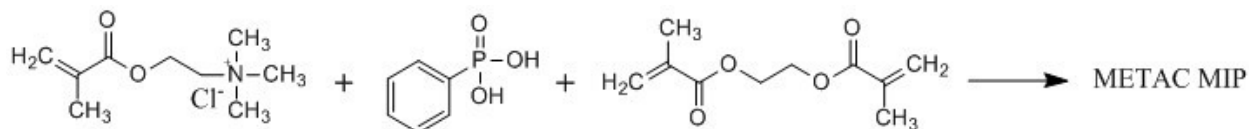
Scheme 4. Chemical Reaction Scheme of METAC MIP Synthesis
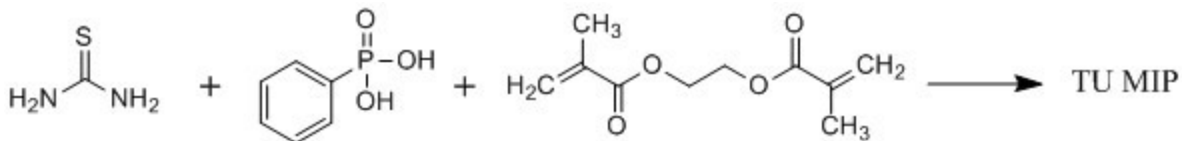
Scheme 5. Chemical Reaction Scheme of TU MIP Synthesis
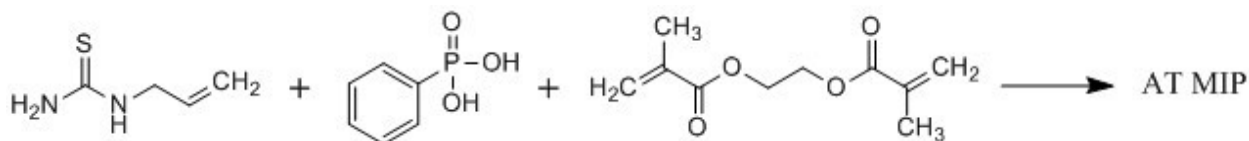
Scheme 6. Chemical Reaction Scheme of AT MIP Synthesis
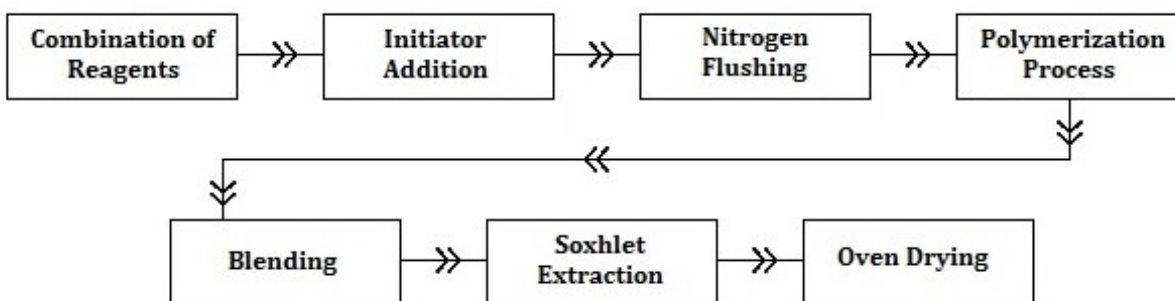
Scheme 7. General MIP Synthesis Procedure

2.2.3. Point of Zero Charge and pH Study

The point of zero charge (PZC) of a material is defined as the pH value at which the net charge on the solid surface is zero. PZC for the MIPs was determined by following the potentiometric mass titrations technique (Vakros et al., 2002). In brief, three solutions of equivalent potassium nitrate were created, but with varying amounts of MIP (0.5 g/100 mL, 1.0 g/100 mL, and 1.5 g/100 mL). The polymer was then left in the KNO_3 solution for 24 hours (in a sealed nitrogen environment) to allow the surface complexes to be saturated with nitrate. After the 24-hour waiting period, the pH of each solution was adjusted with KOH to approximately 11.5 – 12. After reaching this pH range, the samples were then titrated with HCl to a pH value of approximately 3 or lower. pH was measured continuously

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with each volume addition of HCl. The volume of titrant is then plotted against the pH-value

for all three concentrations of the MIP. The point that all three samples intersect is the pH

at which the PZC is reached (Vakros et al., 2002).

The effect of pH on phosphate removal by MIPs was investigated at pH values of 3, 4,

5, 6, 7, 8, 9, 10, 11, and 12. To reach these pH levels, the pH was adjusted by the addition of

1M HCl and 1M KOH to the phosphate solution (5 mg PO_4^{3-}-P/L).

2.2.4. Isotherm Studies

Batch experiments were conducted to understand the isotherm behavior of the

MIPs during phosphate removal. Each batch reactor (50 mL) contained 1 g of MIP and 50

mL of phosphate solution. The polypropylene plastic test reactors and controls (no MIP,

only phosphate solution) were then rotated at room temperature (22 ± 2 °C) at 28 rpm in a

tailor-made end-over-end shaker to reduce mass transfer resistance. All samples were run

in triplicate. Reactors were withdrawn after 24 hours and the used MIPs were separated

from the reactors by centrifugation. The phosphate concentration in the bulk solution was

measured to calculate the sorption capacity of the MIP. Initial concentration of phosphate

was varied from 0.1 – 3500 mg PO_4^{3-}-P/L. The data were fit to Langmuir and Freundlich

isotherm models. The Langmuir isotherm assumes monolayer sorption and that adsorption

only occurs at a finite number of localized sites. Further, the Langmuir isotherm refers to

homogenous sorption, describing all sites as possessing equal affinity for the sorbate (**Eq.**

1).

$$q = \frac{K_L q_m C_e}{1 + K_L C_e} \qquad (1)$$

Where, q_m is the maximum loading of the sorbent (mg PO_4^{3-}-P/g sorbent) and K_L is the Langmuir sorption constant (L/mg sorbent).

The Freundlich isotherm describes heterogeneous and reversible sorption not restricted to the formation of a monolayer (**Eq. 2**).

$$q = KC_e^{\frac{1}{n}}$$ (2)

Where, K is the sorption Fruendlich constant (mg PO_4^{3-}-P/g sorbent)(mg/L)$^{-1/n}$, and n is the Fruendlich exponent. Values of n > 1 represent favorable sorption.

2.2.5. Temperature Studies

In order to assess the behavior of MIPs removal under various temperature conditions, a temperature study was conducted. A range of relevant temperatures that MIPs may experience when in the environment (4, 10, 15, 20, 25, 30, and 35 °C) were selected. Phosphate solution was first placed in an environmental chamber for 2 hours to reach the desired temperature before MIPs were introduced into the reactors. The same batch study procedure as discussed previously was used. The phosphate removal efficiency (η) was calculated using (**Eq. 3**).

$$\eta = \frac{C_0 - C_e}{C_0}$$ (3)

2.2.6. Kinetic Studies

Kinetic studies were conducted to delve into the mechanism of phosphate removal. This testing was also conducted as a batch study, with the same procedure as previously described for the isotherm testing. Three concentrations (0.1, 5, and 50 mg PO_4^{3-}-P/L) were chosen for the kinetic studies. The 0.1 mg PO_4^{3-}-P/L concentration was selected to gain insight into the kinetics of removal at environmentally relevant P concentrations (100

μg/L) found in eutrophic lakes. Sets of three sacrificial reactors were withdrawn at specific time intervals (4, 6, 8, 10, 15, 30, 60, 90, 120, 180, 360, 720, 1080, and 1440 min) to have the phosphate concentrations in the bulk solutions measured. The phosphate removal efficiency (η) was calculated using (**Eq. 3**).

2.2.7. Interference and Partial-selectivity Studies

2.2.7.1. Interference Studies

Effects of possible competing ions and compounds on phosphate sorption by MIPs were investigated by adding common coexisting anions and natural organic matter (NOM) to the phosphate solution. Interference studied were carried out with chloride (Cl^-, 50 – 500 mg/L), bicarbonate (HCO_3^-, 10 – 100 mg/L), sulfate (SO_4^{2-}, 50 – 1000 mg/L), nitrate (NO_3^-, 10 – 100 mg/L), humic acid (2 – 10 mg/L), and Suwanee River NOM (10 – 50 mg/L) mixed with phosphate solution (5 mg PO_4^{3-}-P/L). These tests were conducted as batch studies, with the same procedure as previously described for the isotherm studies.

2.2.7.2. Partial-Selectivity Studies

Further studies were needed to assess whether or not the recognition sites of the MIP showed any sort of selectivity. To evaluate this, 1 g of the MIP was saturated with excessive amounts of a competing anion (110 meq/L NO_3^-, Cl^-, and SO_4^{2-}). The batch reactors were rotated at 28 rpm for 24 hours to allow sufficient time for all non-selective binding sites to become saturated and electrostatic interactions to be completed. After saturation, the saturated MIP was filtered using a 0.45 μm pore size cellulose nitrate membrane filter (Whatman, 47-mm-diameter). The remaining bulk anion solution was tested to examine non-phosphate anion removal, which in turn was used to estimate the

amount of anions adsorbed onto the MIP. The saturated MIP was then placed in 50 mL

polypropylene tubes with equal parts (110 meq/L) solution of phosphate and other

competing anions (nitrate, sulfate, chloride). The samples were then run in same manner as

described in the isotherm study. The bulk solution of each sample was tested for phosphate

and for the competing anion.

2.2.8. Lake Water Study

Eutrophic lake water samples were collected in April 2017 from the following lakes

located in Minnesota, USA: Lake Katrina (45.012913°N, -93.623659°W), Holy Name Lake

(45.014284°N, -93.532872°W), Lake Ardmore (45.032101°N, -93.636703°W), Lake

Independence (45.026063°N, -93.636109°W) , and Half Moon Lake (45.028521°N, -

93.628103°W). The lake water samples were filtered using a 0.45 μm pore size cellulose

nitrate membrane filter (Whatman, 47-mm-diameter) and stored in glass bottles at 4 °C for

further analyses. This testing was also conducted as a batch study, with the same

procedure as previously described for the isotherm studies.

2.2.9. Biodegradability Studies

To assess the potential for the MIP to be broken down biologically, a biodegradation

study was conducted. Biodegradability of a material can be determined by indirect

measure of the parent compound's bioconversion (i.e. biochemical oxygen demand (BOD))

by monitoring the BOD over time using a respirometer. The experiment followed the Hach

Standard Method (Hach, 1995). BOD was monitored using an automated closed-system

respirometer (BODTrak Apparatus, Hach, USA). BODTrak correlates the quantity of oxygen

consumed to the change in headspace pressure. As the organic matter in the sample is

biodegraded, the headspace pressure will decrease. BOD data were automatically collected by the BODTrak via a data-logger and stored on a computer connected to the instrument during the 30-day test period. Mixed liquor suspended solids (MLSS) was collected from the City of Moorhead Wastewater Treatment Plant (Moorhead, MN) and used immediately to preserve the integrity of the microorganisms. The MLSS served as the seed for the bacteria population. The biodegradation studies were conducted in 500 mL amber bottles and ran in triplicate. BOD nutrient solution was prepared by mixing a BOD nutrient buffer pillow (Hach, USA) with 6L of DI water which had previously been oxygenated for 24 hours. Next, each reactor was filled with MIP (100 mg), BOD nutrient (155 mL), and MLSS (5 mL). Upon filling each reactor, a seal cup was placed on the reactor and LiOH was added to adsorb generated CO_2. Special care was taken to ensure the LiOH did not spill into the solution. Recorded BOD values were corrected for dilution and seeding factors.

2.2.10. Desorption Studies

A desorption study was conducted to evaluate the release pattern of sorbed phosphate from the spent MIP. For this, 1 g of MIP was placed in polypropylene tubes filled with 50 mL phosphate solution (2000 mg PO_4^{3-}-P/L) and allowed to react for 24 hours at 28 rpm in the end-over-end shaker. The spent MIP was then filtered using a 0.45 μm pore size cellulose nitrate membrane filter (Whatman, 47-mm-diameter). The bulk phosphate concentration was measured in the samples. After separation from the solution, the spent MIP was placed in a 50 mL polypropylene tube filled with DI water. The reactors were then run for a 4-day period with desorption data collected at different cycle times (0.25, 0.5, 1, 2, 3, and 4 d). After each cycle had completed, the reactors were taken out from the shaker. The MIP was filtered out from solution and placed in 50 mL reactors with fresh DI water.

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The bulk water after each cycle was analyzed for P content to determine how much P was released by the MIP over those periods.

2.2.11. Phosphate, Nitrate, Sulfate, and Chloride Analysis

To quantify the P content in supernatants, the Ascorbic Acid Method was employed (Eaton et al., 2005). Nitrate-nitrogen and total chlorine concentrations were each determined with the use of Hach Test 'N Tube reagent kits. The concentration of sulfate present was measured using a turbidimetric method (Method #375.4) developed by the United States Environmental Protection Agency (USEPA) (USEPA, 1978).

2.2.12. Material Characterization

2.2.12.1. SEM/EDS

MIP particles were characterized using scanning electron microscopy with energy dispersive spectroscopy (SEM/EDS) to analyze the surface morphology of the particles and characterize the elemental composition of the polymer. Fresh (before phosphate removal) and spent (after phosphate removal) MIP samples were used for imaging and EDS analyses.

For high-resolution SEM/EDS, samples were attached to aluminum mounts using silver paint (SPI Products, West Chester, Pennsylvania, USA) or double-stick carbon tape (Ted Pella, Redding, California, USA), and then coated with a conductive layer of carbon in a high-vacuum evaporative coater (Cressington 208c, Ted Pella Inc., Redding, California). Images were obtained with a JEOL JSM-7600F scanning electron microscope (JEOL USA, Inc., Peabody, Massachusetts.)   Energy-dispersive spectroscopy information was acquired using an UltraDry silicon drift X-ray detector and NSS-212e NORAN System 7 X-ray Microanalysis System (Thermo Fisher Scientific, Madison, Wisconsin.)

For low-resolution SEM/EDS, samples were attached to aluminum mounts with silver paint (SPI Products, West Chester, Pennsylvania, USA) or double-stick carbon tape (Ted Pella, Redding, California, USA), and then sputter coated (Cressington 108 auto, Ted Pella, Redding, California, USA) with a conductive layer of gold. Images were obtained with a JEOL JSM-6490LV scanning electron microscope (JEOL USA, Inc., Peabody MA, USA); energy-dispersive X-ray information was collected using a Nanotrace EDS detector with a NORVAR light-element window and Noran System Six imaging system (ThermoFisher Scientific, Madison WI, USA) at an accelerating voltage of 15 kV.

2.2.12.2. FTIR

Fourier transform infrared spectroscopic (FTIR) spectra were obtained ex-situ on fresh and spent MIPs using a Nicolet 8700 FTIR Spectrometer operated with OMNIC software. All spectra were obtained in the range of 4000 – 400 cm^{-1} using potassium bromide (KBr) as a background. Prior to analysis, samples were dried in a vacuum oven at 30 °C under nitrogen gas for 48 hours. Pellets were formed by crushing the dried samples with KBr (sample:KBr mass ratio of 1:10). Spectra were recorded at a resolution of 4 cm^{-1} with each spectrum corresponding to 64 scans. The background collected from KBr was subtracted from the sample spectra. Spectra information collected was plotted in the same scale on the wavenumber axis (x-axis).

2.2.12.3. Swelling Studies, and Prediction of Porosity and Hydrated Diameter

Swelling studies were conducting using dry MM12 particles. The term dry refers to the state of the particles after extraction and vacuum drying of excess methanol. Swelling studies were carried out in DI water. Accurately weighed amounts of MM12 particles

(ranging from 1 to 2 g) were immersed in 25 mL of DI water for 30 minutes. After immersion, the MM12 particles were filtered with a fiber-glass filter (Whatman, 47-mm-diameter). Immediately, they were weighed. The weight change of the beads was calculated according to the formula:

$$\% \ weight \ change = \frac{W_s - W_i}{W_i} * 100\% \tag{4}$$

where W_s is the weight of the MM12 particles in the swollen state and W_i is the initial weight of the MM12 particles.

Upon MM12 immersion in DI water, the average hydrated diameter (D), and the average porosity (ε) of swollen MM12 particles were estimated using **Eqs. 5&6**. The hydrated diameter is based on the assumption that the shape of the particle can be approximated as a sphere.

$$D = \left[6 * \frac{\left(W_i / \rho_{MM12}\right) + \left(\left(W_s - W_i\right) / \rho_w\right)}{\pi}\right]^{1/3} \tag{5}$$

$$\varepsilon = \frac{(W_s - W_i)/\rho_w}{W_i / \rho_{MM12} + (W_s - W_i)/\rho_w} * 100\% \tag{6}$$

where ρ_w is the density of water (1.0 g/cm^3) and ρ_{MM12} is the estimated density of MM12 (1.063 g/cm^3).

2.2.13. Statistical Analysis

The results are presented as the mean ± SD (standard deviation, n = 3). The data were checked for homogeneity of variance. One-way analysis of variance (ANOVA) was performed using General Linear Model, followed by a Tukey's pairwise comparison where appropriate. All statistical analysis was performed on Minitab version 17. Significance was determined based on whether *p*-values were less than 0.05 or not.

2.3.   Results and Discussion

2.3.1. *PZC and Effect of pH*

The PZC for the polymers gives an idea as to which pH the MIPs will be positively or negatively charged on the surface. Determining these values will help understand when the MIPs may remove coexisting anions along with phosphate due to non-selective electrostatic attractions under varying pH regimes.

The PZCs for the TU, AT, and METAC MIPs were obtained by using the potentiometric mass titration method and found to be 2.9 for TU (**Figure 1**), 8.9 for AT(**Figure 2**), and 10.9 for METAC (**Figure 3**). The TU MIP not reaching its PZC until an extremely low pH indicates that it has strong negatively charged surface sites above that pH. Therefore, non-selective electrostatic interactions with other anions are not anticipated. The AT and METAC MIPs were found to have PZCs above the typical pH range for eutrophic lakes (7.5 - 8.5, Michaud, 1991). This will provide active sites for electrostatic interaction and removal of phosphate, although it will also attract coexisting anions.
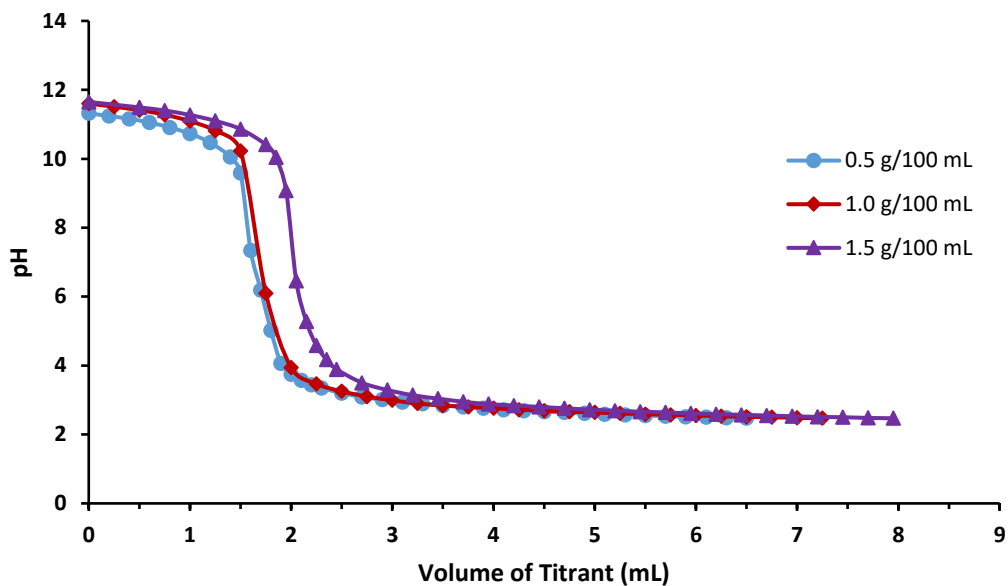
Figure 1. PZC of TU MIP. The PZC was found to be 2.9.
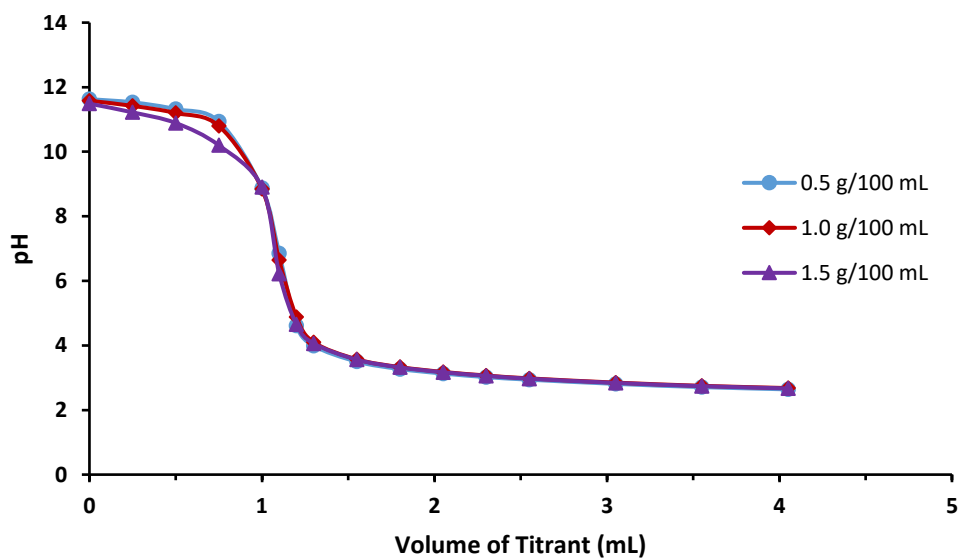
Figure 2. PZC of AT MIP. The PZC was found to be 8.9.

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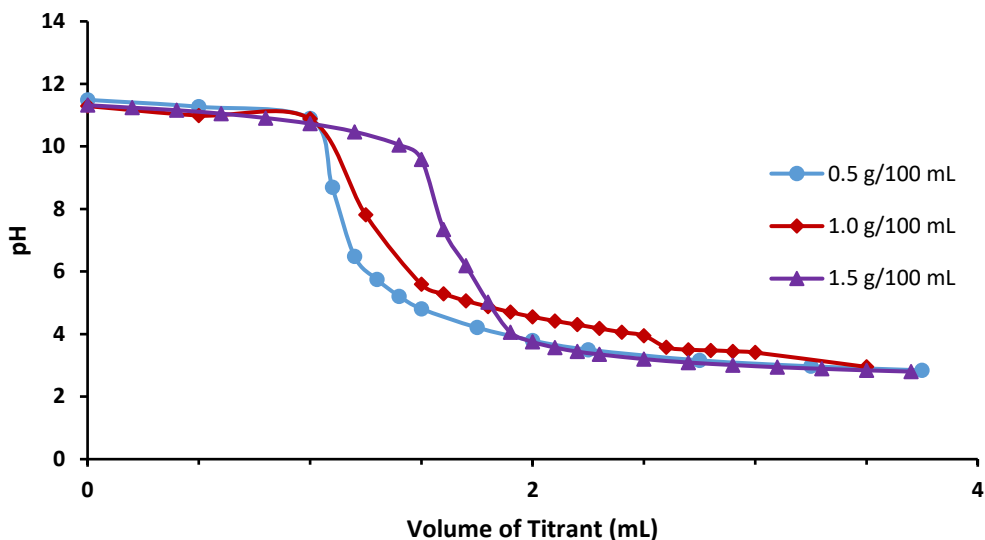
Figure 3. PZC of METAC MIP. The PZC was found to be 10.9.

To confirm the behavior of these MIPs in varying pH regimes, a pH study was conducted in a range of 3 - 12 (**Figure 4**). The TU MIP was shown to have relatively zero phosphate sorption over the entire pH range studied. The lack of removal indicates that there are no electrostatic attractions occurring, which confirms the lack of positively charged surface sites previously found in the PZC analysis.

The AT MIP showed different adsorption behaviors at different pH values. The AT MIP's behavior under different pH ranges is indicative of several things. First, the phosphate removal efficiency above pH 8 is lower than that in the ranges of pH 6 - 8. One-way ANOVA testing under a 95% confidence interval (CI) showed that the difference in removal percentages between these two regions is not statistically significant. However, it is believed that this change in removal efficiency likely corresponds to the polymer's PZC. Above pH 8.9, the positively charged surface sites that contribute to electrostatic removal of anions will be saturated by hydroxides and, therefore, unavailable for removal of

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phosphate. Although, it can be observed that even though the electrostatic attraction sites are saturated, there is still some phosphate removal occurring. This leads to the belief that the AT MIP is removing phosphate through its imprinted recognition sites. As the pH decreased below 6, the AT MIP showed decreased removal efficiency as well. One-way ANOVA testing under a 95% CI confirmed that the removal percentages from pH 3 – 5 and pH 6 – 8 differ significantly. This is attributed to one of two potential phenomena. First, as the pH decreases, the speciation of phosphate in solution will trend more toward phosphoric acid (H_3PO_4). It is possible that these phosphoric acid compounds may have reduced or negligible recognition by the imprinted sites on the MIP. Additionally, as anionic phosphate speciation trends toward phosphoric acid with a formal charge of zero, the polymer surface will become more positively charged due to continued protonation from decreasing pH. This will reduce or possibly eliminate any presence of electrostatic attraction between the compound and the polymer surface. The second phenomenon arises from the dissociation of hydrochloric acid (HCl) used to reduce the pH. Reducing the pH every 1 unit requires an order of magnitude more HCl than the previous unit. As the amount of HCl increases in the phosphate solution, more chloride (Cl^-) anions dissociate. These anions may compete for non-specific electrostatic binding sites or other available binding sites. These phenomena could consequently reduce the phosphate removal efficiency.

For the METAC MIP, the phosphate removal remains relatively constant from a range of pH 4 - 12. Once below a pH of 4, one-way ANOVA testing under a 95% CI shows the removal efficiency exhibiting a significant decrease. This can also likely be explained by the same phenomena used to describe the AT MIP's behavior at low pH. The general

behavior of the METAC MIP over varying pH regimes shows that it is stable and reliable across a wide range of pH.
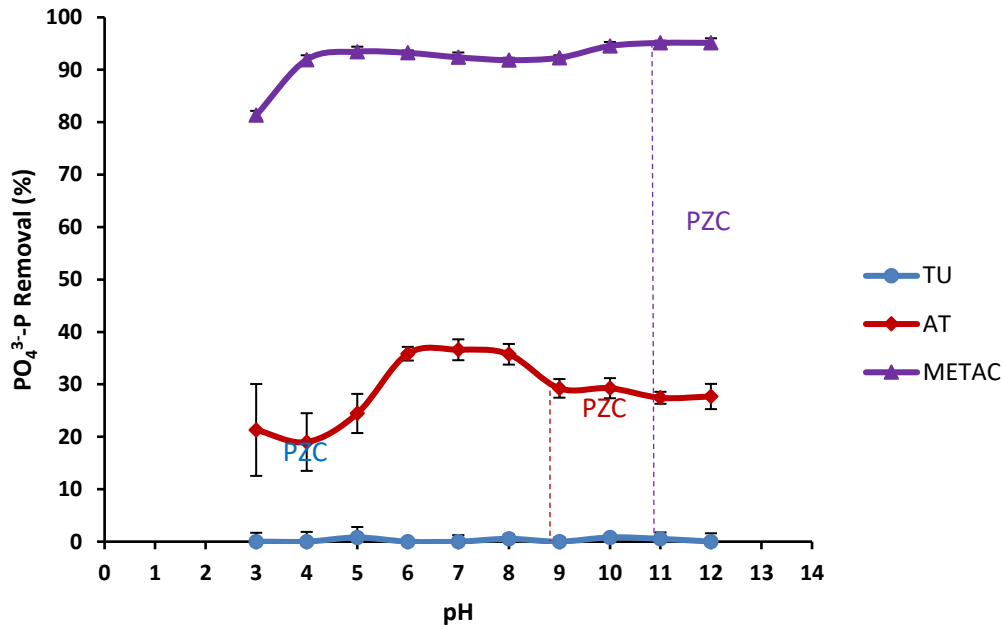
Figure 4. The Role of pH on Phosphate Removal (C_0 = 5 mg PO_4^{3-}-P/L)

2.3.2. Phosphate Removal Studies

2.3.2.1. Sorption Capacity of Polymers

The experimental data from phosphate sorption of the TU (NIP & MIP), AT (NIP & MIP), and METAC (NIP & MIP) are shown (**Figures 5**-**7**). Neither the TU NIP nor MIP showed successful phosphate removal. Any removal of phosphate found was typically associated with standard error. The results from this study corresponds those reported by Warwick et al. (2014a) who indicated that TU NIP was able to remove only 1% phosphate at C_0 = 0.78 mg PO_4^{3-}/L. However, this contradicts the same work done by Warwick et al. (2014a) who reported the TU MIP removing 22% phosphate at C_0 = 0.78 mg PO_4^{3-}/L. This

finding is not entirely surprising as Warwick et al. (2014a) also reported large error associated with phosphate removal by TU MIP.

Difference in P sorption between the AT MIP and its non-imprinted counterpart was deemed as insignificant based on one-way ANOVA testing under a 95% CI interval. While the means show significant differences, the lack of statistical significance is likely due to the large error associated with the sorption results. Additionally, the sorption capacity found for the AT MIP ($\sim$1 mg PO_4^{3-}-P/g polymer) is not high enough to be considered competitive with technologies that are already available (**Table 2**).

The METAC MIP showed a sorption capacity that could very well be considered competitive with available technologies ($\sim$11 mg PO_4^{3-}-P/g polymer). The METAC MIP also had a much lower standard error in the phosphate removal data, making the phosphate removal process more reliable and predictable. However, one-way ANOVA testing under a 95% CI showed that the difference in sorption capacities between the METAC MIP and NIP was not significant. This indicates that electrostatic attraction of phosphate due to a positive surface charge of the METAC polymer significantly contributes to phosphate removal, while the assumedly selective binding sites from imprinting phenlyphosphonic acid possibly only contribute marginally.
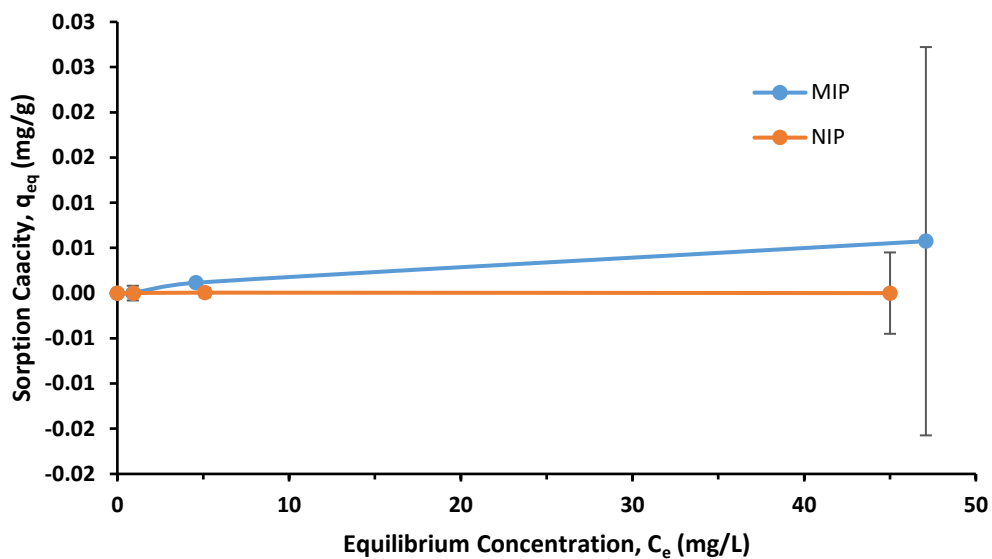
Figure 5. Sorption Capacities of TU NIP and MIP
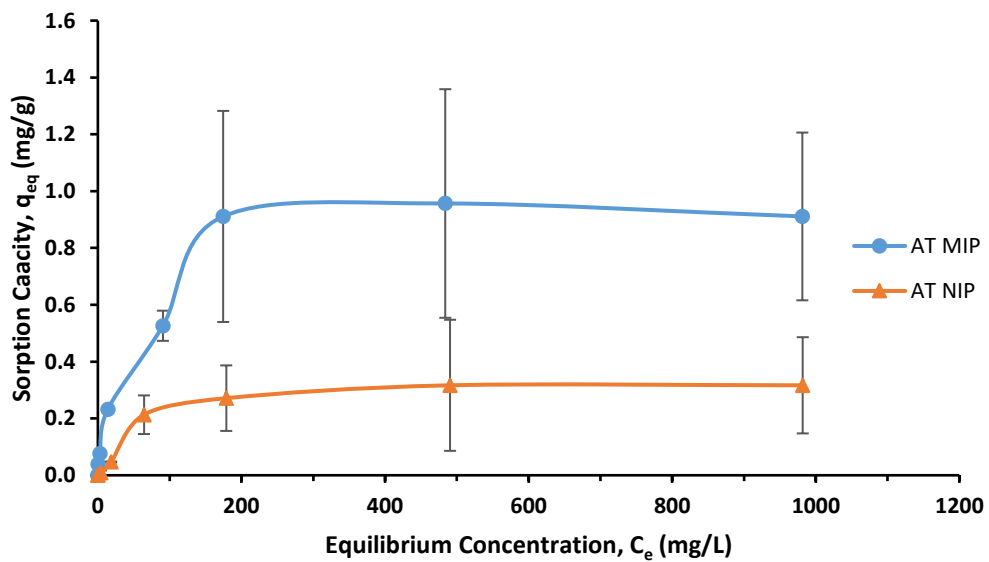
Figure 6. Sorption Capacities of AT NIP and MIP
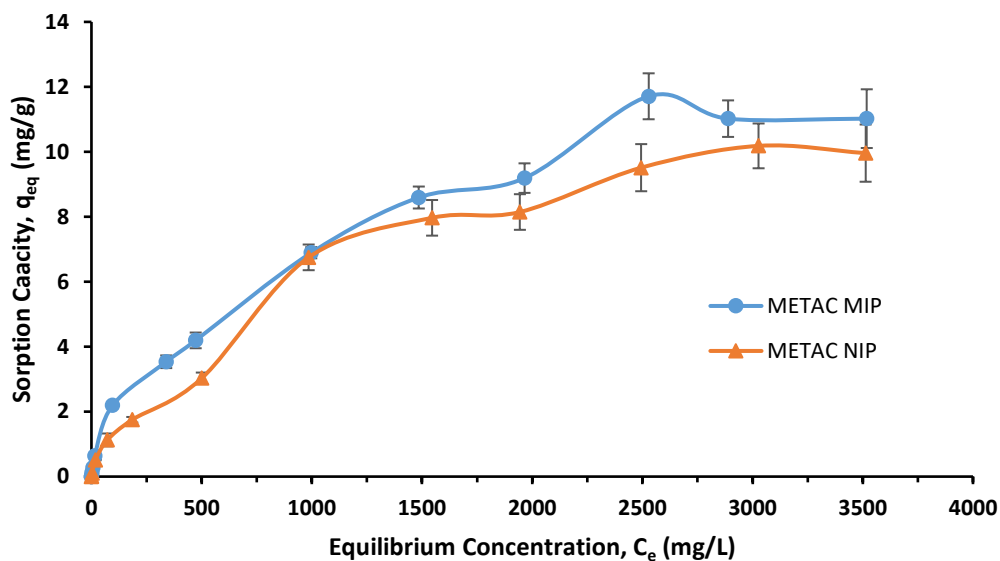
Figure 7. Sorption Capacities of METAC NIP and MIP

2.3.2.2. Interference Studies

It is necessary to determine the feasibility of applying these MIPs in actual lake water for P recovery, specifically the proficiency of the MIPs in the presence of coexisting anions and compounds which might compete for active binding sites. At this point in the study, the TU MIP was no longer considered for testing as it was unable to remove any significant amount of phosphate from solutions under varying pH and initial phosphate concentrations. The AT MIP also showed poor sorption capacity to be considered as a feasible sorbent for P recovery applications. However, Kugimiya et al. (2008) reported that their AT MIP had the ability to selectively remove phosphate from amongst coexisting ions. Kugimiya et al. (2008) did not provide any data from an interference study to support this claim. The AT MIP was included in this study in the hope that it may still provide potential as a low-efficiency selective sorbent. METAC MIPs worked well and were also considered for this study.

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The ions and their associated concentrations used in this interference study were selected due to their typical presence in surface waters. One-way ANOVA testing under a 95% CI showed that the AT MIP experienced significant interference for binding sites from certain competing anions. The AT MIP experienced most of its significant reduction in phosphate removal from nitrate, chloride, and sulfate (~55 - 85% reduction). Significant reduction was also seen with NOM at concentrations of 10 mg/L or higher (~18 – 22% reduction) (**Figure 8**). Minor interference was experienced from NOM while bicarbonate and humic acid showed no significant interference. The METAC MIP followed the same pattern of interference as one-way ANOVA testing under a 95% CI showed that it only experienced significant reduction in phosphate removal from nitrate, chloride, and sulfate (~15 - 85% reduction) (**Figure 9**). Of those three interfering anions, interference from nitrate at a concentration of 10 mg/L did not show to significantly differ from the control's removal. These results indicate that the AT MIP is not selective toward phosphate even though indicated otherwise by Kugimiya et al. (2008). This was somewhat expected as the PZC value indicated the AT MIP as having a positive surface charge below a pH of 8.9, and a positively charged surface provides sites for non-specific binding with any negatively charged compound. The METAC MIP also showed poor selectivity toward phosphate in the presence of coexisting anions. The same explanation given for the AT MIP's failure to selectively remove phosphate can be given to the METAC MIP as it had a PZC at pH 10.9.
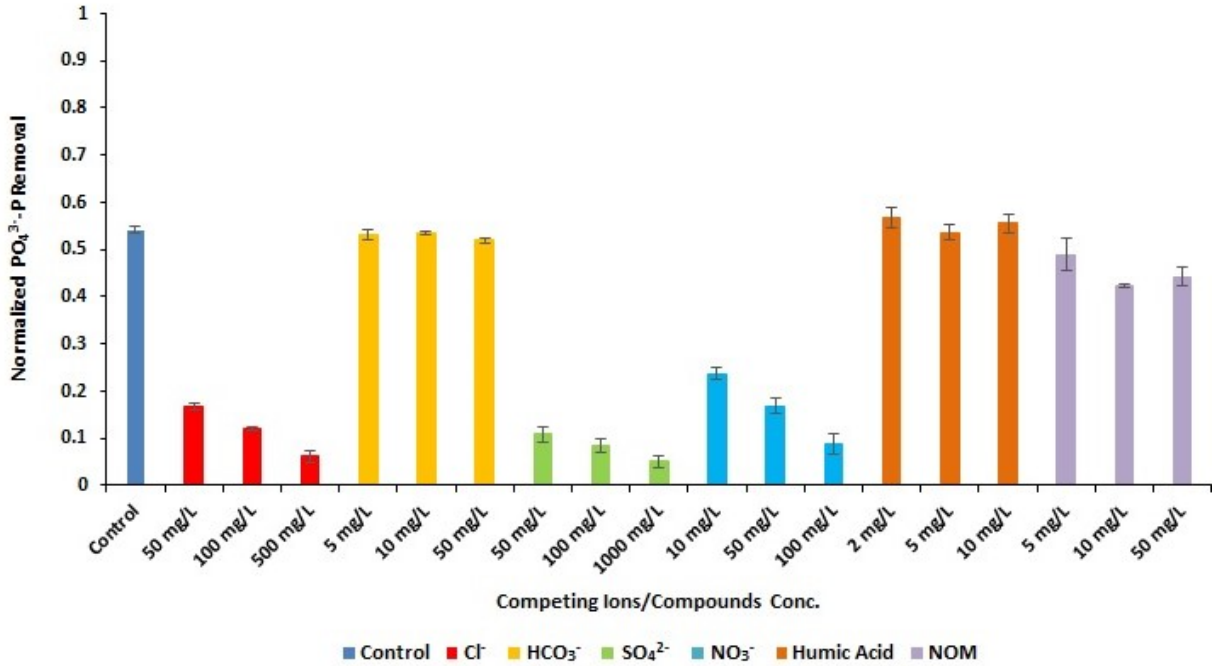
Figure 8. Competition of Common Coexisting Anions and Compounds for Binding Sites on AT MIP (C_0 = 5 mg PO_4^{3-}-P/L)
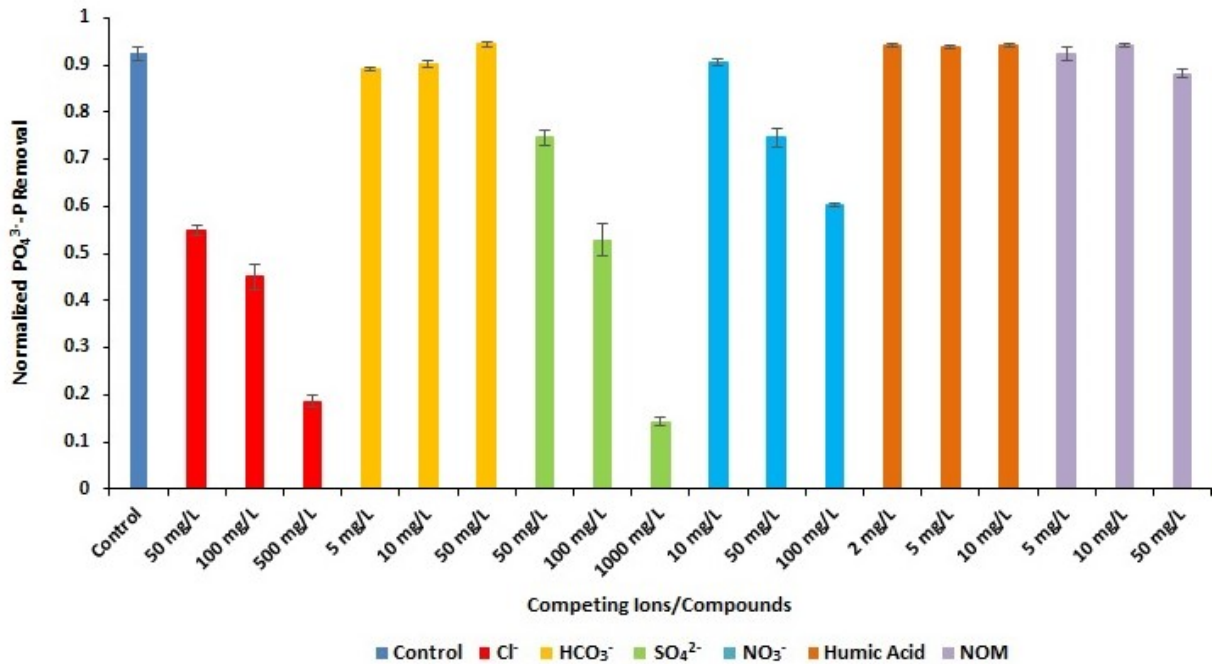
Figure 9. Competition of Common Coexisting Anions and Compounds for Binding Sites on METAC MIP (C_0 = 5 mg PO_4^{3-}-P/L)

At first glance, it may appear that nitrate, chloride, and sulfate are the only ions to be concerned with for interference. This would make any surface water lacking significant quantities of those three anions remiss of interference toward MIP sorption of phosphate. While it is unrealistic to assume that surface waters will be lacking all three of these anions, it was also determined that the physicochemical compositions of those three anions were not the predominant reason for interference. The METAC MIP interference data collected were reorganized, only considering the normalized PO_4^{3-}-P removal and the equivalent charge concentration (meq/L) of the anions applied (**Figure 10**). Fluoride (F^-), Bromide (Br^-), and acetate ($C_2H_3O_2^-$) were included in this reorganization to bolster confidence in the pattern shown. As the presence of negative charge in solution increased, the normalized phosphate removal decreased. It can also be seen that it did not matter as to which anion was used to provide competing negative charge; only the amount of negative charge present in solution impacted phosphate removal. This confirms that the specific anion size, shape, and chemical composition do not play inherently significant roles in the interference with phosphate removal. It also confirms that the METAC MIP is removing phosphate, at least to some degree, through non-selective electrostatic binding to positively charged surface sites.
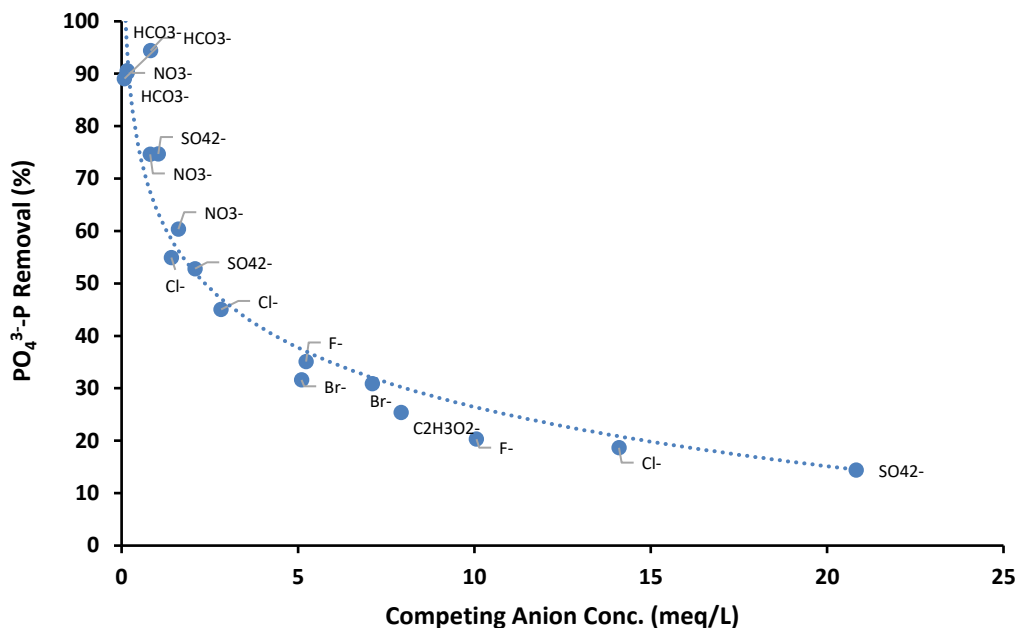
Figure 10. The Effect of Negative Charge Present in Solution on Phosphate Removal (C$_0$ = 5 mg PO$_4$$^{3-}$-P/L)

2.3.2.3.   Effect from Increasing Template:Monomer Ratio

The original template:monomer ratio used for synthesis of all of the MIPs was 1:4. It was hypothesized that increasing this ratio will increase the amount of imprinted recognition sites. Under this assumption, it is also plausible that more imprinted recognition sites could lead to better selectivity toward phosphate.

At this point in the study, the AT MIP was eliminated from consideration for testing as it had already reached its maximum template:monomer ratio. This maximum ratio was reached due to solubility issues upon the addition of more template. Insoluble template does not affect the template:monomer ratio as it does not form complexes with the monomer. The decision to remove AT from the study was made due to poor sorption capacity and lack of selectivity at the maximum (1:4) and minimum (0:4) available template:monomer ratio for design. The METAC MIP was considered for further study

because even though it did not show a resistance to interference from competing anions, it had a sorption capacity which can be considered competitive with many other technologies. There was potential to increase template:monomer ratio in METAC MIP, which could potentially solve the interference issue. Two more template:monomer ratios (4:4 and 12:4) were used to determine their impacts on interference reduction and sorption capacity (**Table 4**). A ratio of 12:4 was the highest ratio considered because that was the point where the template reached saturation.

Table 4. Components for METAC Reagent Mixtures with Various Template:Monomer Ratios

Template: Monomer Ratio	Notation	Monomer Qty.	Cross-linker	Solvent	Template
0:4	MN	830 mg (4 mmol)	3.8 mL (20 mmol)	5 mL	0 mg (0 mmol)
1:4	MM	830 mg (4 mmol)	3.8 mL (20 mmol)	5 mL	158 mg (1 mmol)
4:4	MM4	830 mg (4 mmol)	3.8 mL (20 mmol)	5 mL	632 mg (4 mmol)
12:4	MM12	830 mg (4 mmol)	3.8 mL (20 mmol)	5 mL	1,896 mg (12 mmol)

The impact of the template:monomer ratio on reduction of anion interference was analyzed under the same conditions as previously mentioned, where negative charge was introduced in varying amounts by several different anions. One-way ANOVA testing under a 95% CI indicated that an increase in the template:monomer ratio made an insignificant difference on the METAC MIP's resistance to interference from competing anions (**Figure 11**). This confirms that increasing the template:monomer ratio does not reduce or overcome the presence of electrostatic interactions between anions and the METAC MIP.
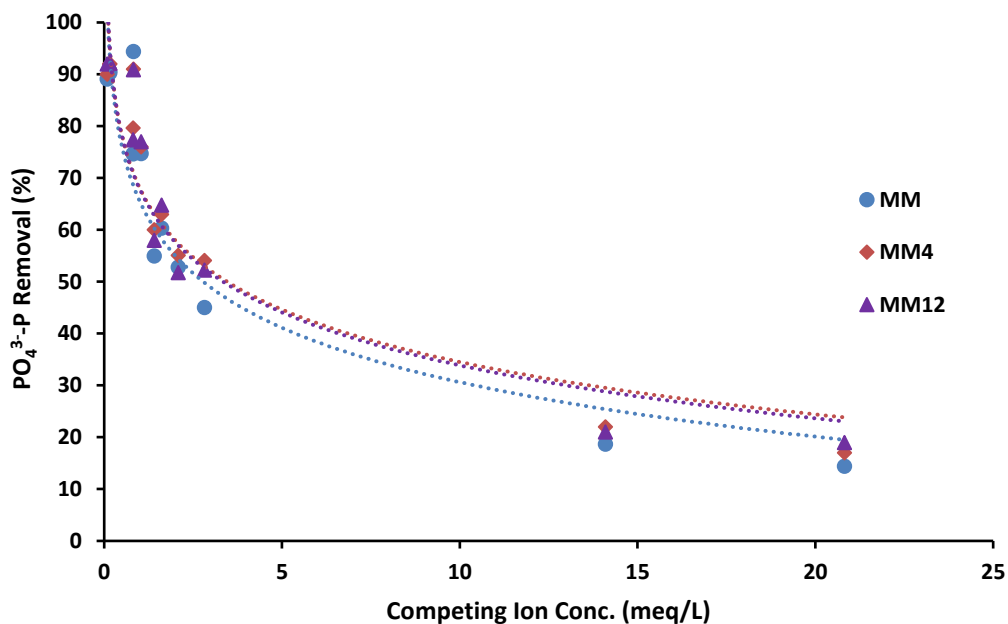
Figure 11. The Effect of Negative Charge Present in Solution on Phosphate Removal by METAC MIPs with Varying Template:Monomer Ratios (C_0 = 5 mg PO_4^{3-}-P/L)

Next, the impact of the template:monomer ratio on the sorption capacity was studied. The sorption capacities of all three METAC MIPs (MM, MM4, and MM12) along with the control, METAC NIP (MN), were compared. The data were fit to the Freundlich and Langmuir isotherm models (**Figures 12 & 13**). The isotherm coefficients (K_F, n, K_L, q_m) and fit (R^2) were determined from the linearized forms of **Eqs. 1 & 2** and are summarized (**Tables 5 & 6**).

Table 5. Coefficients and Fit of Freundlich Isotherm Model

Polymer	Freundlich Coefficients		Fit
	K_F	n	R^2
MM12	0.223	1.682	0.9715
MM4	0.230	1.776	0.9727
MM	0.310	2.180	0.9904
MN	0.061	1.504	0.9690

Table 6. Coefficients and Fit of Langmuir Isotherm Model

	Langmuir Coefficients		Fit
Polymer	K_L	q_m	R^2
MM12	0.0012	30.960	0.7114
MM4	0.0024	19.608	0.9391
MM	0.0037	11.669	0.9507
MN	0.0019	11.403	0.9432
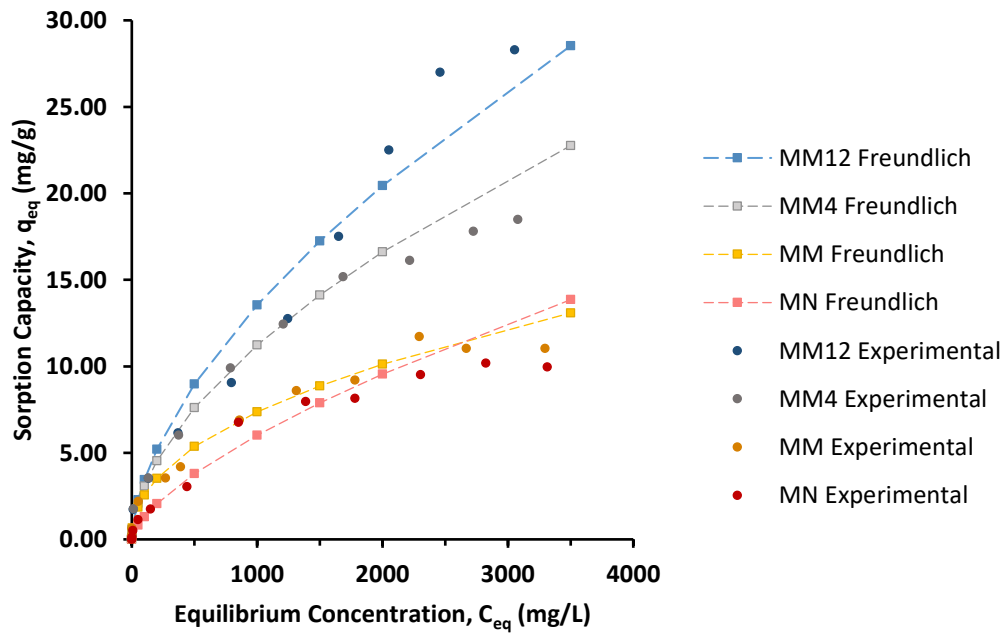
Figure 12. Comparison of METAC Freundlich Isotherm Model with Varying Template:Monomer Ratios
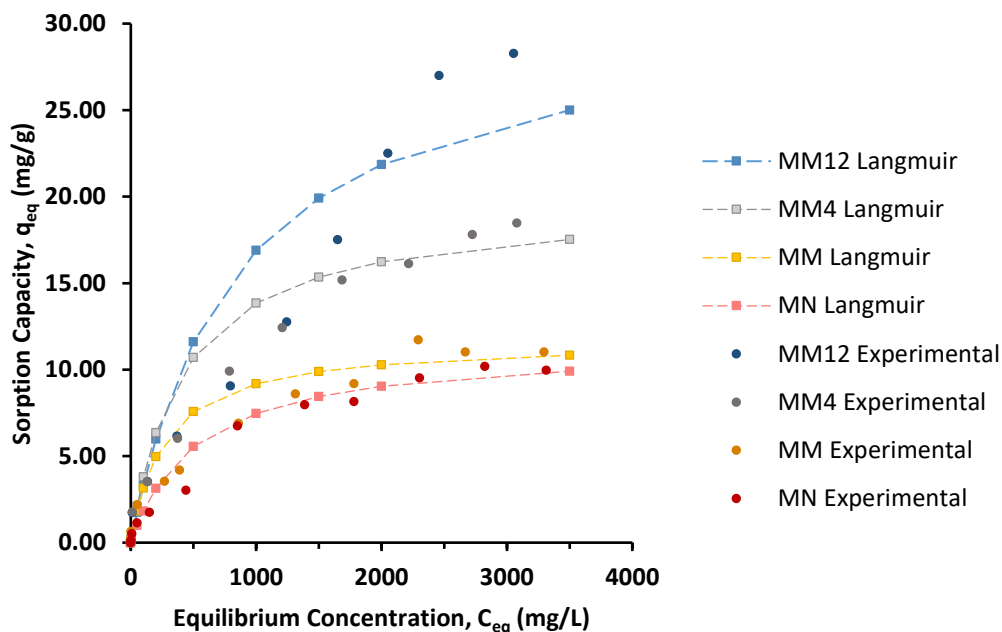
Figure 13. Comparison of METAC Langmuir Isotherm Model with Varying Template:Monomer Ratios

One-way ANOVA testing under a 95% CI showed that with an increase in template:monomer ratio, the sorption capacity significantly increased as well. The increase in sorption capacity indicates that more active phosphate binding sites are becoming available on or within the polymer matrix. Due to the increase being a direct result of increasing the amount of template, it is assumed that these active binding sites are selective recognition sites. Based on this finding, it appears that MIPs could be designed for higher sorption capacities by increasing their template:monomer ratios. While this design parameter is significantly influenced by monomer and template saturation point within the solvent used, this finding could still help provide the framework for future development of MIPs.

2.3.2.4. Removal from Low Concentration Phosphate from Lake Water

In order for a sorbent technology to be feasible for surface water application, it must be able to remove phosphate concentrations below environmentally relevant levels (50 µg PO_4^{3-}-P/L). Thus, the MM12 was tested in a low concentration phosphate solution (~150 µg PO_4^{3-}-P/L). Even though interference studies were conducted on the METAC MIPs, it was still necessary to see how MM12 behaves in actual lake water. MM12 was selected for further study over the other METAC MIPs (MM and MM4) due its heightened sorption capacity, making it the sorbent with the most potential. After retrieving the samples and analyzing P content, it was found that concentrations of P were lower than 50 µg PO_4^{3-}-P/L. Therefore, the lake water samples were artificially spiked with KH_2PO_4 to bring their concentrations to a range of 100 – 200 µg PO_4^{3-}-P/L. Lakes sampled and spiked are as follows: Lake Katrina (185 µg PO_4^{3-}-P/L), Holy Name Lake (180 µg PO_4^{3-}-P/L), Lake Ardmore (212 µg PO_4^{3-}-P/L), Lake Independence (187 µg PO_4^{3-}-P/L), and Half Moon Lake (157 µg PO_4^{3-}-P/L) (**Figure 14**). It was observed that MM12 was able to remove phosphate to levels lower than 50 µg PO_4^{3-}-P/L in synthetic water and lake waters. One-way ANOVA testing under a 95% CI indicated that differences in removal between synthetic water and lake waters were insignificant for all except Lake Independence and Holy Name Lake. This difference is likely attributed to competing anions present in the lake waters. This study reinforces the concept of using MIPs to remove phosphate below environmentally relevant levels; a feat that several established technologies for phosphate removal cannot accomplish.
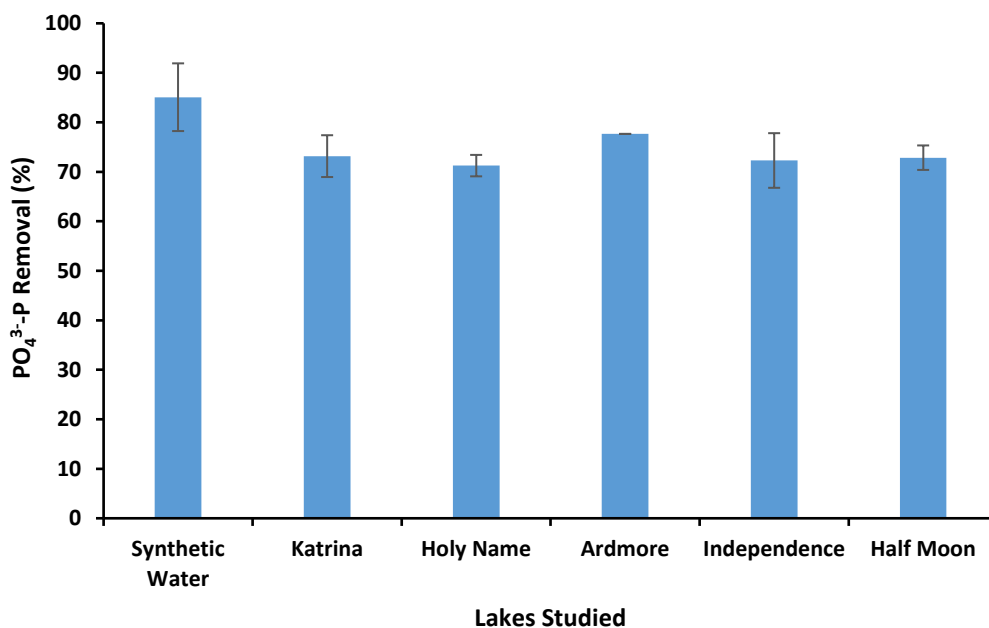
Figure 14. Phosphate Removal from Select Minnesota Lakes with MM12

2.3.2.5.   Influence from Temperature

For MIPs to have any future as sorbents for eutrophic lake remediation, they must

be able to handle seasonal variations in water temperatures. Lake water temperatures can

reach as low as 4 °C (before freezing begins to occur) in the winter and can occasionally

break 30 °C in summer time conditions (http://www.gvsu.edu/videticp/stratification.htm,

2001). To assess the behavior of MM12 in a wide range of temperature conditions, the

polymer and phosphate solution were studied in an environmental chamber where the

temperature could be adjusted, monitored, and kept constant for extended periods of time

(**Figure 15**). One-way ANOVA testing under a 95% CI showed there was no significant

variation in phosphate removal efficiencies observed between temperatures. This suggests

that MM12, and potentially most other MIPs, can be used effectively during any season.

70
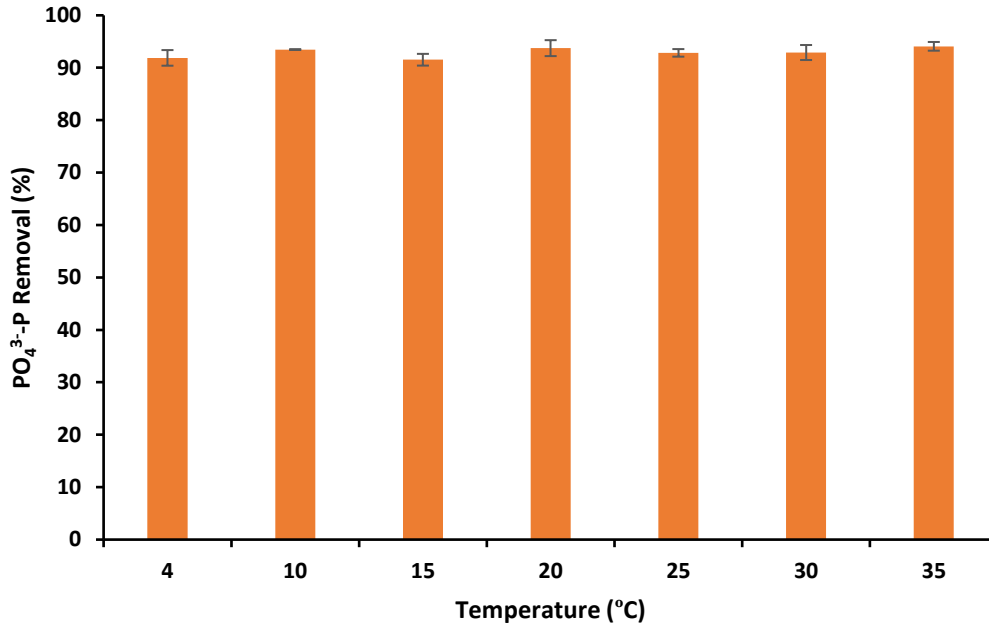
Figure 15. Influence of Temperature on Phosphate Removal Efficiency with MM12 (C_0 = 5 mg PO_4^{3-}-P/L)

2.3.3. Biochemical Availability and Potential for P Recovery

2.3.3.1. Biodegradability

In order for used MM12 to be reused in agriculture, it must be biodegradable. Non-biodegradable materials will reside in precious soil space for years on end without sorbed P being bioavailable to plants. Toxicity toward microorganisms and plants is one limiting factor that has prevented many wastewater P-recovery technology products from being feasible for reuse. Wastewater also contains heavy metals that are also removed by adsorbents. It is these heavy metals that cause toxicity toward microorganisms and plants, resulting in detrimental effects on croplands (Satorius et al., 2012). To assess MM12's potential for biodegradability, a respirometer experiment was conducted using the BODTrak (Hach, USA). Triplicate samples of the MM12 in a solution of bacteria and BOD nutrient were compared against triplicate samples of a control with only bacteria and BOD

nutrient. After correcting the BOD reading for dilution and seeding factors (**Appendix 3.6**)

the average readings were reported here (**Figure 16**). Samples with MM12 present showed

higher levels of BOD when compared to the samples without them. This indicates that the

MM12 served as a source of food for microorganism consumption, and that MM12 is

biodegradable. This conclusion opens up the possibility that MM12 (with P sorbed) may be

applicable for fertilizer use. Further studies would need to be conducted to ensure that

MM12 biodegradation and its by-products do not result in toxicity in plants.
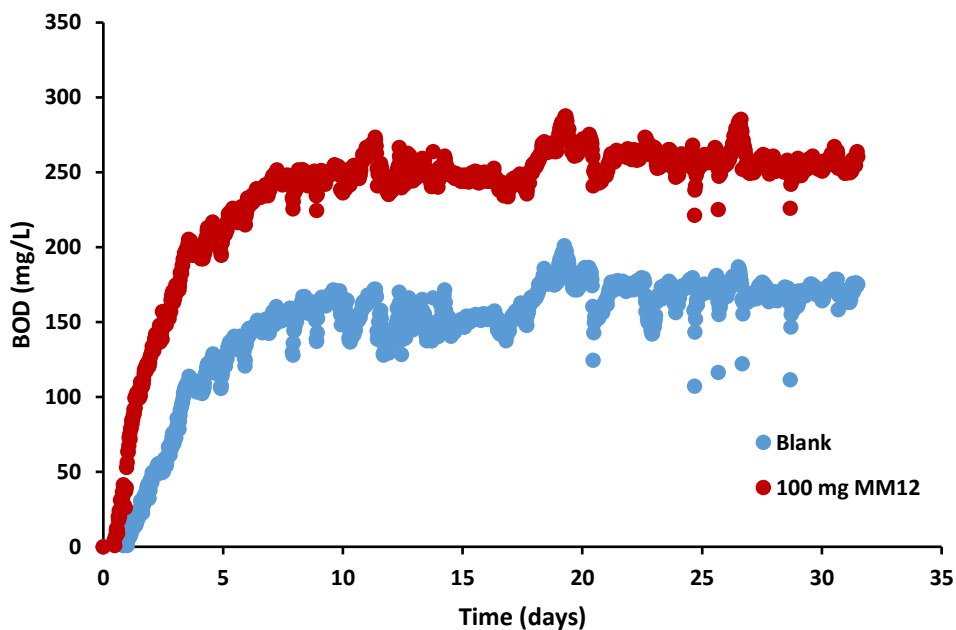
Figure 16. Respirometric Data of Biodegradability of MM12

2.3.3.2.   Desorption Study

To understand how MM12 may behave as a soil amendment, the P-release

mechanism needs to be understood. Fertilizer supplements that release P immediately are

not as advantageous as slow P-release technologies. This is because plants cannot uptake

all of the P that is supplied to them immediately. The current problem with fertilizer

applications is that 40% of P applied results as surface runoff due to crops having the

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inability to uptake the P at high rates (Rittmann et al., 2011). Technologies that can release P in small amounts over a long period of time are more beneficial to plant growth as the slow release behavior more ideally matches with plant uptake kinetics.

A desorption study was conducted using spent (**Figure 17**) MM12. Fresh MM12 was initially placed in high phosphate concentration solution (2000 mg PO_4^{3-}-P/L). Analysis of phosphate removal from bulk solution showed that the MM12 had ~17 mg of sorbed PO_4^{3-}-P available for release from MM12. After the six testing cycles (four total days), the desorption study indicates that MM12 released 29.3 % (~5 mg PO_4^{3-}-P) of sorbed phosphate. The majority (19.2 %) of that 29.3 % was released during the first cycle (6 hours). Each subsequent cycle released less P than the cycle before it. It is believed that this release of P is coming from the phosphate that is bound to the polymer electrostatically. This pattern of continuously decreasing P-release rates is expected to continue until the remaining majority of P, which is assumed to be strongly bound to imprinted recognition sites within the polymer matrix, is released. To be able to access the P bound in this manner, the polymer would need to be degraded over a time. This mechanism of P-release would establish a slow release regiment that would be highly beneficial for plant growth. Studies in actual conditions (soil with growing plants) would need to be conducted in order to assess the P-release mechanism.
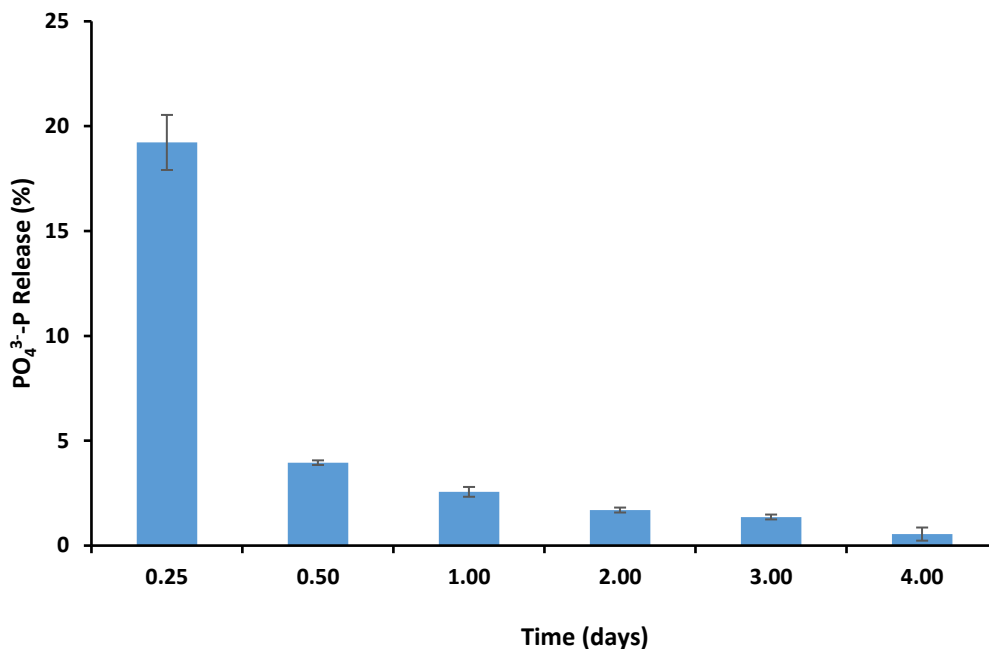
Figure 17. Phosphate Release from MM12 during Desorption Study

2.3.4. MIP Characterization

2.3.4.1. SEM/EDS Analysis

Characterization of the METAC MIP's surface via SEM/EDS analysis was done to gain

an insight to the polymer's morphology. Additionally, the morphology of the polymer was

studied under variable template:monomer ratios to see what affect the amount of

templating could have on surface morphology. The first analysis was completed at the

micron-scale using a low-resolution SEM (**Figure 18(a-d)**). An immediate trend can be

found from the change in surface morphologies from one template:monomer ratio to the

next. At the lowest ratio, MN (0:4), the surface appears extremely rough and heterogeneous

in nature (**Figure 18(a)**). As the template:monomer ratio increases, the surface tends to

become smoother and more homogenous than the previous until reaching the maximum

ratio, MM12 (12:4), where the surface appears extremely smooth and flat (**Figure 18(b-**

d)). (Wu, 2012) previously discussed how MIPs tend to have heterogeneous chemical structures at low template:monomer ratios. Surface characterization was never completed in tandem with that work. However from that work, it's reasonable to believe that increasing the template:monomer ratio will consequently increase homogeneous structure of MIPs. An increase in the overall homogeneity of the polymer matrix could explain the trend seen in surface morphology as homogeneous nature may generate more uniformity in surface structure.
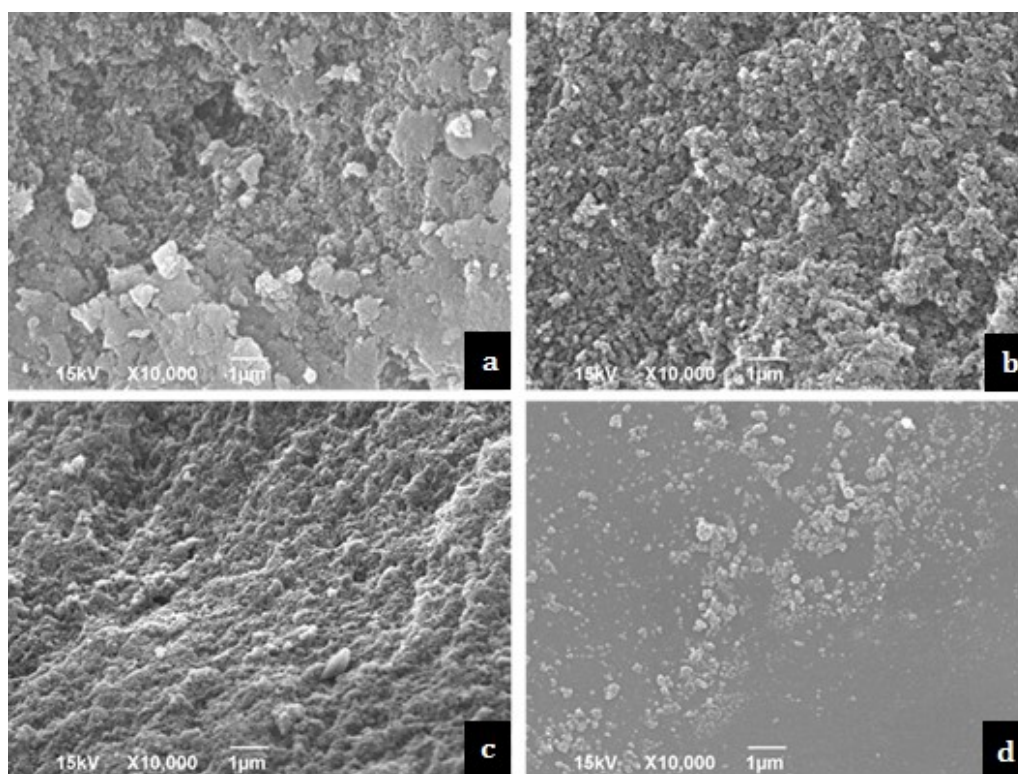
Figure 18. Low-resolution SEM Characterization of MN (a), MM (b), MM4 (c), and MM12 (d)

After seeing the changes to the surface on the micron-scale, it was deemed necessary to investigate what type of changes may have happened at the nano-scale. To view the polymer surface at this scale, high-resolution SEM was employed (**Figure 19(a-d)**). One of the primary targets of this analysis was to characterize potential changes in the

pore distribution or quantity. The high-resolution images are not of the highest quality as the polymers heated easily, resulting in image drift. The non-imprinted polymer (MN) was the only polymer void of explicit nanopores. While it was originally believed that the development of nanopores would be the result of template extraction from the polymer matrix, the nanopores are of larger size than the template molecules. Additionally, there was no distinguishable pattern change in nanopore distribution or quantity between the different template:monomer ratios at this scale. One general trend found was the localization of nanopores in clusters, which were scattered across the polymer particles. The template molecules may be bound into the polymer matrix in localized clusters, and the extraction of those clusters may be responsible for the development of the observed nanopores, however that remains to be unknown. It should be noted that even though MN did not show explicit nanopores, it still has a porous structure. The general porous structure of MN and the other MIPs is likely the result of the extraction of the porogen. During polymerization, the solvent (porogen) phase-separates and is trapped within the polymer matrix. The extraction process then pulls the porogen out, leaving behind a porous polymeric structure. Many of the nanopores were pointed out with white arrows to help distinguish them from the natural porous structure.
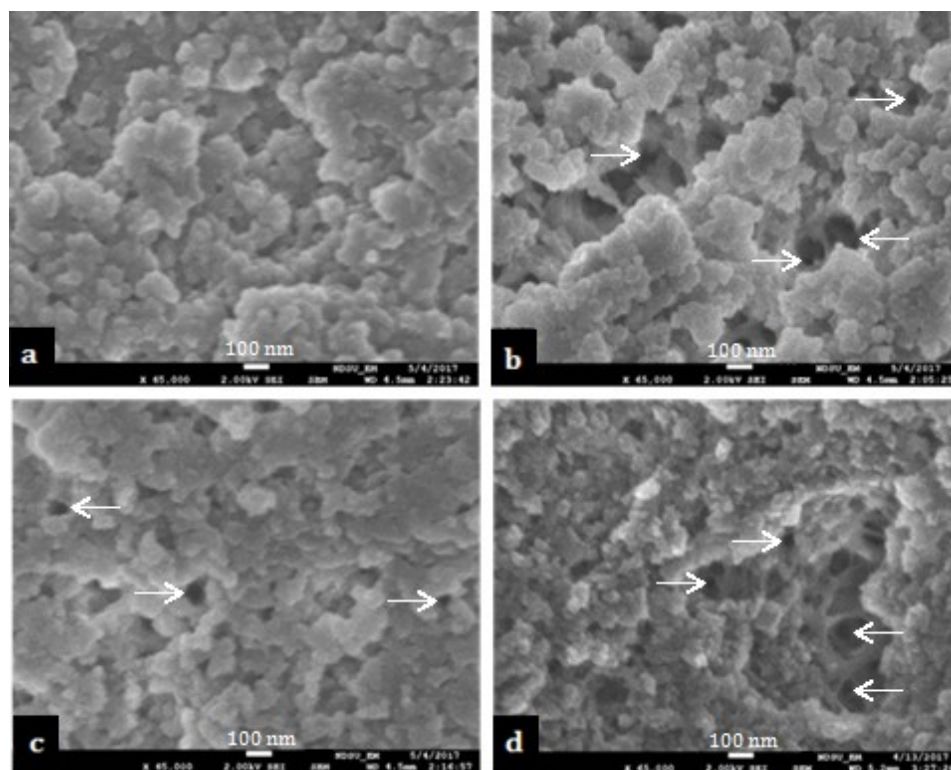
Figure 19. High-resolution SEM Characterization of MN (a), MM (b), MM4 (c), and MM12 (d)

Lastly, EDS analysis was performed on MM12 (**Figure 20**). This analysis was performed to validate that the polymer is physically removing phosphate, as opposed to precipitating the phosphate from solution. For a control, EDS analysis was also performed on MM12 which had not been placed in a phosphate solution (**Figure 21**). On the blank MM12, EDS results showed the surface consisting of predominantly carbon and oxygen. Trace amounts of chloride were found, likely trapped in the polymer matrix after the dissociation and polymerization of the METAC monomer. Gold was found as well; however, the presence of gold is attributed to the coating that was used for sputtering on the polymer to provide conductivity to the sample. On the MM12 with sorbed phosphate, EDS results showed that the surface once again consisted of predominantly carbon and oxygen. Trace amounts of chloride was also found. Potassium was also detected in trace amounts

on the surface, which is likely the result of some potassium dissociating from KH_2PO_4 and being adsorbed onto the surface of the polymer. Lastly, phosphorus was found on the surface of the polymer. The presence of phosphorus confirms that phosphorus was adsorbed as phosphate onto the surface of the polymer.
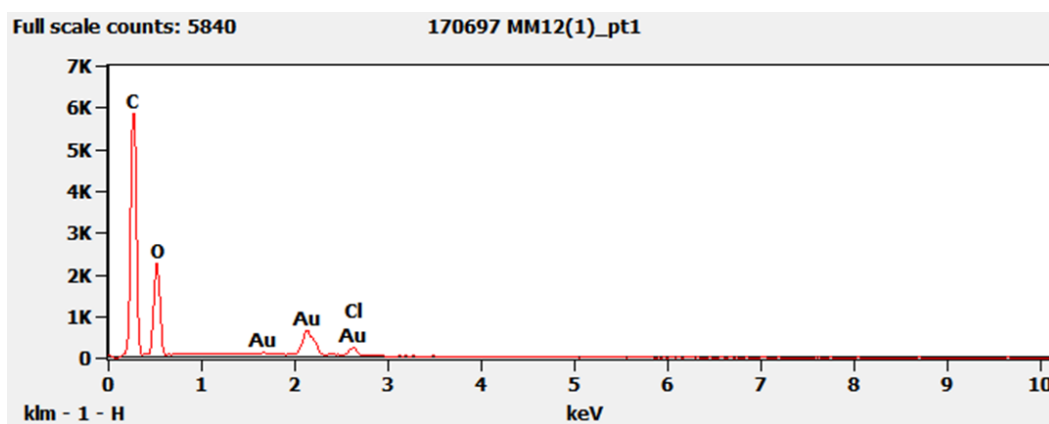
Figure 20. EDS Data from Analysis on Blank MM12
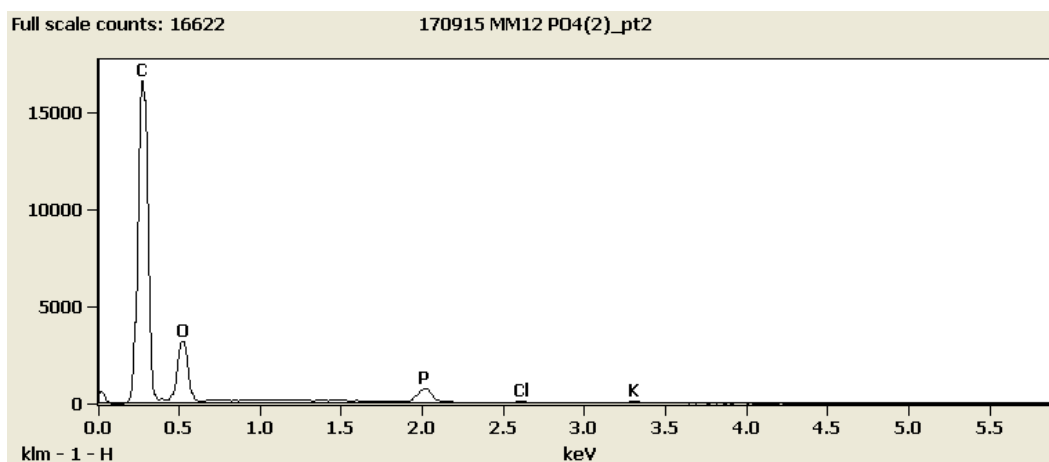
Figure 21. EDS Data from Analysis with Adsorbed Phosphate

2.3.4.2. *FTIR Spectroscopy*

FTIR spectroscopy can help determine what time of bonding is present within a material based on the unique rotational and vibrational resonances that result from bonds being gently excited by infrared radiation. FTIR analysis can help determine whether or not the template, phenylphosphonic acid, is being imprinted. To be imprinted, the template

must first be bound in a complex with the monomer. Presence of the template bonds in the polymer matrix will confirm that the template is being imprinted. On top of that, loss of the template bonds after the extraction process will indicate effective template removal from the Soxhlet extraction. Lastly, if there is any selective recognition of phosphate in the MIP, it will be observed here. Phosphate, which is selectively removed by the recognition sites in the MM12, will be bound into the polymer matrix. Therefore, P binding should be seen. FTIR analysis was performed on unfinished MM12 just after synthesis without extraction (MM12 UNF), fresh MM12 after extraction (MM12), and spent MM12 after phosphate sorption (MM12 PF) (**Figure 22**).

In the MM12 sample before template extraction, there are several bands unique to the spectra as compared to after extraction. Weak bands at 1600 and 1500 cm^{-1} indicate the presence of C=C stretching in a mono-alkyl arene ring structure (R-C_6H_5). The band at 990 cm^{-1} is indicative of a phosphorus ester functional group (P-OR). Lastly, strong band intensities at 700 and 680 cm^{-1} identify =C-H bending in mono-alkyl arenes (R- C_6H_5). Phenylphosphonic is the only compound with a ring structure that was introduced into the reagent mixture for synthesis. Thus, it is clear that these bands are representative of phenylphosphonic acid's binding within the polymer matrix. Additionally, it can be seen in the spectra after extraction (MM12) that these bands disappear entirely. This suggests that phenylphosphonic acid is being effectively extracted from the polymer matrix, leaving imprinted recognition sites behind.

Signals from P=O stretching can generally be seen in the 1100 - 1200 cm^{-1} range. The 1130 cm^{-1} band is located right in that region. However, there are also intense signals coming from C-O stretching in that region. The coexistence of these two signals in the same

region makes it impossible to use that band region as an identifier for phosphate sorption. However, comparing the spectra of MM12 after phosphate sorption (MM12 PF) to the other spectra before phosphate sorption shows one unique difference. In the spectra before phosphate sorption, the relative intensities of the bands at 510 cm^{-1} and 490 cm^{-1} are equal. After phosphate sorption, the relative intensity of the band at 510 cm^{-1} significantly increases compared to that of the band at 490 cm^{-1}, which remains constant. The increase in relative intensity is likely constituted by phosphate conjugation around nitrogen, forming $(PNCl_2)_3$. From observation, it can be seen that the 510 cm^{-1} band was present before phosphate extraction. Before phosphate extraction, the appearance of this band was likely due to the presence of an alkyl halide with chlorine as its halogen component (C-Cl). Both of these explanations would justify the appearance of chlorine in the EDS analysis.
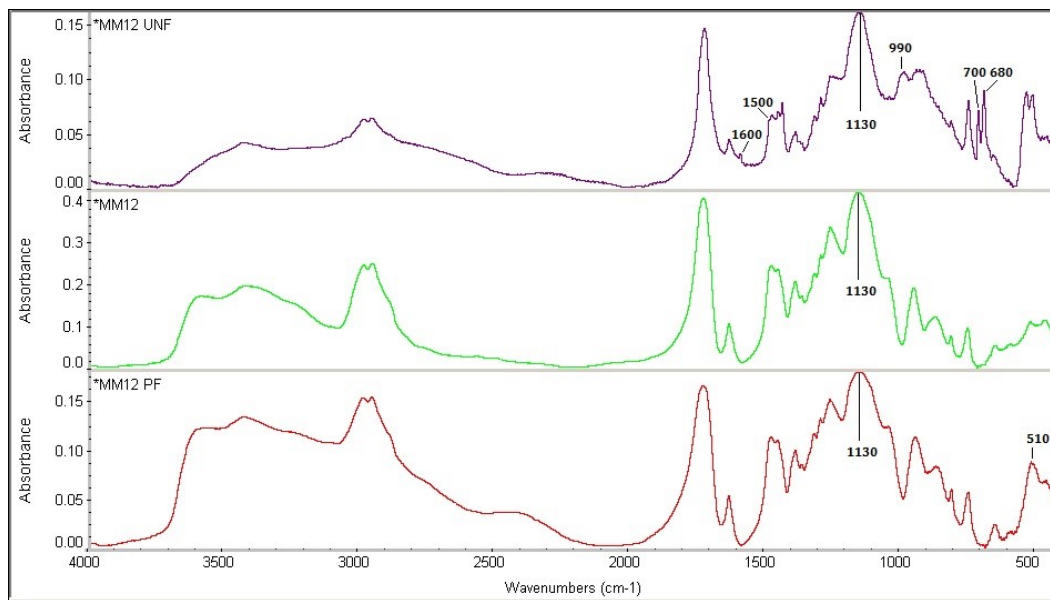
Figure 22. FTIR Spectra of MM12 during Different Phases of Life Cycle

While the analysis of the FTIR spectra does not prove selective binding of phosphate, it does validate the existence of bound phosphate. This suggests that the two

primary mechanisms of phosphate removal with the MM12 are electrostatic attraction to the surface and phosphate-binding with nitrogen in the recognition sites.

2.3.4.3. *Swelling Studies, and Prediction of Porosity and Hydrated Diameter*

Upon immersion of MM12 particles in DI water for 30 minutes, it was found that the MM12 particles exhibited the following swelling capability (**Table 7**). From this information, the porosity of and hydrated diameter of the particle average was determined (**Table 7**). Such a high degree of swelling is indicative of the polymer likely having higher available surface area for anion interaction when immersed in water.

Table 7. Swelling, Predicted Porosity, and Predicted Hydrated Diameter of Hydrated MM12 particles

MM12 Samples	Initial Dry Weight (W_i, g)	Hydrated Weight (W_s, g)	Swelling (%) eq. (4)	Predicted Porosity of Hydrated Particles (ε, %)	Predicted Diam. of Hydrated Particles (D, mm)
1	1.0236	2.5222	146.4	60.9	1.68
2	1.1650	3.0674	163.3	63.5	1.79
3	1.4737	3.6570	148.2	61.2	1.90
Average	1.2208	3.0822	152.6 ± 7.6	61.9 ± 1.1	1.79 ± 0.09

2.3.5. **Mechanism of Phosphate Removal**

2.3.5.1. *Investigation of Partial-selectivity*

At this point, there has been little to no evidence of the synthesized MIPs having any form of selective phosphate recognition. The MM12 can be seen to have a significantly higher P sorption capacity than MM4, MM, and its non-imprinted counterpart, MN (**Figure 23**). This increased sorption capacity can be attributed to one of two situations. First, the increase in template:monomer ratio could result in greater porosity of the polymer, allowing for more anions to be trapped within the polymer pores by non-selective

electrostatic attraction. The prospective alternative is that the increased

template:monomer ratio generates a greater quantity of recognition sites within the

polymer matrix which have the capability of selectively binding phosphate. To validate

whether the increase in sorption capacity was from selective phosphate binding or non-

specific anion attraction, an experiment was designed. It has already been shown from

interference studies that MM12 binds anions non-specifically due to having a positive

surface charge. The underlying question is whether or not all of the phosphate removal

observed was non-specific binding.

Mixing MM12 in extremely high anion concentration (110 meq/L) solution allows

the non-specific binding surface sites to become saturated by the anion. If any of the

recognition sites generated from imprinting are actually selective toward phosphate, their

phosphate recognition capabilities will not be affected by the presence of anions. Then once

introducing phosphate along with the competing anion in equal parts (110 meq/L), the

removal of phosphate will be uninterrupted. For predictive capabilities, we can look at

Figure 23. When MM12 was placed in a phosphate solution of 3500 mg PO_4^{3-}-P/L (110

meq/L), its sorption capacity was ~28 mg PO_4^{3-}-P. On the other hand when MN was placed

in a phosphate solution of 3500 mg PO_4^{3-}-P/L (110 meq/L), its sorption capacity was ~10

mg PO_4^{3-}-P. If we consider MM12's removal to be the combination of electrostatic attraction

and selective recognition and MN's removal to be the combination of purely electrostatic

attraction, then the difference between the two polymers' sorption capacities will be the

sorption capacity of the selective recognition sites (~18 mg PO_4^{3-}-P). From this study, it

can be seen that after the non-specific binding sites were saturated, approximately 17 mg

PO_4^{3-}-P was removed from solution (**Figure 24**). This level of removal is extremely similar

to that of the 18 mg PO_4^{3-}-P sorption capacity predicted for the selective recognition sites. The ability for the MM12 to reach this P sorption capacity after continuous immersion in high concentrations of competing anions suggests that MM12 does in fact have imprinted recognition sites that selectively remove phosphate.
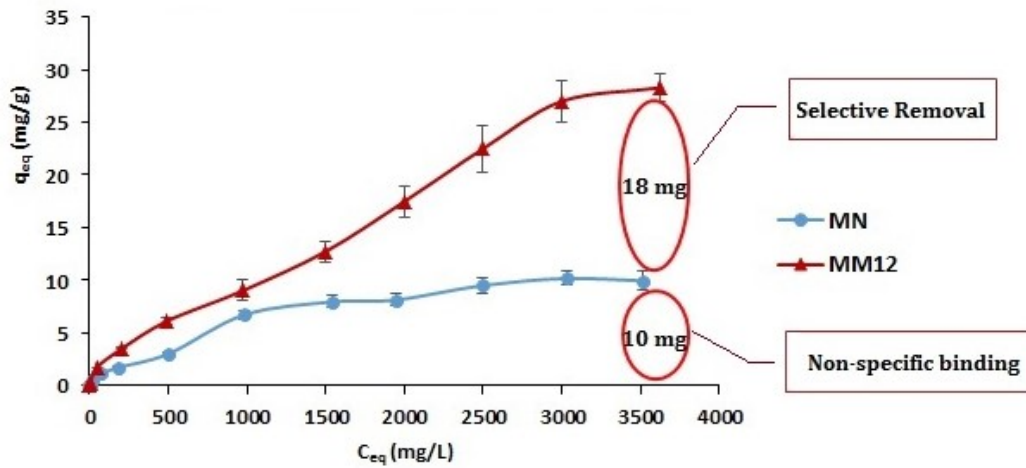
Figure 23. Comparison of Sorption Capacities between Non-specific Binding and Selective Removal
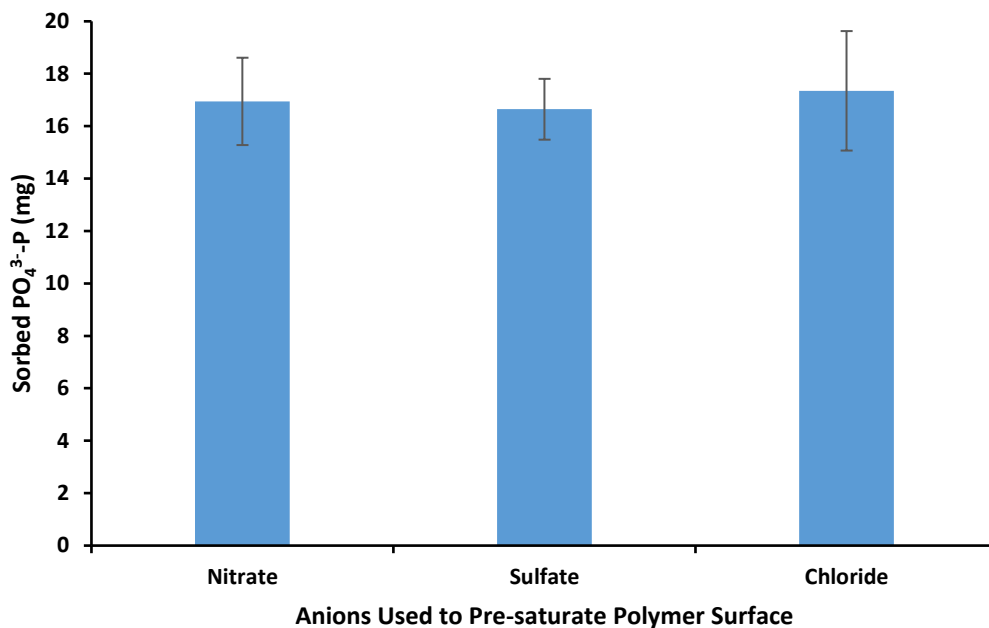
Figure 24. Phosphate Removal after Saturation of Electrostatic Attraction Sites

2.3.5.2.   Kinetic Study

Understanding the rate at which the sorbent removes phosphate from solution can

provide key insight into the removal mechanism. Therefore, kinetic studies were conducted

to understand the phosphate removal mechanism of MM12. Removal behavior using

several initial phosphate concentrations (0.1, 5, and 50 mg/L) were recorded (**Figure 25**).

Several pieces of information were discerned from the data shown. First, the reaction

kinetics was identical with all initial concentrations of phosphate. Next, the removal of

phosphate on the polymer surface occurred immediately. This immediate removal is likely

the cause of electrostatic interactions taking place nearly instantaneously (Adamson and

Gast, 1997). After the initial removal, a pseudo-equilibrium between sorption and

desorption of phosphate onto the polymer surface was reached. Because of the unchanging

nature of this pseudo-equilibrium, the data are reported up to 200 minutes (**Figure 25**)

(although studies were conducted up to 1440 minutes).

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It is common to reach pseudo-equilibrium with electrostatic interactions. Nevertheless, it was believed that the presence of selective binding (chemical binding) sites, this pseudo-equilibrium should be breached after adequate mechanical mixing of solution. The mechanical mixing provides means for phosphate ions to come in contact with the binding sites over time. Once the ions come in contact with these binding sites, desorption should not occur. Thus, the pseudo-equilibrium would be breached and the phosphate concentration would be slowly lowered until it reaches nearly zero or until the sorbent reaches its sorption capacity (**Figure 26**).
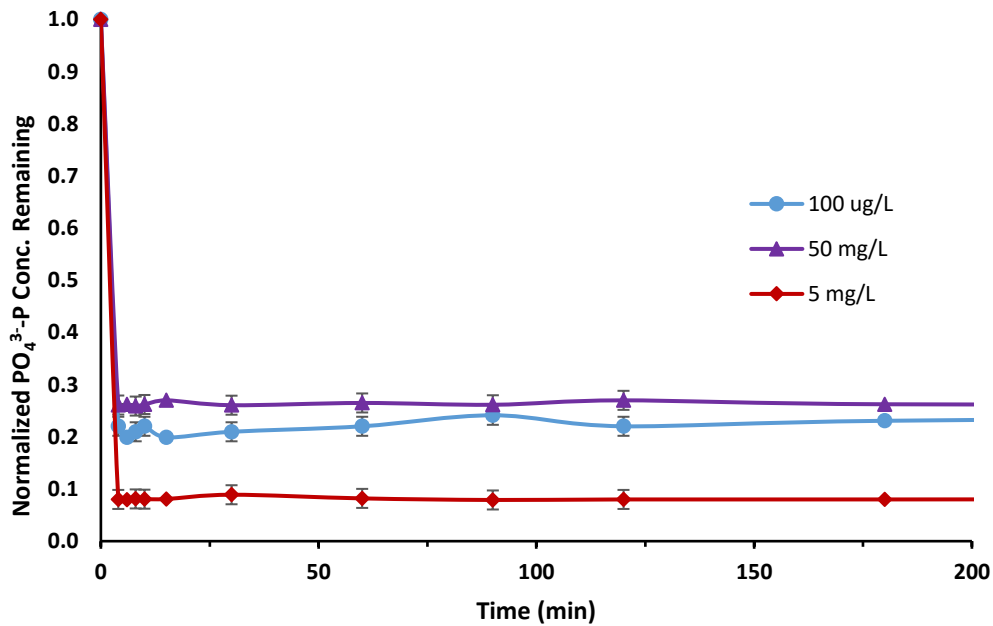
Figure 25. Assessment of MM12's Phosphate Removal Rate at Varying Initial Concentrations
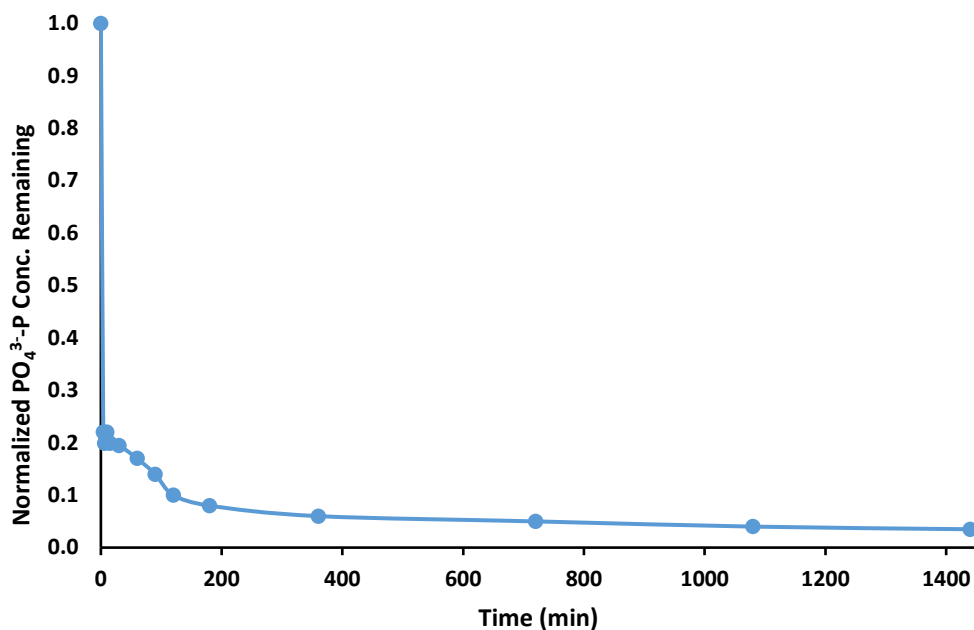
Figure 26. Expected Phosphate Removal Rate Behavior of MM12

With the observed kinetic behavior at hand, it appears that the recognition sites are not experiencing interactions with the phosphate ions. Further investigation on this behavior was thus conducted. The seemingly probable cause for this behavior was deemed to be that the phosphate ions were not reaching the imprinted recognition sites. The root of this issue is likely due to the formation of an electric double layer (**Figure 27**). When a charged surface is present in solution, ions of opposite charge will be attracted through electrostatic attraction. The attraction of anions to the surface of the positively charged polymer will generate a new charge on the surface of the polymer. This attraction of anions to the surface will continue to happen until the total net charge in the solution is balanced by the equal and opposite net charge on the surface. This then generates a double layer of charge, one localized on the surface of the plane and the other developed in the diffuse region extending into solution. The diffuse layer is the result of ions of one sign being held more strongly with those of the opposite sign. The pseudo-equilibrium that is developed

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over the diffuse layer is due to the osmotic pressure of solvent being reduced in the region between two planes. The two planes, in this case, are the surface of MM12 and the bulk solution. The equilibrium between these two "planes" can only be shifted by changing the external ionic strength in solution (Adamson and Gast, 1997). This helps to explain why imprinted recognition sites only appeared to take significant predominance at higher concentrations. As the phosphate concentration was increased, so was the ionic strength of the bulk solution. This increase in the ionic strength of the bulk solution likely grew so large that it diminished the equilibrium formed across the double layer and pushed phosphate anions past the surface of the polymer and into the polymer matrix. (Adamson and Gast, 1997) also reported that if specific chemical interactions are not dominant, the adsorption of an ionic species is largely determined by its charge (Adamson and Gast, 1997). Within that given charge type, the sequence of removal is likely that of increasing hydration enthalpy (Adamson and Gast, 1997). Based on MM12's removal kinetics witnessed at lower phosphate concentrations, it is believed that specific chemical interactions from the imprinted recognition sites are not the dominant mechanism for phosphate removal. In fact, it appears very likely that the dominant mechanism for phosphate removal is based on electrostatic attraction and is charge determinant.
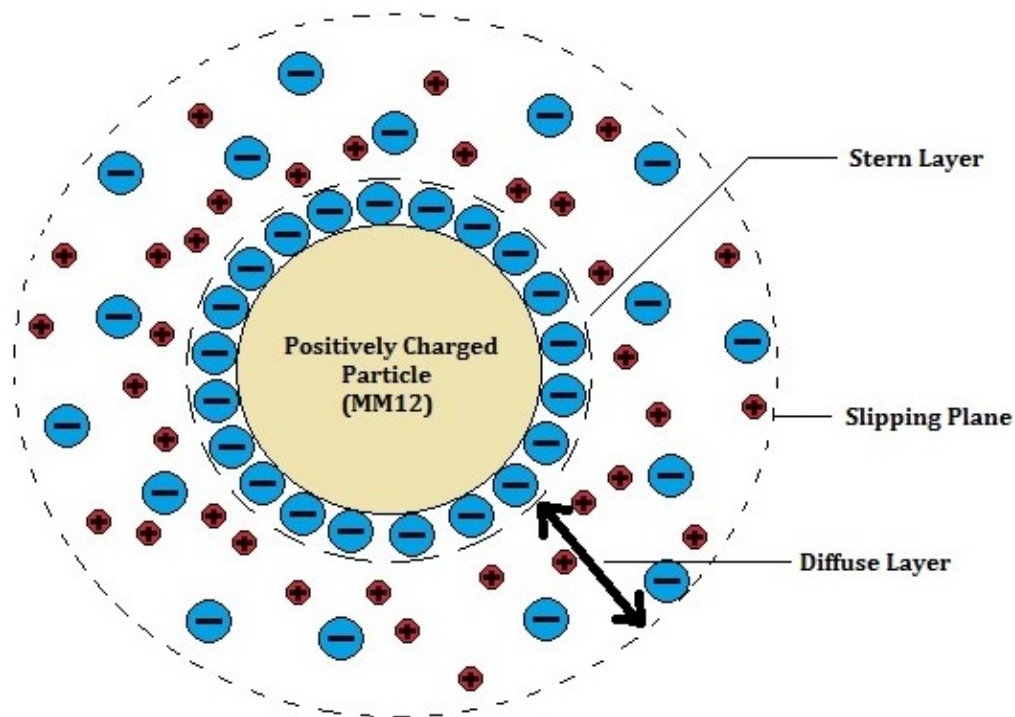
Figure 27. Hypothesized Electric Double Layer Phenomenon with MM12

2.4. Conclusions

Of the three MIPs initially selected for study, only one (METAC MIP) showed promise as a feasible P-sorbent. The METAC MIP's sorption capacity was found to be significantly increased by its template:monomer ratio; a ratio of 1:4 provided a sorption capacity of ~11 mg PO_4^{3-}-P while a ratio of 12:4 provided a sorption capacity of ~28 mg PO_4^{3-}-P under the same conditions. The MM12 experienced significant interference from competing anions, as evidence collected throughout the study suggests that electrostatic attraction of anions is the predominant mechanism for P-removal. However, FTIR analysis and a partial-selectivity study confirmed that selective chemical binding of phosphate is occurring in the imprinted recognition sites. Nevertheless, this selective binding is inhibited at lower phosphate concentrations of relevance due to the development of an electric double layer from the polymer's positive surface charge.

2.5.   Significance and Future Perspective

The work completed herein is the first work done indicating that molecularly imprinted polymers can attain sorption capacities significant enough to be deemed as an effective P-removal technology. Additionally, this study provided evidence that the molecular recognition sites generated from imprinting can bind phosphate selectively in the presence of competing anions. While the MIP studied within this report (MM12) has limitations which currently bar it from being applicable to P-recovery and reuse, it shows that molecular imprinting technology has potential to produce sorbents which can selectively recover P for reuse. It should be noted that this study focused only on the ability of MIPs to remove orthophosphate. It is believed that the MIPs can also show successful removal of polyphosphates and organophosphates due to the shared functionalities of these different phosphate forms. Specifically, organophosphates are expected to be sorbed quite effectively as the MIPs were imprinted with phenylphosphonic acid, which is an organophosphate itself.

To best further the work in this area, several items of recommendation are provided. First, the available pool of functional monomers that can be used to synthesize MIPs for phosphate recognition is rather small. Screening of additional functional monomers, which can be completed through methods such as computational modeling, is necessary to find the optimal monomer. Another approach is to neutralize the surface charge on the MIP. As shown from the study, polymers with positive surface charge (high PZC) suffer from significant interference with other anions. Selecting functional monomers, which form complexes with phenylphosphonic acid while having a lower PZC, could help negate the effect from non-specific surface binding in MIPs. Additionally, copolymerization

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with monomers that have negatively charged surfaces could produce a polymer with a neutral net charge to negate this interference and electric double layer formation as well. Lastly, optimization of synthesis conditions is necessary for developing the best MIP for this application. In this study, only one design parameter (template:monomer ratio) was altered. Optimizing design parameters can require tedious effort and large dedications of time. High throughput well-plate testing as identified by (Vasapollo et al., 2011) can be a practical means to higher efficiency design for developing MIPs that may work for P removal.

CHAPTER 3. SUMMARY OF WORK, CONCLUSIONS, AND FUTURE

PERSPECTIVES

3.1. Introduction

Society is trending toward more sustainable methods for phosphorus (P) production and application in order to reduce pressure on natural and finite sources of phosphate rock. Recovery of P from new sources has been identified as a pivotal point in moving toward this direction. The majority of P harvested from phosphate rock is applied in agricultural applications. Of this P applied in agriculture, much of the P is transported through surface runoff into surface waters. As surface waters continue to be polluted with otherwise valuable P, their viability as alternative sources to P-recovery continues to rise. In tandem to generating an alternative source for P-recovery, the health of socio-economically important lakes could be restored. Restoring the quality of lakes (i.e. reducing and preventing eutrophication) across the United States could prevent an annual economic loss of $2.2 billion. However, available technologies are not up to par to be implemented as effective vehicle for P-recovery and -reuse. Effective P-removal below environmentally relevant concentrations, selective removal of P in the presence of competing constituents, and the opportunity for recovery and reuse of P are critical attributes that many available technologies commonly lack. In the research presented in this thesis, molecularly imprinted polymers (MIPs) were identified as a potential vehicle for meeting all of the desired attributes for P-recovery and -reuse. Almost all works on MIP related to P have focused on phosphate recognition for sensing, not for targeted P recovery and reuse.

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3.2.   Evaluation of Several MIPs for P Removal

Three candidate functional monomers for MIP formulations were selected for this work based on extensive literature survey. Thiourea (TU), 1-Allyl-2-thiourea (AT), and 2-methacryloyloxyethyltrimethylammonium chloride (METAC) were the three functional monomers used here. TU showed no promise as a P-sorbent as it removed little to no phosphate across various P concentrations and pH. The AT MIP was found to have a P sorption capacity of only ~1 mg PO_4^{3-}-P/g. This polymer showed no promise as a competitive P-sorbent as it had a sorption capacity which was far lower than many other available technologies and did not resist interference from competing anions. The METAC MIP was the only polymer which showed potential as a P-sorbent with a competitive sorption capacity. The sorption capacity was largely affected by the template:monomer ratio. Ratios ranging from 0:4 to 12:4 lead to P sorption capacities of 10 mg PO_4^{3-}-P/g to 28 mg PO_4^{3-}-P/g, respectively. However, it was found that phosphate removal was also significantly reduced by the presence of coexisting anions.

3.3.   Potential for P Recovery/Reuse with METAC MIPs

The biodegradability and rate of phosphate desorption were the primary parameters tested to assess METAC MIP's (with template:monomer of 12:4, MM12) potential for recovery and reuse of P. Respirometer studies indicated that MM12 was an available food source for bacteria. MM12 can possibly be applied as a soil amendment as it will be biodegraded over time. Desorption studies indicated that MM12 releases a large bulk of loosely-bound phosphate immediately, but the majority of the bound phosphate is not released. It is believed that the phosphate not released immediately is strongly bound into the polymer matrix and cannot be accessed until those bonds are broken through

(bio)degradation processes. This form of phosphate release indicates that MM12 could provide some phosphate to plants for immediate nourishment, and then work as a slow-release fertilizer to provide subtle nourishment over time. Phosphate supplementation in this way can lead to more economically and environmentally favorable fertilizer-application.

3.4. P Removal Characteristics and Mechanism of Removal for METAC MIPs

METAC MIPs were shown to have partial-selectivity toward P removal. Evidence suggests that the MIPs have non-selective removal of anions due to non-specific binding sites on the surface of the polymer. These non-specific binding sites are likely due to the polymer's high PZC, giving it a positive surface charge at the pH of interest (7.5 - 8.5). The positive surface charge therein contributes to electrostatic attraction of all negatively charged species in water. Once all non-specific binding sites are saturated, phosphate was selectively removed by the imprinted recognition sites. Kinetic studies indicated that electrostatic attraction is the predominant mechanism of P removal over chemical binding by the imprinted recognition sites. In fact, evidence suggests that chemical binding will not occur unless adequate ionic strength is present in solution. The formation of an electric double layer on the surface of the charged polymer surface is hypothesized. This electric double layer engenders pseudo-equilibrium between the charged surface and the bulk solution in a "diffuse layer," preventing the accumulation of additional ions on the polymer surface.

3.5. Conclusions

This work has found that, of the functional monomers previously investigated for phosphate recognition (sensors) can also act as phosphate sorbents for recovery and reuse.

Of the three monomers tested, only METAC was shown to have potential viability as a selective P-sorbent when used in the synthesis of MIPs. The METAC MIPs showed partial selectivity as they also contained an abundance of non-specific binding sites due to their surface charge. METAC MIPs showed competitive sorption capacities when compared to available technologies. Also, METAC MIPs demonstrated characteristics which could lead to the successful implementation of recoverable/reusable sorbents. METAC MIPs are not feasible as selective P-sorbents from low ionic strength solutions like eutrophic surface waters and municipal wastewaters at this time due to their severe reduction in phosphate removal when in the presence of coexisting anions. However, they may be a feasible option as a selective P sorbent from high ionic strength solutions (i.e. powder coating industry and fertilizer plant effluents). Even without the presence of coexisting anions, their sorption capacities could be severely limited by the supposed formation of an electric double layer around the charged polymer surface. These results suggest that if better materials or design parameters are found, MIPs have promise to be implemented as sorbents for a more sustainable P-recovery and reuse cycle.

3.6.   Future Perspectives

Currently, there are far too few functional monomers which have been screened for phosphate recognition. Screening more functional monomers to develop a larger selection pool for study is a crucial first step in advancing the area of MIPs for P-recovery and -reuse. Another approach is to strengthen or at least optimize the MIPs that are currently available for P recovery. In this study it was revealed that non-specific binding from electrostatic attraction of phosphate eventually led to detrimental effects that include interference by coexisting anions and possible sorption capacity limitations in low ionic strength solutions.

The electrostatic attraction caused by the functional monomer's natural positive surface charge led to limitations in selective removal of P by MIPs. However, neutralizing this surface charge may be one of the options to negate the effect of electrostatic interactions. Selecting functional monomers which have a PZC below 7.5 (the lower pH range of typical eutrophic lakes) could result in MIPs which are void of non-specific electrostatic binding sites in the typical pH range. Another alternative is copolymerization with another monomer which can lead to neutrality on the MIP surface. Copolymerization with monomers of negative charge could result in charge-neutral polymers. The last option identified to advance this area is the optimization of MIPs through a better understanding of design parameters and their inherent influences. While template:monomer ratio was identified to be critical in influencing sorption capacity, there may be several other design parameters. High throughput well-plate testing can lead to a better understanding of design parameters and their influences in reasonable time and with reasonable efforts.

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APPENDIX

A.1. Thiourea (TU)

A.1.1. TU PZC

Table A.1. PZC Data for TU MIP

0.5 g/100 mL		1.0 g/100 mL		1.5 g/100 mL	
Volume Titrant (mL)	pH	Volume Titrant (mL)	pH	Volume Titrant (mL)	pH
0	11.32	0	11.6	0	11.65
0.2	11.24	0.25	11.51	0.5	11.49
0.4	11.16	0.5	11.41	0.75	11.4
0.6	11.05	0.75	11.28	1	11.27
0.8	10.91	1	11.09	1.25	11.1
1	10.73	1.25	10.81	1.5	10.86
1.2	10.47	1.5	10.23	1.75	10.41
1.4	10.05	1.75	6.1	1.85	10.04
1.5	9.59	2	3.94	1.95	9.08
1.6	7.34	2.25	3.47	2.05	6.46
1.7	6.19	2.5	3.25	2.15	5.28
1.8	5.02	2.75	3.1	2.25	4.58
1.9	4.06	3	2.99	2.35	4.17
2	3.75	3.25	2.9	2.45	3.89
2.1	3.57	3.5	2.84	2.7	3.5
2.2	3.44	3.75	2.8	2.95	3.29
2.3	3.35	4	2.76	3.2	3.14
2.5	3.2	4.25	2.72	3.45	3.04
2.7	3.09	4.5	2.69	3.7	2.95
2.9	3.01	4.75	2.66	3.95	2.88
3.1	2.94	5	2.64	4.2	2.84
3.3	2.89	5.25	2.61	4.45	2.8
3.5	2.84	5.5	2.59	4.7	2.76
3.7	2.8	5.75	2.57	4.95	2.72
3.9	2.76	6	2.55	5.2	2.69
4.1	2.72	6.25	2.53	5.45	2.66
4.3	2.69	6.5	2.51	5.7	2.64
4.5	2.66	6.75	2.5	5.95	2.61

Table A.1. PZC Data for TU MIP (Continued)

Volume Titrant (mL)	pH	Volume Titrant (mL)	pH	Volume Titrant (mL)	pH
4.9	2.61	7.25	2.47	6.45	2.57
5.1	2.59	-	-	6.7	2.55
5.3	2.57	-	-	6.95	2.53
5.5	2.55	-	-	7.2	2.51
5.7	2.53	-	-	7.45	2.5
5.9	2.51	-	-	7.7	2.48
6.1	2.5	-	-	7.95	2.47
6.3	2.48	-	-	-	-
6.5	2.47	-	-	-	-

A.1.2. TU pH

Table A.2. P Removal for TU MIP over Varying pH (C_0 = 5 mg PO_4^{3-}-P/L)

pH	Normalized	Normalized StdDev
3	0.000	0.017
4	0.000	0.019
5	0.008	0.020
6	0.000	0.005
7	0.000	0.012
8	0.005	0.008
9	0.000	0.008
10	0.008	0.005
11	0.005	0.012
12	0.000	0.016

A1.3. TU Sorption Capacity

Table A.3. TU NIP Sorption Capacity Data

Experiment Title	Actual Initial Conc. (mg/L)	Sorbed Phosphate (mg/L)	Standard Deviation	Remaining P (mg/L)	Sorbed P (mg/g)	StdDev (mg/g)
0	0.00	0.00	0.00	0.00	0.00	0.00
1 mg/L	0.95	0.00	0.00	0.95	0.00	0.00
5 mg/L	5.10	0.00	0.01	5.10	0.00	0.00
50 mg/L	45.00	0.00	0.09	45.00	0.00	0.00

Table A.4. TU MIP Sorption Capacity Data

Experiment Title	Actual Initial Conc. (mg/L)	Sorbed Phosphate (mg/L)	Standard Deviation	Remaining P (mg/L)	Sorbed P (mg/g)	StdDev (mg/g)
0	0.00	0.00	0.00	0.00	0.00	0.00
1 mg/L	0.90	0.00	0.02	0.90	0.00	0.00
5 mg/L	4.60	0.02	0.00	4.57	0.00	0.00
50 mg/L	47.21	0.11	0.43	47.09	0.01	0.02

A.2. 1-Allyl-2-thiourea (AT)

A.2.1. AT PZC

Table A.5. PZC Data for AT MIP

0.5 g/100 mL		1.0 g/100 mL		1.5 g/100 mL	
Volume Titrant (mL)	pH	Volume Titrant (mL)	pH	Volume Titrant (mL)	pH
0	11.63	0	11.58	0	11.49
0.25	11.53	0.25	11.42	0.25	11.22
0.5	11.32	0.5	11.2	0.5	10.89
0.75	10.94	0.75	10.8	0.75	10.2
1	8.87	1	8.85	1	8.9
1.1	6.85	1.1	6.64	1.1	6.22
1.2	4.61	1.2	4.88	1.2	4.65

Table A.5. PZC Data for AT MIP (Continued)

Volume Titrant (mL)	pH	Volume Titrant (mL)	pH	Volume Titrant (mL)	pH
1.3	3.99	1.3	4.1	1.3	4.06
1.55	3.5	1.55	3.57	1.55	3.56
1.8	3.27	1.8	3.33	1.8	3.32
2.05	3.13	2.05	3.18	2.05	3.17
2.3	3.02	2.3	3.07	2.3	3.06
2.55	2.94	2.55	2.98	2.55	2.97
3.05	2.81	3.05	2.85	3.05	2.84
3.55	2.71	3.55	2.75	3.55	2.74
4.05	2.64	4.05	2.68	4.05	2.67

A.2.2. AT pH Study

Table A.6. P Removal for AT MIP over Varying pH (C_0 = 5 mg PO_4^{3-}-P/L)

pH	Normalized	Normalized StdDev
3	0.213	0.088
4	0.190	0.055
5	0.244	0.037
6	0.358	0.013
7	0.366	0.020
8	0.357	0.020
9	0.292	0.018
10	0.293	0.019
11	0.274	0.011
12	0.277	0.024

A.2.3. AT Sorption Capacity

Table A.7. AT MIP Sorption Capacity Data

Experiment Title	Actual Initial Conc. (mg/L)	Sorbed Phosphate (mg/L)	Standard Deviation	Remaining P (mg/L)	Sorbed P (mg/g)	StdDev (mg/g)
0	0.00	0.00	0.00	0.00	0.00	0.00
1 mg/L	1.21	0.77	0.05	0.44	0.04	0.00
5 mg/L	4.48	1.53	0.09	2.95	0.08	0.00
20 mg/L	18.48	4.64	0.11	13.83	0.23	0.01
100 mg/L	101.08	10.52	1.06	90.56	0.53	0.05
200 mg/L	192.79	18.22	7.43	174.57	0.91	0.37
500 mg/L	503.07	19.13	8.04	483.95	0.96	0.4
1000 mg/L	999.5	18.22	5.9	981.28	0.91	0.3

Table A.8. AT NIP Sorption Capacity Data

Experiment Title	Actual Initial Conc. (mg/L)	Sorbed Phosphate (mg/L)	Standard Deviation	Remaining P (mg/L)	Sorbed P (mg/g)	StdDev (mg/g)
0	0.00	0.00	0.00	0.00	0.00	0.00
1 mg/L	0.42	0.01	0.02	0.41	0.00	0.00
5 mg/L	4.38	0.16	0.17	4.21	0.01	0.01
20 mg/L	19.39	0.94	0.03	18.45	0.05	0.00
100 mg/L	68.46	4.26	1.36	64.19	0.21	0.07
200 mg/L	184.24	5.43	2.31	178.81	0.27	0.12
500 mg/L	496.95	6.33	4.61	490.61	0.32	0.23
1000 mg/L	988.19	6.33	3.39	981.86	0.32	0.17

A.2.4. AT Interference Study

Table A.9. P Removal by AT MIP amongst Competing Anions (C₀ = 5 mg PO₄³⁻-P/L)

Cl-								
Cl⁻ (50 mg/L)			Cl⁻ (100 mg/L)			Cl⁻ (500 mg/L)		
Time (hr)	Phosphate conc. (mg/L)	standard deviation	Time (hr)	Phosphate conc. (mg/L)	standard deviation	Time (hr)	Phosphate conc. (mg/L)	standard deviation
0 h	4.44	0.00	0 h	4.42	0.02	0 h	4.49	0.04
24 h	3.69	0.03	24 h	3.89	0.02	24 h	4.21	0.06
Blank	4.42	0.03	Blank	4.41	0.03	Blank	4.49	0.02
Amount Sorbed (mg/L)	Percent Sorption		Amount Sorbed (mg/L)	Percent Sorption		Amount Sorbed (mg/L)	Percent Sorption	
0.74	16.78		0.53	12.01		0.28	6.22	
Normalized Sorption	0.168	0.006	Normalized Sorption	0.120	0.004	Normalized Sorption	0.062	0.013

Table A.9. P Removal by AT MIP amongst Competing Anions (C₀ = 5 mg PO₄³⁻-P/L) (Continued)

HCO₃⁻								
HCO₃⁻ (5 mg/L)			**HCO₃⁻ (10 mg/L)**			**HCO₃⁻ (50 mg/L)**		
Time (hr)	**Phosphate conc. (mg/L)**	**standard deviation**	**Time (hr)**	**Phosphate conc. (mg/L)**	**standard deviation**	**Time (hr)**	**Phosphate conc. (mg/L)**	**standard deviation**
0 h	4.35	0.02	0 h	4.39	0.02	0 h	4.41	0.02
24 h	2.04	0.04	24 h	2.04	0.02	24 h	2.12	0.03
Blank	4.37	0.02	Blank	4.34	0.07	Blank	4.39	0.03
Amount Sorbed (mg/L)	**Percent Sorption**		**Amount Sorbed (mg/L)**	**Percent Sorption**		**Amount Sorbed (mg/L)**	**Percent Sorption**	
2.31	53.09		2.34	53.45		2.29	51.91	
Normalized Sorption	0.531	0.010	**Normalized Sorption**	0.534	0.004	**Normalized Sorption**	0.519	0.006

Table A.9. P Removal by AT MIP amongst Competing Anions (C_0 = 5 mg PO_4^{3-}-P/L) (Continued)

SO_4^{2-}								
SO_4^{2-} (50 mg/L)			SO_4^{2-} (100 mg/L)			SO_4^{2-} (1000 mg/L)		
Time (hr)	Phosphate conc. (mg/L)	standard deviation	Time (hr)	Phosphate conc. (mg/L)	standard deviation	Time (hr)	Phosphate conc. (mg/L)	standard deviation
0 h	4.45	0.02	0 h	4.53	0.02	0 h	4.45	0.04
24 h	3.97	0.07	24 h	4.15	0.07	24 h	4.22	0.06
Blank	4.39	0.04	Blank	4.52	0.03	Blank	4.42	0.03
Amount Sorbed (mg/L)	Percent Sorption		Amount Sorbed (mg/L)	Percent Sorption		Amount Sorbed (mg/L)	Percent Sorption	
0.49	10.91		0.38	8.47		0.23	5.07	
Normalized Sorption	0.109	0.016	Normalized Sorption	0.085	0.015	Normalized Sorption	0.051	0.013
Humic Acid								
Humic Acid (2 mg/L)			Humic Acid (5 mg/L)			Humic Acid (10 mg/L)		
Time (hr)	Phosphate conc. (mg/L)	standard deviation	Time (hr)	Phosphate conc. (mg/L)	standard deviation	Time (hr)	Phosphate conc. (mg/L)	standard deviation
0 h	4.70	0.04	0 h	4.76	0.02	0 h	4.76	0.07
24 h	2.04	0.10	24 h	2.20	0.07	24 h	2.11	0.09
Blank	4.70	0.02	Blank	4.77	0.07	Blank	4.73	0.02
Amount Sorbed (mg/L)	Percent Sorption		Amount Sorbed (mg/L)	Percent Sorption		Amount Sorbed (mg/L)	Percent Sorption	
2.66	56.69		2.55	53.64		2.64	55.54	
Normalized Sorption	0.567	0.021	Normalized Sorption	0.536	0.015	Normalized Sorption	0.555	0.019

Table A.9. P Removal by AT MIP amongst Competing Anions (C_0 = 5 mg PO_4^{3-}-P/L) (Continued)

NOM								
NOM (5 mg/L)			**NOM (10 mg/L)**			**NOM (50 mg/L)**		
Time (hr)	**Phosphate conc. (mg/L)**	**standard deviation**	**Time (hr)**	**Phosphate conc. (mg/L)**	**standard deviation**	**Time (hr)**	**Phosphate conc. (mg/L)**	**standard deviation**
0 h	4.99	0.02	0 h	4.89	0.04	0 h	4.84	0.03
24 h	2.55	0.18	24 h	2.82	0.02	24 h	2.69	0.09
Blank	4.86	0.06	Blank	4.94	0.06	Blank	4.85	0.04
Amount Sorbed (mg/L)	**Percent Sorption**		**Amount Sorbed (mg/L)**	**Percent Sorption**		**Amount Sorbed (mg/L)**	**Percent Sorption**	
2.45	48.98		2.07	42.40		2.14	44.29	
Normalized Sorption	0.490	0.036	**Normalized Sorption**	0.424	0.003	**Normalized Sorption**	0.443	0.018

Control		
Control (5 mg/L)		
Time (hr)	**Phosphate conc. (mg/L)**	**standard deviation**
0 h	4.48	0.23
24 h	2.05	0.03
Blank	4.55	0.09
Amount Sorbed (mg/L)	**Percent Sorption**	
2.42	54.14	
Normalized Sorption	0.541	0.007

A.2.5. AT Statistical Analysis

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One-way ANOVA: % Removal versus pH

```
Method

Null hypothesis         All means are equal
Alternative hypothesis  At least one mean is different
Significance level      α = 0.05

Equal variances were assumed for the analysis.


Factor Information

Factor  Levels  Values
pH          10  10, 11, 12, 3, 4, 5, 6, 7, 8, 9


Analysis of Variance

Source  DF  Adj SS  Adj MS  F-Value  P-Value
pH       9   999.3  111.03     7.69    0.000
Error   20   288.9   14.45
Total   29  1288.2


Model Summary

      S    R-sq  R-sq(adj)  R-sq(pred)
3.80088  77.57%     67.48%      49.53%


Means

pH  N    Mean  StDev        95% CI
10  3   29.27   1.92  ( 24.69,  33.85)
11  3  27.438  1.143  (22.861, 32.016)
12  3   27.69   2.41  ( 23.11,  32.27)
3   3   21.31   8.76  ( 16.73,  25.88)
4   3   19.00   5.50  ( 14.43,  23.58)
5   3   24.45   3.73  ( 19.87,  29.03)
6   3  35.849  1.303  (31.271, 40.426)
7   3   36.60   1.99  ( 32.02,  41.18)
8   3   35.74   1.97  ( 31.16,  40.32)
9   3   29.24   1.78  ( 24.66,  33.82)

Pooled StDev = 3.80088
```

Tukey Pairwise Comparisons

```
Grouping Information Using the Tukey Method and 95% Confidence
```

```
pH  N    Mean  Grouping
7   3   36.60  A
6   3   35.849 A
8   3   35.74  A
10  3   29.27  A B
9   3   29.24  A B
12  3   27.69  A B
11  3   27.438 A B
5   3   24.45    B
3   3   21.31    B
4   3   19.00    B
```

Means that do not share a letter are significantly different.

Tukey Simultaneous 95% CIs

Interval Plot of % Removal vs pH

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One-way ANOVA: Sorption (mg) versus Polymer

```
Method

Null hypothesis         All means are equal
Alternative hypothesis  At least one mean is different
Significance level      α = 0.05

Equal variances were assumed for the analysis.


Factor Information

Factor   Levels  Values
Polymer       8  100 MIP, 100 NIP, 20 MIP, 20 NIP, 200 MIP, 200 NIP, 5 MIP, 5 NIP


Analysis of Variance

Source   DF  Adj SS   Adj MS  F-Value  P-Value
Polymer   7  2.4226  0.34608    11.63    0.000
Error    16  0.4763  0.02977
Total    23  2.8989


Model Summary

       S    R-sq  R-sq(adj)  R-sq(pred)
0.172539  83.57%     76.38%      63.03%
```

```
Means


Polymer  N    Mean    StDev         95% CI
100 MIP  3   0.4861   0.0650  (  0.2749,  0.6973)
100 NIP  3   0.2319   0.0832  (  0.0207,  0.4430)
20 MIP   3  0.23218  0.00686  ( 0.02101, 0.44336)
20 NIP   3  0.04450  0.00203  (-0.16667, 0.25568)
200 MIP  3    0.934    0.455  (   0.722,   1.145)
200 NIP  3   0.7215   0.1412  (  0.5103,  0.9326)
5 MIP    3  0.08006  0.00524  (-0.13111, 0.29124)
5 NIP    3  0.00820  0.00884  (-0.20298, 0.21937)


Pooled StDev = 0.172539
```

Tukey Pairwise Comparisons

```
Grouping Information Using the Tukey Method and 95% Confidence

Polymer  N    Mean  Grouping
200 MIP  3   0.934  A
200 NIP  3  0.7215  A
100 MIP  3  0.4861  A B
20 MIP   3 0.23218    B
100 NIP  3  0.2319    B
5 MIP    3 0.08006    B
20 NIP   3 0.04450    B
5 NIP    3 0.00820    B


Means that do not share a letter are significantly different.
```

Tukey Simultaneous 95% CIs

Interval Plot of Sorption (mg) vs Polymer

One-way ANOVA: Sorption (mg) versus Polymer

```
* NOTE * Cannot draw the interval plot for the Tukey procedure. Interval plots for
         comparisons are illegible with more than 45 intervals.


Method

Null hypothesis         All means are equal
Alternative hypothesis  At least one mean is different
Significance level      α = 0.05

Equal variances were assumed for the analysis.


Factor Information

Factor   Levels  Values
Polymer      12  100 MIP, 100 NIP, 1000 MIP, 1000 NIP, 20 MIP, 20 NIP, 200 MIP, 200
NIP, 5
                 MIP, 5 NIP, 500 MIP, 500 NIP
```

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```
Analysis of Variance

Source   DF  Adj SS   Adj MS  F-Value  P-Value
Polymer  11   3.983  0.36205     6.23    0.000
Error    24   1.395  0.05812
Total    35   5.377


Model Summary

       S    R-sq  R-sq(adj)  R-sq(pred)
0.241083  74.06%     62.17%      41.64%


Means

Polymer   N     Mean    StDev          95% CI
100 MIP   3   0.4861   0.0650  (  0.1988,  0.7734)
100 NIP   3   0.2319   0.0832  ( -0.0554,  0.5191)
1000 MIP  3    1.002    0.361  (   0.715,   1.289)
1000 NIP  3    0.452    0.207  (   0.165,   0.740)
20 MIP    3  0.23218  0.00686  (-0.05509, 0.51945)
20 NIP    3  0.04450  0.00203  (-0.24277, 0.33177)
200 MIP   3    0.934    0.455  (   0.646,   1.221)
200 NIP   3   0.7215   0.1412  (  0.4342,  1.0087)
5 MIP     3  0.08006  0.00524  (-0.20721, 0.36734)
5 NIP     3  0.00820  0.00884  (-0.27908, 0.29547)
500 MIP   3    0.729    0.493  (   0.441,   1.016)
500 NIP   3    0.226    0.207  (  -0.061,   0.513)


Pooled StDev = 0.241083
```

Tukey Pairwise Comparisons

```
Grouping Information Using the Tukey Method and 95% Confidence

Polymer   N     Mean  Grouping
1000 MIP  3    1.002  A
200 MIP   3    0.934  A B
500 MIP   3    0.729  A B C
200 NIP   3   0.7215  A B C
100 MIP   3   0.4861  A B C D
1000 NIP  3    0.452  A B C D
20 MIP    3  0.23218    B C D
100 NIP   3   0.2319    B C D
500 NIP   3    0.226    B C D
5 MIP     3  0.08006      C D
20 NIP    3  0.04450      C D
5 NIP     3  0.00820        D

Means that do not share a letter are significantly different.
```

Interval Plot of Sorption (mg) vs Polymer

————— 7/4/2017 12:43:37 PM —————————————————————

One-way ANOVA: Normalized Removal versus Interference

```
* NOTE * Cannot draw the interval plot for the Tukey procedure. Interval plots for
         comparisons are illegible with more than 45 intervals.
```

Method

```
Null hypothesis        All means are equal
Alternative hypothesis  At least one mean is different
Significance level     α = 0.05
```

Equal variances were assumed for the analysis.

Factor Information

```
Factor        Levels  Values
Interference      19  Cl 100, Cl 50, Cl 500, Control, HA 10, HA 2, HA 5, HCO3 10, HCO3
5,
                      HCO3 50, NO3 10, NO3 100, NO3 50, NOM 10, NOM 5, NOM 50, SO4
100, SO4
                      1000, SO4 50
```

Analysis of Variance

Source	DF	Adj SS	Adj MS	F-Value	P-Value
Interference	18	2.34609	0.130338	355.87	0.000
Error	38	0.01392	0.000366		
Total	56	2.36000			

Model Summary

S	R-sq	R-sq(adj)	R-sq(pred)
0.0191377	99.41%	99.13%	98.67%

Means

Interference	N	Mean	StDev	95% CI
Cl 100	3	0.12011	0.00443	(0.09774, 0.14248)
Cl 50	3	0.16781	0.00763	(0.14544, 0.19018)
Cl 500	3	0.06220	0.01554	(0.03984, 0.08457)
Control	3	0.54137	0.00885	(0.51900, 0.56374)
HA 10	3	0.5554	0.0229	(0.5331, 0.5778)
HA 2	3	0.5669	0.0260	(0.5445, 0.5893)
HA 5	3	0.5364	0.0188	(0.5141, 0.5588)
HCO3 10	3	0.53449	0.00441	(0.51212, 0.55685)
HCO3 5	3	0.53091	0.01175	(0.50854, 0.55327)
HCO3 50	3	0.51912	0.00760	(0.49675, 0.54149)
NO3 10	3	0.23754	0.01612	(0.21517, 0.25991)
NO3 100	3	0.0867	0.0269	(0.0643, 0.1090)
NO3 50	3	0.1687	0.0196	(0.1463, 0.1911)
NOM 10	3	0.42396	0.00399	(0.40160, 0.44633)
NOM 5	3	0.4898	0.0435	(0.4675, 0.5122)

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```
NOM 50          3   0.4429   0.0225  ( 0.4205,  0.4653)
SO4 100         3   0.0847   0.0188  ( 0.0624,  0.1071)
SO4 1000        3  0.05072  0.01584  (0.02836, 0.07309)
SO4 50          3   0.1091   0.0201  ( 0.0867,  0.1314)

Pooled StDev = 0.0191377
```

Tukey Pairwise Comparisons

```
Grouping Information Using the Tukey Method and 95% Confidence

Interference   N     Mean  Grouping
HA 2           3   0.5669  A
HA 10          3   0.5554  A
Control        3  0.54137  A B
HA 5           3   0.5364  A B
HCO3 10        3  0.53449  A B
HCO3 5         3  0.53091  A B
HCO3 50        3  0.51912  A B
NOM 5          3   0.4898    B C
NOM 50         3   0.4429      C D
NOM 10         3  0.42396        D
NO3 10         3  0.23754          E
NO3 50         3   0.1687            F
Cl 50          3  0.16781           F G
Cl 100         3  0.12011           F G H
SO4 50         3   0.1091             G H I
NO3 100        3   0.0867               H I
SO4 100        3   0.0847               H I
Cl 500         3  0.06220              H I
SO4 1000       3  0.05072                 I

Means that do not share a letter are significantly different.
```

Interval Plot of Normalized Removal vs Interference

A.3. 2-Methacryloyloxyethyltrimethylammonium Chloride (METAC)

A.3.1. METAC MIP PZC

Table A.10. PZC Data for METAC MIP

0.5 g/100 mL		1.0 g/100 mL		1.5 g/100 mL	
Volume Titrant (mL)	pH	Volume Titrant (mL)	pH	Volume Titrant (mL)	pH
0	11.49	0	11.29	0	11.32
0.5	11.27	0.5	10.99	0.2	11.24
1	10.89	1	10.88	0.4	11.16
1.1	8.69	1.25	7.81	0.6	11.05
1.2	6.48	1.5	5.59	0.8	10.91
1.3	5.75	1.6	5.29	1	10.73
1.4	5.21	1.7	5.06	1.2	10.47
1.5	4.81	1.8	4.88	1.4	10.05
1.75	4.21	1.9	4.7	1.5	9.59
2	3.79	2	4.55	1.6	7.34
2.25	3.5	2.1	4.42	1.7	6.19
2.75	3.16	2.2	4.3	1.8	5.02
3.25	2.97	2.3	4.18	1.9	4.06
3.75	2.84	2.4	4.06	2	3.75
-	-	2.5	3.95	2.1	3.57
-	-	2.6	3.58	2.2	3.44
-	-	2.7	3.5	2.3	3.35
-	-	2.8	3.48	2.5	3.2
-	-	2.9	3.45	2.7	3.09
-	-	3	3.41	2.9	3.01
-	-	3.5	2.95	3.1	2.94
-	-	-	-	3.3	2.89
-	-	-	-	3.5	2.84
-	-	-	-	3.7	2.8

A.3.2. METAC pH Study

Table A.11. P Removal for METAC MIP Over Varying pH (C_0 = 5 mg PO_4^{3-}-P/L)

pH	Normalized	Normalized StdDev
3	0.813	0.008
4	0.919	0.008
5	0.935	0.009
6	0.932	0.005
7	0.924	0.009
8	0.919	0.005
9	0.923	0.005
10	0.945	0.008
11	0.951	0.004
12	0.951	0.009

A.3.3. METAC Sorption Capacities (MN, MM, MM4, and MM12)

Table A.12. MN Sorption Capacity Data

Experiment Title	Actual Initial Conc. (mg/L)	Sorbed Phosphate (mg/L)	Standard Deviation	Remaining P (mg/L)	Sorbed P (mg/g)	StdDev (mg/g)
0	0.00	0.00	0.00	0.00	0.00	0.00
0.5 mg/L	0.57	0.26	0.02	0.31	0.01	0.00
5 mg/L	4.00	2.18	0.15	1.82	0.11	0.01
20 mg/L	18.22	10.28	0.29	7.94	0.51	0.01
100 mg/L	72.42	22.58	3.95	49.84	1.13	0.20
200 mg/L	185.15	35.00	1.72	150.15	1.75	0.09
500 mg/L	500.71	60.61	3.44	440.10	3.03	0.17
1000 mg/L	985.63	135.00	7.90	850.62	6.75	0.40
1500 mg/L	1545.68	159.35	10.96	1386.33	7.97	0.55
2000 mg/L	1944.05	162.93	10.96	1781.12	8.15	0.55
2500 mg/L	2494.52	190.23	14.50	2304.28	9.51	0.73
3000 mg/L	3027.17	203.66	13.80	2823.51	10.18	0.69
3500 mg/L	3512.82	199.19	17.62	3313.64	9.96	0.88

Table A.13. MM Sorption Capacity Data

Experiment Title	Actual Initial Conc. (mg/L)	Sorbed Phosphate (mg/L)	Standard Deviation	Remaining P (mg/L)	Sorbed P (mg/g)	StdDev (mg/g)
0	0.00	0.00	0.00	0.00	0.00	0.00
0.5 mg/L	0.51	0.50	0.01	0.01	0.03	0.00
2.5 mg/L	2.62	2.47	0.09	0.14	0.12	0.00
5 mg/L	5.99	5.46	0.17	0.54	0.27	0.01
15 mg/L	14.62	12.83	0.12	1.78	0.64	0.01
100 mg/L	95.46	43.86	1.58	51.60	2.19	0.08
300 mg/L	339.23	70.72	3.90	268.51	3.54	0.19
500 mg/L	472.96	83.87	4.84	389.09	4.19	0.24
1000 mg/L	997.33	137.85	3.44	859.48	6.89	0.17
1500 mg/L	1485.34	171.86	6.75	1313.48	8.59	0.34
2000 mg/L	1965.08	183.81	9.10	1781.27	9.19	0.45
2500 mg/L	2528.36	234.19	14.15	2294.16	11.71	0.71
3000 mg/L	2888.83	220.42	11.25	2668.41	11.02	0.56
3500 mg/L	3517.93	220.42	18.08	3297.52	11.02	0.90

Table A.14. MM4 Sorption Capacity Data

Experiment Title	Actual Initial Conc. (mg/L)	Sorbed Phosphate (mg/L)	Standard Deviation	Remaining P (mg/L)	Sorbed P (mg/g)	StdDev (mg/g)
0	0.00	0.00	0.00	0.00	0.00	0.00
0.5 mg/L	0.42	0.37	0.01	0.05	0.02	0.00
5 mg/L	4.40	4.13	0.03	0.27	0.21	0.00
50 mg/L	49.47	35.43	0.16	14.04	1.77	0.01
200 mg/L	201.58	70.63	1.45	130.95	3.53	0.07
500 mg/L	494.25	120.48	4.52	373.77	6.02	0.23
1000 mg/L	984.49	198.03	9.05	786.46	9.90	0.45
1500 mg/L	1458.31	248.56	10.80	1209.75	12.43	0.54
2000 mg/L	1990.12	303.42	44.60	1686.71	15.17	2.23
2500 mg/L	2540.42	322.45	44.73	2217.97	16.12	2.24
3000 mg/L	3080.08	356.04	38.78	2724.04	17.80	1.94
3500 mg/L	3449.55	369.47	26.87	3080.08	18.47	1.34

Table A.15. MM12 Sorption Capacity Data

Experiment Title	Actual Initial Conc. (mg/L)	Sorbed Phosphate (mg/L)	Standard Deviation	Remaining P (mg/L)	Sorbed P (mg/g)	StdDev (mg/g)
0.00	0.00	0.00	0.00	0.00	0.00	0.00
0.5 mg/L	0.33	0.30	0.03	0.04	0.01	0.00
5 mg/L	4.28	4.02	0.00	0.26	0.20	0.00
50 mg/L	46.21	34.58	0.92	11.63	1.73	0.05
200 mg/L	203.99	70.33	1.47	133.66	3.52	0.07
500 mg/L	491.96	122.85	6.99	369.11	6.14	0.35
1000 mg/L	976.41	180.99	19.80	795.42	9.05	0.99
1500 mg/L	1514.36	255.26	21.20	1259.10	12.76	1.06
2000 mg/L	2079.72	351.30	29.80	1728.42	17.57	1.49
2500 mg/L	2416.33	457.04	45.07	1959.29	22.85	2.25
3000 mg/L	3099.18	539.89	38.21	2559.29	26.99	1.91
3500 mg/L	3619.34	565.56	26.72	3053.79	28.28	1.34

A.3.4. METAC Interference Study (MM, MM4, and MM12)

Table A.16. P Removal by MM Amongst Competing Anions (C_0 = 5 mg PO_4^{3-}-P/L)

	Cl-								
	Cl- (50 mg/L)				**Cl- (100 mg/L)**			**Cl- (500 mg/L)**	
Time (hr)	**Phosphate conc. (mg/L)**	**standard deviation**	**Time (hr)**	**Phosphate conc. (mg/L)**	**standard deviation**	**Time (hr)**	**Phosphate conc. (mg/L)**	**standard deviation**
0 h	4.46	0.04	0 h	4.47	0.03	0 h	4.13	0.05
24 h	2.01	0.04	24 h	2.46	0.12	24 h	3.36	0.05
Blank	4.49	0.02	Blank	4.45	0.03	Blank	4.08	0.08
Amount Sorbed (mg/L)	**Percent Sorption**		**Amount Sorbed (mg/L)**	**Percent Sorption**		**Amount Sorbed (mg/L)**	**Percent Sorption**	
2.45	54.94		2.01	45.05		0.77	18.65	
Normalized Sorption	0.549	0.009	**Normalized Sorption**	0.450	0.027	**Normalized Sorption**	0.187	0.013

Table A.16. P Removal by MM Amongst Competing Anions (C_0 = 5 mg PO_4^{3-}-P/L) (Continued)

SO$_4^{2-}$								
SO$_4^{2-}$ (50 mg/L)			SO$_4^{2-}$ (100 mg/L)			SO$_4^{2-}$ (1000 mg/L)		
Time (hr)	Phosphate conc. (mg/L)	standard deviation	Time (hr)	Phosphate conc. (mg/L)	standard deviation	Time (hr)	Phosphate conc. (mg/L)	standard deviation
0 h	4.32	0.02	0 h	4.29	0.04	0 h	4.35	0.02
24 h	1.09	0.07	24 h	2.03	0.15	24 h	3.73	0.04
Blank	4.30	0.00	Blank	4.33	0.02	Blank	4.35	0.04
Amount Sorbed (mg/L)	Percent Sorption		Amount Sorbed (mg/L)	Percent Sorption		Amount Sorbed (mg/L)	Percent Sorption	
3.22	74.69		2.27	52.82		0.63	14.38	
Normalized Sorption	0.747	0.017	Normalized Sorption	0.528	0.035	Normalized Sorption	0.144	0.010

Table A.16. P Removal by MM Amongst Competing Anions (C_0 = 5 mg PO_4^{3-}-P/L) (Continued)

	NO$_3^-$							
	NO$_3^-$ (10 mg/L)			NO$_3^-$ (50 mg/L)			NO$_3^-$ (100 mg/L)	
Time (hr)	**Phosphate conc. (mg/L)**	**standard deviation**	**Time (hr)**	**Phosphate conc. (mg/L)**	**standard deviation**	**Time (hr)**	**Phosphate conc. (mg/L)**	**standard deviation**
0 h	4.32	0.02	0 h	4.35	0.03	0 h	4.36	0.02
24 h	0.41	0.03	24 h	1.10	0.09	24 h	1.73	0.02
Blank	4.30	0.03	Blank	4.36	0.02	Blank	4.36	0.02
Amount Sorbed (mg/L)	**Percent Sorption**		**Amount Sorbed (mg/L)**	**Percent Sorption**		**Amount Sorbed (mg/L)**	**Percent Sorption**	
3.91	90.56		3.25	74.63		2.63	60.35	
Normalized Sorption	0.906	0.007	**Normalized Sorption**	0.746	0.020	**Normalized Sorption**	0.604	0.004

Table A.16. P Removal by MM Amongst Competing Anions (C_0 = 5 mg PO_4^{3-}-P/L) (Continued)

Humic Acid								
Humic Acid (2 mg/L)			Humic Acid (5 mg/L)			Humic Acid (10 mg/L)		
Time (hr)	Phosphate conc. (mg/L)	standard deviation	Time (hr)	Phosphate conc. (mg/L)	standard deviation	Time (hr)	Phosphate conc. (mg/L)	standard deviation
0 h	4.88	0.02	0 h	4.81	0.03	0 h	4.86	0.02
24 h	0.28	0.02	24 h	0.29	0.02	24 h	0.28	0.02
Blank	4.74	0.15	Blank	4.78	0.11	Blank	4.85	0.03
Amount Sorbed (mg/L)	Percent Sorption		Amount Sorbed (mg/L)	Percent Sorption		Amount Sorbed (mg/L)	Percent Sorption	
4.60	94.23		4.52	93.91		4.58	94.20	
Normalized Sorption	0.942	0.003	Normalized Sorption	0.939	0.003	Normalized Sorption	0.942	0.003
NOM								
NOM (5 mg/L)			NOM (10 mg/L)			NOM (50 mg/L)		
Time (hr)	Phosphate conc. (mg/L)	standard deviation	Time (hr)	Phosphate conc. (mg/L)	standard deviation	Time (hr)	Phosphate conc. (mg/L)	standard deviation
0 h	4.98	0.02	0 h	4.88	0.03	0 h	4.77	0.07
24 h	0.37	0.07	24 h	0.28	0.02	24 h	0.56	0.04
Blank	4.81	0.18	Blank	4.81	0.10	Blank	4.79	0.03
Amount Sorbed (mg/L)	Percent Sorption		Amount Sorbed (mg/L)	Percent Sorption		Amount Sorbed (mg/L)	Percent Sorption	
4.61	92.53		4.60	94.23		4.20	88.18	
Normalized Sorption	0.925	0.015	Normalized Sorption	0.942	0.003	Normalized Sorption	0.882	0.009

Table A.16. P Removal by MM Amongst Competing Anions (C$_0$ = 5 mg PO$_4^{3-}$-P/L) (Continued)

	Control	
	Control (5 mg/L)	
Time (hr)	**Phosphate Conc. (mg/L)**	**Standard deviation**
0	4.95	0.03
24	0.37	0.07
Blank	4.91	0.06
Amount Sorbed (mg/L)	**Percent Sorption**	
4.58	92.48	
Normalized Sorption	0.925	0.015

Table A.17. P Removal of METAC MIPs in the presence of various competing charges (C_0 = 5 mg PO_4^{3-}-P/L)

Anion Species	MM		MM4		MM12	
	Anion Charge (meq/L)	Normalized P Removal	Anion Charge (meq/L)	Normalized P Removal	Anion Charge (meq/L)	Normalized P Removal
Control	0.00	0.92	0.00	0.92	0.00	0.92
Cl^-	1.41	0.55	1.41	0.60	1.41	0.58
Cl^-	2.82	0.45	2.82	0.54	2.82	0.52
Cl^-	14.10	0.19	14.10	0.22	14.10	0.21
HCO_3^-	0.08	0.89	0.08	0.90	0.08	0.92
HCO_3^-	0.16	0.90	0.16	0.92	0.16	0.92
HCO_3^-	0.82	0.94	0.82	0.91	0.82	0.91
SO_4^{2-}	1.04	0.75	1.04	0.76	1.04	0.77
SO_4^{2-}	2.08	0.53	2.08	0.55	2.08	0.52
SO_4^{2-}	20.82	0.14	20.82	0.17	20.82	0.19
NO_3^-	0.16	0.91	0.16	0.92	0.16	0.92
NO_3^-	0.81	0.75	0.81	0.80	0.81	0.77
NO_3^-	1.61	0.60	1.61	0.63	1.61	0.65
F^-	10.06	0.20	-	-	-	-
Br^-	7.11	0.31	-	-	-	-
F^-	5.23	0.35	-	-	-	-
Br^-	5.10	0.32	-	-	-	-
$C_2H_3O_2^-$	7.92	0.25	-	-	-	-

A.3.5. METAC Desorption Study

Table A.18. Desorption of P from MM12 Over Six Time-Variable Cycles

Cumulative Days	Cycles	Normalized P Removal	Normalized StdDev
0	0	0.00	0.00
0.25	Cycle 1 (6 hr)	3.27	0.22
0.5	Cycle 2 (6 hr)	0.67	0.02
1	Cycle 3 (12 hr)	0.44	0.04
2	Cycle 4 (24 hr)	0.29	0.02
3	Cycle 5 (24 hr)	0.23	0.02
4	Cycle 6 (24 hr)	0.09	0.05

A.3.6. METAC Biodegradability Respirometer Data

Table A.19. BOD of Blank and Sample with 100 mg MM12

Days	Blank Corrected Reading	Blank Corrected StdDev	Sample Corrected Reading	Sample Corrected StdDev
0.00	0.00	0.00	0.00	0.00
0.02	-21.82	3.62	-21.82	6.26
0.04	-34.38	4.74	-27.62	2.46
0.06	-36.32	5.34	-30.03	2.46
0.08	-39.70	6.26	-36.80	4.27
0.10	-37.77	4.10	-37.77	1.81
0.12	-36.32	9.40	-36.80	2.46
0.14	-34.87	7.20	-37.28	4.16
0.16	-33.42	10.34	-31.48	6.73
0.18	-34.38	9.86	-29.55	2.98
0.20	-31.00	13.34	-26.65	3.62
0.22	-33.42	10.07	-28.10	1.37
0.25	-27.13	14.71	-21.33	2.98
0.27	-26.65	17.17	-21.33	1.81

Days	Blank Corrected Reading	Corrected StdDev	Sample Corrected Reading	Corrected StdDev
0.31	-25.20	15.99	-17.95	2.46
0.33	-28.10	16.80	-14.08	2.37
0.35	-28.10	14.80	-17.47	2.98
0.37	-22.78	15.41	-11.18	4.93
0.39	-23.75	17.44	-10.70	8.89
0.41	-21.82	18.08	-2.00	7.76
0.43	-21.33	18.43	-4.42	10.12
0.45	-20.85	19.68	4.28	11.92
0.47	-22.78	19.38	0.90	8.89
0.50	-18.92	20.41	6.70	10.85
0.52	-14.08	17.57	7.18	12.44
0.54	-15.05	19.03	11.05	14.32
0.56	-13.60	23.51	12.50	17.68
0.58	-16.50	21.34	11.53	17.01
0.60	-11.67	21.52	9.12	18.79
0.62	-4.90	26.93	20.23	17.57
0.64	-10.70	23.29	17.82	20.59
0.66	-5.87	24.39	24.10	19.92
0.68	-7.80	29.34	20.23	21.34
0.70	-9.73	20.38	23.13	25.40
0.72	-2.48	23.51	31.35	20.38
0.75	-4.90	22.38	27.00	20.17
0.77	-3.45	20.95	32.80	23.27
0.79	-1.52	23.94	36.67	19.70
0.81	-2.97	20.14	33.28	23.24
0.83	0.90	25.12	41.50	23.27
0.85	-3.93	22.18	40.05	25.54
0.87	1.38	39.16	37.15	27.31
0.89	-22.30	43.41	27.48	20.54
0.91	-30.03	29.74	26.03	21.32
0.93	-8.77	31.11	37.15	20.86
0.95	-7.80	22.49	39.57	25.54
0.97	4.28	28.42	53.10	25.64
1.00	0.90	24.19	56.48	29.74
1.02	8.15	27.97	63.73	28.42
1.04	4.77	20.86	63.25	35.56
1.06	6.22	22.35	67.60	35.33
1.10	12.98	25.06	75.33	29.98

Days	**Blank** Corrected Reading	Corrected StdDev	**Sample** Corrected Reading	Corrected StdDev
1.16	9.12	23.05	79.20	37.45
1.18	17.33	28.42	84.52	37.11
1.20	11.05	25.51	82.10	38.50
1.22	12.50	25.06	85.00	36.78
1.25	21.68	28.25	87.90	36.17
1.27	19.27	28.42	91.77	33.49
1.29	19.75	31.79	88.87	29.10
1.31	15.88	29.22	99.50	36.12
1.33	13.95	29.88	92.73	32.86
1.35	15.88	31.26	100.95	32.43
1.37	14.92	31.26	98.05	30.62
1.39	19.27	33.49	102.88	32.69
1.41	18.30	33.49	99.02	32.96
1.43	21.20	33.00	102.88	33.66
1.45	18.78	34.03	102.40	32.34
1.47	24.10	34.98	103.37	31.69
1.50	21.68	34.69	99.50	35.30
1.52	22.65	34.15	105.78	34.77
1.54	26.03	32.52	100.95	35.30
1.56	30.87	33.18	110.13	31.39
1.58	27.00	33.84	105.78	33.66
1.60	26.03	36.61	106.75	34.15
1.62	24.10	35.80	107.23	33.72
1.64	22.65	34.70	107.23	32.34
1.66	27.48	36.03	109.17	34.65
1.68	28.45	32.90	112.07	36.45
1.70	32.80	31.32	115.93	38.26
1.72	34.73	36.70	118.35	38.59
1.75	34.73	37.83	117.87	36.82
1.77	34.73	34.03	120.28	38.42
1.79	34.25	34.93	117.38	37.27
1.81	38.12	38.73	121.73	35.21
1.83	40.05	36.50	119.80	38.26
1.85	38.12	38.31	122.70	35.30
1.87	41.98	42.56	123.18	36.82
1.89	32.80	43.89	120.28	32.77
1.91	32.32	41.70	120.77	32.18
1.95	38.12	37.89	126.08	38.11

Table A.19. BOD of Blank and Sample with 100 mg MM12 (Continued)

Days	**Blank** Corrected Reading	Corrected StdDev	**Sample** Corrected Reading	Corrected StdDev
2.02	42.95	38.26	130.43	38.91
2.04	49.72	38.93	133.82	38.11
2.06	47.30	38.59	130.92	38.77
2.08	47.30	39.41	132.85	39.27
2.10	47.78	38.93	130.92	40.19
2.12	48.75	37.14	132.85	40.10
2.14	48.27	42.30	139.13	38.26
2.16	48.27	38.04	138.65	41.87
2.18	51.65	41.39	142.03	39.22
2.20	51.17	39.93	138.65	41.05
2.22	48.27	39.16	138.65	40.57
2.25	54.07	40.10	140.10	41.54
2.27	55.03	41.19	139.13	39.08
2.29	54.07	39.38	141.55	41.54
2.31	55.52	43.21	142.03	41.22
2.33	55.52	42.37	143.97	43.40
2.35	55.03	41.02	147.83	43.54
2.37	52.62	40.08	136.72	42.76
2.39	52.62	40.57	142.52	40.95
2.41	51.65	42.56	141.55	40.96
2.43	54.55	42.51	143.48	40.19
2.45	49.72	46.30	138.65	43.60
2.47	53.10	43.70	157.02	47.80
2.50	50.20	47.30	146.87	42.11
2.52	55.03	46.64	147.35	42.11
2.54	57.93	44.08	151.70	44.23
2.56	58.90	45.32	149.28	43.06
2.58	58.42	45.32	156.53	44.87
2.60	54.07	46.16	154.60	44.34
2.62	54.07	44.36	148.32	43.54
2.64	56.48	44.56	155.57	44.70
2.66	60.35	47.96	151.22	46.00
2.68	60.35	43.23	154.60	44.97
2.70	60.83	47.22	153.63	44.84
2.72	66.63	46.54	154.12	45.49
2.75	62.77	43.42	152.67	44.36
2.77	67.60	48.37	159.43	44.49
2.81	66.15	48.35	157.02	44.05

Days	**Blank** Corrected Reading	Corrected StdDev	**Sample** Corrected Reading	Corrected StdDev
2.87	73.40	45.03	162.82	44.49
2.89	75.82	47.78	170.55	44.70
2.91	72.43	51.02	167.17	45.32
2.93	72.43	50.91	163.30	43.70
2.95	69.05	49.29	170.07	39.66
2.97	71.47	45.81	165.72	41.87
3.00	75.33	51.06	168.13	44.05
3.02	72.92	50.91	174.90	44.97
3.04	77.75	50.43	169.58	44.56
3.06	80.17	48.78	171.52	44.99
3.08	85.97	47.96	171.03	47.31
3.10	82.10	50.91	172.48	41.47
3.12	82.58	51.69	173.93	45.81
3.14	84.52	52.71	173.93	43.92
3.16	78.72	48.33	171.52	47.69
3.18	83.55	50.57	174.42	49.77
3.20	87.90	48.11	178.28	50.91
3.22	91.28	49.90	183.12	46.47
3.25	93.22	51.28	183.60	47.49
3.27	94.67	52.18	182.63	49.29
3.29	96.60	49.69	187.95	49.91
3.31	96.60	51.09	187.95	47.45
3.33	99.50	53.04	191.82	51.39
3.35	102.88	55.49	191.82	50.51
3.37	101.43	55.71	193.27	49.35
3.39	105.30	53.53	196.17	47.67
3.41	104.82	50.83	195.68	48.45
3.43	107.23	53.03	195.68	50.91
3.45	106.27	51.09	198.58	52.07
3.47	106.75	49.90	196.17	49.59
3.50	109.17	55.28	198.10	50.26
3.52	107.72	52.13	199.55	51.88
3.54	108.68	53.28	202.45	52.22
3.56	107.72	52.58	205.35	50.41
3.58	114.00	54.61	198.58	52.89
3.60	111.58	52.18	198.58	45.95
3.62	112.55	52.37	200.52	50.08
3.66	110.62	52.54	199.55	50.73

Days	Blank Corrected Reading	Corrected StdDev	Sample Corrected Reading	Corrected StdDev
3.72	105.78	56.61	194.72	50.41
3.75	109.65	57.09	200.03	51.68
3.77	108.68	54.63	199.07	52.72
3.79	109.17	53.71	195.68	51.42
3.81	109.17	53.92	196.65	53.37
3.83	109.17	56.61	199.55	53.03
3.85	108.20	55.98	196.65	49.42
3.87	105.78	53.86	197.62	50.57
3.89	103.37	55.94	196.17	49.91
3.91	107.72	55.04	198.10	48.70
3.93	110.62	57.52	196.65	51.42
3.95	105.30	55.71	196.65	52.72
3.97	108.20	54.15	194.23	51.80
4.00	108.68	57.28	196.65	52.71
4.02	108.68	51.50	195.20	53.88
4.04	107.72	56.38	198.58	51.22
4.06	105.78	55.27	192.78	51.42
4.08	105.78	56.98	192.30	52.87
4.10	104.82	57.34	193.27	51.09
4.12	102.40	55.27	193.27	53.74
4.14	108.68	60.48	193.75	54.13
4.16	106.75	55.98	192.30	56.81
4.18	104.33	58.87	194.72	53.74
4.20	106.27	59.07	195.20	53.49
4.22	109.17	56.88	195.68	52.58
4.25	114.48	60.63	198.10	51.42
4.27	115.93	58.88	205.35	54.53
4.29	115.93	57.35	202.45	51.22
4.31	119.80	56.68	207.28	51.04
4.33	117.87	58.41	211.15	51.57
4.35	122.22	58.17	213.08	52.87
4.37	120.77	55.49	209.22	56.81
4.39	120.28	55.49	209.22	51.80
4.41	118.83	57.73	206.80	55.00
4.43	119.80	57.35	205.35	53.37
4.45	116.90	59.15	203.90	56.84
4.47	118.83	57.82	208.25	53.53
4.52	125.60	57.29	211.15	51.93

Days	Blank Corrected Reading	Corrected StdDev	Sample Corrected Reading	Corrected StdDev
4.58	128.98	59.29	213.57	55.04
4.60	122.22	60.67	211.63	54.53
4.62	119.32	59.51	209.70	54.13
4.64	118.83	58.88	208.25	53.88
4.66	123.18	59.62	207.28	54.53
4.68	121.25	57.09	208.25	52.23
4.70	115.93	62.01	206.32	53.69
4.72	121.73	60.21	206.32	53.37
4.75	118.35	60.48	206.80	53.37
4.77	119.80	59.96	206.80	54.88
4.79	121.25	59.98	208.73	49.91
4.81	117.87	60.29	205.35	53.69
4.83	123.18	58.88	209.22	53.49
4.85	124.63	62.67	211.63	55.69
4.87	120.77	68.72	206.80	59.16
4.89	106.75	67.09	198.10	51.28
4.91	105.78	64.52	196.17	49.11
4.93	107.72	61.76	194.72	51.72
4.95	114.00	61.32	203.42	52.37
4.97	115.93	62.21	202.93	53.37
5.00	121.25	64.27	206.80	57.36
5.02	121.73	63.71	210.18	55.49
5.04	127.53	61.34	210.18	56.64
5.06	121.25	64.47	210.18	54.68
5.08	124.15	64.08	210.18	58.14
5.10	120.77	60.01	208.73	57.14
5.12	124.63	61.14	210.67	58.65
5.14	127.53	61.09	212.12	58.52
5.16	125.60	57.54	211.63	58.30
5.18	129.47	64.47	217.43	52.87
5.20	132.85	62.09	217.92	55.99
5.22	133.82	61.31	222.27	54.34
5.25	135.27	62.67	221.78	55.49
5.27	136.72	62.65	223.23	55.33
5.29	134.78	59.34	221.30	57.47
5.31	131.40	63.14	218.88	56.98
5.33	131.88	63.71	222.75	54.88
5.37	134.30	59.81	222.27	53.37

Days	Blank Corrected Reading	Corrected StdDev	Sample Corrected Reading	Corrected StdDev
5.43	139.62	60.29	223.72	56.21
5.45	139.13	62.01	225.17	55.55
5.47	140.58	63.77	222.75	53.69
5.50	136.23	65.41	220.82	53.69
5.52	133.33	62.88	219.37	56.21
5.54	137.68	67.14	220.82	50.91
5.56	131.88	66.47	219.37	51.57
5.58	134.30	65.60	218.40	52.89
5.60	134.30	65.56	218.88	53.37
5.62	132.37	62.72	216.47	54.70
5.64	132.37	67.21	219.85	53.08
5.66	132.37	64.94	219.37	56.21
5.68	139.62	60.29	222.27	57.64
5.70	140.58	63.32	223.72	55.74
5.72	139.62	62.47	224.68	54.41
5.75	140.10	66.07	225.65	53.03
5.77	135.27	64.47	227.10	53.75
5.79	140.10	63.55	224.68	55.84
5.81	138.17	62.43	224.20	57.05
5.83	140.58	60.63	227.10	55.66
5.85	140.58	63.14	225.17	57.35
5.87	130.43	68.70	225.17	63.61
5.89	126.08	70.70	216.47	53.49
5.91	120.77	71.05	215.98	54.21
5.93	124.63	63.99	215.02	55.95
5.95	134.30	62.65	220.82	59.81
5.97	136.23	64.47	222.75	59.74
6.00	141.55	62.24	225.17	60.19
6.02	140.58	63.99	232.90	59.98
6.04	140.10	59.60	230.00	60.86
6.06	145.42	60.95	228.07	58.63
6.08	141.07	58.63	230.97	62.20
6.10	145.90	62.01	231.45	59.30
6.12	143.48	60.88	230.48	58.17
6.14	143.48	60.88	231.45	58.78
6.16	143.48	61.98	232.42	57.28
6.18	144.45	64.89	229.03	56.83
6.22	146.38	63.14	234.83	58.63

Days	Blank Corrected Reading	Corrected StdDev	Sample Corrected Reading	Corrected StdDev
6.29	144.93	60.42	231.45	56.61
6.31	146.87	61.54	233.87	62.21
6.33	146.87	62.75	233.87	59.51
6.35	147.35	58.41	233.38	61.31
6.37	148.32	62.47	235.80	59.51
6.39	148.80	59.98	239.18	62.43
6.41	149.77	61.31	236.28	62.28
6.43	151.22	59.93	239.18	64.01
6.45	145.90	62.87	237.73	60.19
6.47	150.25	64.37	237.25	55.27
6.50	144.45	64.66	237.73	58.17
6.52	146.38	62.94	235.80	58.41
6.54	145.42	65.78	235.80	54.34
6.56	146.38	63.34	233.38	53.56
6.58	144.45	65.88	235.80	55.28
6.60	143.97	68.27	235.80	56.21
6.62	147.83	66.56	233.87	54.18
6.64	143.00	64.70	234.35	55.33
6.66	145.90	68.26	237.25	55.00
6.68	148.32	67.35	237.25	55.99
6.70	147.35	65.62	236.77	53.95
6.72	148.80	67.23	236.77	55.49
6.75	146.38	66.01	237.73	53.03
6.77	148.80	62.43	241.60	54.70
6.79	152.18	64.70	241.60	57.16
6.81	150.25	66.05	240.63	57.64
6.83	149.77	67.79	239.67	56.64
6.85	144.93	66.47	237.73	56.68
6.87	144.45	66.56	234.35	56.81
6.89	147.35	68.69	239.67	59.27
6.91	142.52	66.90	237.25	56.49
6.93	143.48	65.78	240.15	56.49
6.95	144.93	64.01	238.70	56.99
6.97	146.38	65.11	240.63	57.64
7.00	147.35	63.99	241.60	57.49
7.02	148.80	64.94	245.47	59.81
7.04	152.67	60.86	244.50	64.55
7.08	157.02	65.34	247.88	60.91

Table A.19. BOD of Blank and Sample with 100 mg MM12 (Continued)

Days	**Blank**		**Sample**	
	Corrected Reading	Corrected StdDev	Corrected Reading	Corrected StdDev
7.14	148.32	64.47	244.50	63.81
7.16	151.70	61.53	245.95	61.08
7.18	156.53	68.26	246.92	61.76
7.20	155.57	62.75	249.82	63.06
7.22	160.40	62.87	248.85	58.80
7.25	157.50	63.77	251.75	60.75
7.27	157.98	67.57	247.40	59.03
7.29	154.12	69.37	248.37	60.91
7.31	155.57	68.91	244.98	56.84
7.33	157.98	67.79	244.50	60.46
7.35	153.63	66.68	243.53	54.25
7.37	151.22	67.40	243.53	56.06
7.39	150.73	69.61	243.05	55.33
7.41	155.08	71.21	244.50	54.84
7.43	154.12	70.36	244.02	52.87
7.45	152.18	70.03	244.02	57.64
7.47	155.08	72.31	241.60	55.94
7.50	154.12	70.62	244.02	55.66
7.52	154.60	69.43	244.02	56.46
7.54	153.63	71.30	244.50	54.39
7.56	153.63	71.57	242.57	53.88
7.58	154.60	72.27	243.53	54.39
7.60	151.70	70.62	246.92	52.87
7.62	154.60	69.61	244.02	55.84
7.64	155.08	71.30	244.50	53.53
7.66	155.08	68.85	243.53	53.23
7.68	155.57	71.63	248.37	52.54
7.70	157.98	71.39	245.47	54.18
7.72	155.57	71.60	243.05	53.03
7.75	157.02	70.96	244.50	52.54
7.77	157.50	71.30	245.47	53.09
7.79	159.43	70.78	247.88	55.69
7.81	156.05	73.75	245.95	54.01
7.83	157.50	72.13	246.92	51.28
7.85	152.67	73.61	245.47	52.37
7.87	151.70	78.48	240.63	58.39
7.89	129.95	79.90	232.90	49.11
7.93	136.23	74.35	230.48	50.64

Days	Blank Corrected Reading	Corrected StdDev	Sample Corrected Reading	Corrected StdDev
8.00	146.87	71.63	237.25	57.23
8.02	155.08	71.37	238.70	56.84
8.04	153.15	72.81	240.15	59.33
8.06	150.25	68.02	239.18	61.08
8.08	156.05	67.12	245.47	62.27
8.10	157.50	68.70	243.05	60.42
8.12	159.43	70.04	248.85	58.45
8.14	154.60	69.85	251.75	59.03
8.16	157.50	73.40	246.92	57.36
8.18	157.98	72.50	249.33	59.68
8.20	157.02	70.96	247.88	58.80
8.22	158.95	72.07	248.85	61.41
8.25	160.40	70.89	249.82	64.46
8.27	162.82	71.04	248.37	62.56
8.29	159.92	70.76	245.47	61.91
8.31	164.27	70.48	251.75	63.54
8.33	167.17	70.48	251.75	65.10
8.35	163.30	69.01	250.78	63.17
8.37	164.27	71.16	250.78	63.57
8.39	161.85	72.31	250.78	64.98
8.41	167.17	72.66	250.30	60.46
8.43	161.85	67.79	246.43	57.87
8.45	162.33	71.16	245.47	59.81
8.47	161.85	70.93	245.47	57.30
8.50	162.33	72.74	245.95	57.49
8.52	157.98	74.09	244.02	59.45
8.54	156.05	72.50	241.12	59.60
8.56	157.02	74.59	243.05	56.15
8.58	159.43	75.37	245.95	62.06
8.60	160.88	75.69	244.98	62.39
8.62	160.88	72.07	248.85	59.30
8.64	158.95	75.52	243.53	58.95
8.66	163.78	73.23	247.88	60.10
8.68	161.37	79.49	244.98	58.29
8.70	158.47	77.20	246.92	56.84
8.72	158.47	74.43	245.95	57.52
8.75	155.08	77.42	242.57	58.69
8.79	162.33	75.64	244.98	60.84

Days	Blank Corrected Reading	Corrected StdDev	Sample Corrected Reading	Corrected StdDev
8.85	160.40	75.44	247.88	59.81
8.87	151.22	81.80	244.02	69.24
8.89	138.65	80.42	234.35	48.93
8.91	137.20	81.38	224.68	53.69
8.93	143.00	78.23	235.32	60.75
8.95	152.67	76.73	240.15	56.58
8.97	158.95	73.49	243.53	64.20
9.00	163.78	74.84	242.57	62.27
9.02	160.40	76.73	245.95	62.22
9.04	163.30	75.13	249.82	60.10
9.06	163.78	74.01	250.78	65.84
9.08	165.72	75.94	248.85	59.81
9.10	166.20	75.69	251.27	62.41
9.12	166.68	74.96	247.88	63.43
9.14	162.82	74.69	247.40	65.50
9.16	163.30	76.08	246.43	64.87
9.18	163.78	75.94	248.37	61.49
9.20	161.85	79.04	248.85	62.84
9.22	162.82	76.88	247.88	61.41
9.25	164.27	76.62	245.47	62.78
9.27	165.23	77.41	242.08	60.60
9.29	160.88	77.72	248.37	58.80
9.31	163.78	79.18	247.40	59.95
9.33	164.27	80.00	247.40	58.91
9.35	164.75	77.55	246.43	64.34
9.37	166.68	77.82	249.33	60.97
9.39	164.75	82.44	248.37	60.46
9.41	163.78	78.91	250.78	61.57
9.43	166.20	78.36	248.85	61.67
9.45	165.23	77.29	249.33	64.08
9.47	168.62	77.82	249.82	60.97
9.50	164.75	79.25	249.82	62.27
9.52	167.65	80.00	248.85	62.41
9.54	169.10	75.44	250.78	62.78
9.56	168.62	75.11	249.33	61.62
9.58	168.62	77.29	252.23	64.08
9.60	171.52	80.00	252.72	60.08
9.64	170.55	79.85	254.65	60.10

Table A.19. BOD of Blank and Sample with 100 mg MM12 (Continued)

Days	Blank Corrected Reading	Corrected StdDev	Sample Corrected Reading	Corrected StdDev
9.70	170.07	75.24	251.75	60.97
9.72	169.58	80.00	252.72	56.99
9.75	169.10	78.91	252.23	60.60
9.77	169.10	78.14	252.72	62.30
9.79	170.55	79.75	253.20	59.03
9.81	168.13	82.33	254.17	61.11
9.83	166.68	78.93	253.20	61.26
9.85	167.17	79.59	250.78	62.92
9.87	171.03	83.23	249.82	66.67
9.89	158.47	85.35	247.88	54.18
9.91	155.08	89.89	238.70	55.07
9.93	148.32	85.45	237.73	56.99
9.95	155.08	82.04	247.88	62.78
9.97	159.92	83.80	239.18	60.46
10.00	157.50	83.81	236.28	63.43
10.02	159.43	84.28	237.25	65.24
10.04	159.43	81.08	239.67	61.67
10.06	161.37	79.85	240.63	65.89
10.08	160.40	80.27	243.53	64.59
10.10	161.37	82.44	243.05	65.10
10.12	163.78	80.94	239.18	61.77
10.14	164.75	83.60	244.02	64.01
10.16	164.75	80.60	244.98	64.46
10.18	164.27	83.39	244.02	61.11
10.20	155.57	85.58	251.75	66.45
10.22	146.87	84.49	242.57	63.94
10.25	139.62	84.36	243.53	61.11
10.27	137.68	83.55	241.60	67.69
10.29	141.07	83.55	241.60	63.81
10.31	136.72	82.62	243.05	65.38
10.33	139.62	78.98	248.85	65.88
10.35	141.07	79.65	248.37	63.87
10.37	140.10	81.00	251.75	66.83
10.39	141.55	81.00	251.27	65.89
10.41	141.07	80.10	253.68	68.47
10.43	145.42	81.27	252.72	67.50
10.45	141.55	80.32	254.65	67.32
10.50	144.45	79.21	249.82	65.62

Table A.19. BOD of Blank and Sample with 100 mg MM12 (Continued)

Days	**Blank** Corrected Reading	Corrected StdDev	**Sample** Corrected Reading	Corrected StdDev
10.54	150.25	78.09	247.40	65.75
10.56	152.67	80.59	248.85	62.27
10.58	152.67	78.54	247.40	65.10
10.60	151.70	80.10	249.33	65.75
10.62	152.67	80.79	253.20	67.69
10.64	151.22	81.70	248.85	64.34
10.66	151.22	82.14	252.23	67.33
10.68	153.63	80.35	245.95	67.19
10.70	153.15	78.76	249.82	64.86
10.72	153.15	81.44	248.37	66.40
10.75	156.53	79.21	251.27	67.84
10.77	157.98	80.32	256.58	69.50
10.79	157.98	81.23	257.07	69.12
10.81	162.33	82.81	257.07	70.01
10.83	161.37	80.55	261.90	68.85
10.85	163.78	83.68	261.90	69.29
10.87	163.78	79.43	261.90	69.88
10.89	163.78	80.32	259.97	71.69
10.91	163.78	80.55	262.38	68.70
10.93	163.30	80.55	262.38	68.89
10.95	160.88	81.95	263.35	68.21
10.97	162.33	79.21	266.73	70.29
11.00	164.27	80.10	259.97	68.34
11.02	162.82	78.09	264.32	69.88
11.04	168.13	81.00	265.77	72.98
11.06	163.30	76.75	265.77	71.31
11.08	166.20	80.10	266.73	71.69
11.10	165.72	83.68	263.83	70.40
11.12	166.68	83.68	264.80	68.85
11.14	165.23	78.09	263.35	68.60
11.16	165.72	78.33	263.83	72.21
11.18	167.17	77.22	268.67	69.45
11.20	169.10	77.42	266.73	70.29
11.22	169.10	79.88	268.67	70.01
11.25	167.17	80.13	267.70	69.76
11.27	167.65	81.67	269.63	71.18
11.29	169.10	80.77	266.25	70.15
11.33	171.52	83.72	270.60	71.26

Table A.19. BOD of Blank and Sample with 100 mg MM12 (Continued)

Days	Blank Corrected Reading	Corrected StdDev	Sample Corrected Reading	Corrected StdDev
11.39	167.17	79.49	267.70	66.40
11.41	167.17	84.40	262.87	71.82
11.43	143.97	93.04	252.72	57.64
11.45	138.65	90.04	241.12	62.39
11.47	141.07	81.89	248.85	61.08
11.50	150.25	80.32	251.27	66.18
11.52	150.73	81.00	249.82	65.75
11.54	155.57	77.86	251.75	68.14
11.56	155.57	79.67	251.75	65.38
11.58	157.98	81.52	256.10	65.52
11.60	152.67	82.14	253.68	65.62
11.62	147.83	85.03	249.82	68.48
11.64	141.07	81.72	250.30	68.72
11.66	136.72	82.47	249.82	69.37
11.68	128.98	82.85	247.88	69.64
11.70	128.02	81.52	245.95	73.79
11.72	129.95	81.00	242.57	69.43
11.75	129.95	81.44	242.08	68.26
11.77	133.82	79.67	243.53	72.62
11.79	131.88	82.94	241.12	70.44
11.81	134.30	82.76	238.70	69.08
11.83	134.78	83.32	238.70	70.28
11.85	134.78	81.99	239.67	68.72
11.87	138.17	85.02	236.77	68.70
11.89	134.30	83.73	236.28	67.95
11.91	136.72	82.85	235.32	69.53
11.93	129.47	83.05	238.22	72.61
11.95	130.43	81.80	239.67	68.60
11.97	131.40	82.80	239.18	71.24
12.00	133.33	82.78	237.25	70.40
12.02	137.20	83.79	237.25	67.84
12.04	140.10	83.47	240.63	70.81
12.06	139.62	85.72	241.60	67.43
12.08	139.62	83.32	243.53	67.98
12.10	142.03	79.06	242.57	68.79
12.12	143.48	80.67	241.12	69.29
12.14	141.55	81.33	245.95	68.48
12.18	146.38	80.67	242.57	67.20

Days	Blank Corrected Reading	Corrected StdDev	Sample Corrected Reading	Corrected StdDev
12.25	150.25	80.55	242.08	69.79
12.27	159.43	81.22	252.23	67.62
12.29	152.67	84.35	246.92	67.84
12.31	158.47	79.88	252.23	73.50
12.33	160.40	82.11	254.65	69.16
12.35	170.07	79.21	266.73	72.22
12.37	162.82	85.26	260.93	71.45
12.39	154.60	83.25	258.52	66.79
12.41	152.18	87.52	249.82	75.83
12.43	138.17	95.70	241.60	64.20
12.45	128.50	88.46	239.67	65.38
12.47	142.03	86.94	245.95	65.38
12.50	139.13	84.36	244.50	68.48
12.52	139.13	81.92	245.95	72.21
12.54	140.58	79.67	242.57	70.06
12.56	139.13	83.79	241.60	70.60
12.58	145.90	83.68	244.98	65.62
12.60	140.58	81.00	244.02	70.81
12.62	139.62	81.46	243.05	70.50
12.64	150.25	81.89	245.95	74.02
12.66	154.60	80.13	252.23	70.40
12.68	160.40	82.80	259.00	71.82
12.70	160.88	83.55	262.87	69.96
12.72	157.02	76.12	254.17	75.71
12.75	166.20	79.68	261.42	71.77
12.77	162.82	81.90	259.48	78.64
12.79	162.82	80.39	261.42	77.64
12.81	161.85	80.10	258.52	73.38
12.83	152.67	85.02	258.52	71.68
12.85	151.70	82.19	256.10	73.15
12.87	147.83	83.52	247.40	70.81
12.89	147.35	83.92	250.30	71.96
12.91	144.93	81.44	244.50	68.08
12.93	145.42	82.59	244.50	70.80
12.95	152.67	83.46	248.85	68.60
12.97	150.73	81.44	247.88	70.93
13.00	153.63	83.47	252.23	71.05
13.04	146.87	83.01	245.95	67.95

Days	Blank Corrected Reading	Corrected StdDev	Sample Corrected Reading	Corrected StdDev
13.10	149.28	83.47	246.92	70.28
13.12	161.37	82.78	251.27	72.94
13.14	160.40	81.46	255.13	70.17
13.16	149.77	84.39	249.33	70.28
13.18	153.15	81.72	251.27	73.63
13.20	155.57	84.81	253.68	71.77
13.22	156.53	87.27	250.78	70.28
13.25	153.15	84.35	248.37	69.64
13.27	151.22	80.59	248.85	71.57
13.29	157.02	78.79	253.20	73.63
13.31	165.23	83.68	259.48	73.04
13.33	157.98	84.14	257.07	72.09
13.35	153.15	83.46	250.78	69.70
13.37	147.83	82.85	252.72	70.40
13.39	146.87	82.85	254.65	72.94
13.41	155.57	80.86	258.03	70.28
13.43	152.67	82.59	251.27	70.50
13.45	143.97	84.13	250.30	70.53
13.47	141.07	85.89	250.78	74.02
13.50	137.68	80.34	247.40	68.79
13.52	146.87	82.85	248.37	69.24
13.54	138.17	82.11	251.75	71.05
13.56	143.97	81.52	249.82	71.14
13.58	150.25	81.72	249.82	68.08
13.60	143.97	83.13	247.40	68.07
13.62	141.07	80.13	246.92	70.93
13.64	144.45	83.05	247.88	70.80
13.66	140.58	83.92	245.95	72.34
13.68	143.97	83.46	243.05	72.86
13.70	137.68	84.18	240.63	72.51
13.72	137.68	83.72	240.63	70.50
13.75	141.07	79.82	247.40	75.59
13.77	166.20	83.47	263.83	74.28
13.79	145.42	83.23	245.95	71.69
13.81	141.07	84.57	246.43	72.86
13.83	143.48	83.47	243.53	70.15
13.85	142.52	86.39	246.43	68.89
13.89	144.45	83.32	246.43	71.57

Table A.19. BOD of Blank and Sample with 100 mg MM12 (Continued)

Days	Blank Corrected Reading	Corrected StdDev	Sample Corrected Reading	Corrected StdDev
13.95	142.52	84.18	248.85	66.56
13.97	140.58	83.68	240.15	69.24
14.00	140.58	82.59	244.50	71.24
14.02	151.22	85.52	253.20	69.76
14.04	145.90	83.61	249.33	69.64
14.06	152.18	79.00	251.75	72.41
14.08	161.85	84.75	253.20	68.16
14.10	152.18	83.13	247.40	67.52
14.12	151.22	83.69	247.88	71.05
14.14	157.50	85.70	251.27	70.28
14.16	157.50	83.25	249.82	70.28
14.18	153.63	87.05	252.72	69.24
14.20	155.57	81.23	248.85	74.67
14.22	168.13	83.47	257.55	72.86
14.25	171.52	88.16	260.93	71.57
14.27	163.30	85.47	259.00	70.93
14.29	157.98	87.27	253.68	68.60
14.31	153.15	84.59	255.13	70.28
14.33	154.12	86.18	249.82	68.08
14.35	149.28	88.84	250.30	68.48
14.37	147.35	84.57	250.30	67.95
14.39	146.38	82.66	251.27	68.16
14.41	143.00	83.52	248.85	71.18
14.43	153.15	85.72	248.85	67.20
14.45	148.32	85.98	249.82	67.09
14.47	146.87	84.93	249.33	68.26
14.50	144.45	84.18	250.78	68.16
14.52	140.58	85.47	248.85	68.72
14.54	143.97	86.45	248.37	67.84
14.56	144.93	85.79	247.88	67.73
14.58	148.80	89.01	247.40	69.78
14.60	147.83	86.45	247.40	65.92
14.62	149.28	88.83	247.88	70.01
14.64	147.83	87.94	248.85	69.12
14.66	143.48	86.45	248.37	68.48
14.68	147.83	86.36	245.95	69.64
14.70	148.32	85.26	247.40	69.43
14.75	147.83	88.25	246.43	69.76

Table A.19. BOD of Blank and Sample with 100 mg MM12 (Continued)

Days	Blank Corrected Reading	Corrected StdDev	Sample Corrected Reading	Corrected StdDev
14.81	148.80	86.81	247.40	66.27
14.83	149.28	87.06	247.88	68.34
14.85	147.83	89.52	248.37	66.79
14.87	147.35	88.61	245.47	64.22
14.89	148.32	86.39	247.40	65.62
14.91	148.80	87.27	246.92	68.79
14.93	151.70	88.84	244.02	67.84
14.95	151.70	90.52	244.98	67.43
14.97	151.22	87.87	246.43	64.22
15.00	150.73	87.31	247.40	65.75
15.02	150.73	87.72	245.95	70.53
15.04	151.22	88.85	246.43	68.34
15.06	146.87	87.48	248.37	68.99
15.08	146.87	89.72	246.43	66.89
15.10	149.28	89.50	244.02	67.05
15.12	149.77	85.69	246.43	65.52
15.14	150.73	86.85	244.98	67.09
15.16	154.12	91.58	247.88	70.80
15.18	149.28	89.78	244.50	64.86
15.20	150.25	85.94	244.02	65.28
15.22	149.77	88.38	246.92	69.37
15.25	149.77	89.95	243.53	67.43
15.27	150.73	88.61	244.98	68.26
15.29	152.67	90.65	244.98	69.37
15.31	150.25	86.14	246.43	66.91
15.33	151.70	89.98	248.85	67.95
15.35	154.12	88.91	248.37	69.24
15.37	153.15	89.72	248.85	69.12
15.39	152.18	87.49	249.82	67.69
15.41	153.63	87.71	247.40	68.60
15.43	153.63	89.05	249.33	66.40
15.45	153.15	87.52	249.82	68.21
15.47	153.15	88.61	247.88	62.74
15.50	154.12	89.81	247.40	68.21
15.52	152.67	85.93	250.78	67.09
15.54	155.08	87.48	246.43	67.56
15.56	154.12	89.72	245.47	64.34
15.60	154.12	88.45	243.53	67.33

Table A.19. BOD of Blank and Sample with 100 mg MM12 (Continued)

Days	Blank Corrected Reading	Corrected StdDev	Sample Corrected Reading	Corrected StdDev
15.66	154.12	86.39	247.88	66.56
15.68	153.15	89.31	243.05	67.56
15.70	152.18	86.14	249.82	66.99
15.72	154.12	89.78	245.95	66.14
15.75	151.70	87.05	248.85	63.43
15.77	151.22	90.19	247.40	66.54
15.79	153.63	89.28	245.95	64.87
15.81	151.70	89.39	244.02	66.89
15.83	152.67	86.14	248.37	66.79
15.85	155.57	88.45	245.47	66.14
15.87	152.18	84.85	247.40	67.19
15.89	155.57	87.27	245.47	64.59
15.91	153.15	87.49	250.30	67.05
15.93	152.18	88.39	249.33	64.08
15.95	154.60	89.58	246.92	65.38
15.97	152.18	85.69	248.85	63.87
16.00	153.63	85.92	245.95	66.99
16.02	153.15	89.20	248.85	62.53
16.04	153.15	89.52	244.02	67.73
16.06	151.70	85.69	248.85	65.38
16.08	155.08	89.73	248.85	65.18
16.10	152.67	91.32	249.33	62.78
16.12	153.63	92.63	247.88	67.84
16.14	152.67	87.06	246.43	66.27
16.16	153.63	87.26	249.33	69.50
16.18	154.12	87.03	244.02	67.73
16.20	156.05	88.84	247.40	66.40
16.22	156.05	88.39	249.82	67.52
16.25	156.05	90.39	246.43	64.11
16.27	155.57	89.95	246.92	64.11
16.29	156.05	89.28	251.27	68.34
16.31	156.05	87.52	252.72	67.31
16.33	157.50	87.48	247.40	66.99
16.35	152.67	89.28	245.95	69.50
16.37	152.67	86.39	246.92	66.79
16.39	154.60	88.18	245.47	64.46
16.41	149.28	88.18	243.53	66.40
16.45	144.93	89.28	242.57	67.95

Table A.19. BOD of Blank and Sample with 100 mg MM12 (Continued)

Days	**Blank** Corrected Reading	Corrected StdDev	**Sample** Corrected Reading	Corrected StdDev
16.52	151.70	89.28	238.22	68.21
16.54	143.00	87.06	240.63	66.03
16.56	142.03	88.61	244.02	64.46
16.58	145.42	89.50	241.12	67.20
16.60	146.87	88.38	242.57	66.45
16.62	146.87	87.78	241.12	65.62
16.64	145.90	88.85	238.22	68.26
16.66	146.87	91.32	242.08	65.39
16.68	146.38	89.05	238.70	67.62
16.70	144.45	88.54	238.22	68.49
16.72	141.07	86.14	234.35	66.45
16.75	143.00	89.39	236.77	66.56
16.77	141.07	86.65	235.80	69.12
16.79	140.10	90.19	238.22	66.26
16.81	137.68	88.38	235.80	65.10
16.83	145.42	91.53	236.28	66.67
16.85	141.55	89.73	236.28	66.03
16.87	145.42	89.95	235.32	66.40
16.89	144.93	89.95	233.87	68.36
16.91	144.93	90.44	236.77	65.50
16.93	146.38	87.98	240.63	66.18
16.95	143.97	88.16	241.12	64.55
16.97	151.70	90.24	241.12	66.45
17.00	149.77	91.96	244.02	66.56
17.02	150.73	90.19	243.05	67.09
17.04	150.73	91.96	242.08	65.50
17.06	153.63	89.28	240.63	65.28
17.08	154.60	88.85	245.47	66.03
17.10	154.12	89.06	244.02	62.01
17.12	155.08	89.52	244.98	63.58
17.14	151.70	92.20	243.53	66.03
17.16	155.57	90.86	245.47	63.75
17.18	155.57	89.28	246.43	67.43
17.20	156.05	89.05	246.43	65.55
17.22	157.50	90.18	244.50	64.01
17.25	155.57	91.53	246.43	63.43
17.27	155.08	87.48	245.95	64.22
17.31	157.98	90.17	246.92	66.91

Days	Blank Corrected Reading	Corrected StdDev	Sample Corrected Reading	Corrected StdDev
17.37	158.47	89.75	247.40	63.47
17.39	158.95	86.18	248.85	64.22
17.41	159.43	89.99	244.50	66.18
17.43	162.82	89.39	246.92	67.31
17.45	154.60	87.71	243.05	62.65
17.47	157.98	87.96	243.05	63.69
17.50	154.12	88.38	241.12	66.56
17.52	154.12	85.47	241.60	66.45
17.54	151.22	89.72	242.57	65.62
17.56	152.18	88.38	239.67	63.37
17.58	152.67	88.18	239.67	63.91
17.60	150.73	89.72	238.70	62.53
17.62	152.18	89.07	242.57	64.22
17.64	148.32	88.16	238.70	64.64
17.66	150.25	89.06	237.25	65.39
17.68	147.83	84.85	235.80	66.03
17.70	154.12	86.86	238.22	63.58
17.72	155.57	91.10	243.53	64.37
17.75	157.02	86.68	245.95	66.99
17.77	159.92	88.16	247.40	64.64
17.79	161.37	90.18	243.05	65.50
17.81	161.85	88.03	250.78	61.67
17.83	165.23	87.36	249.33	62.27
17.85	166.20	88.29	250.30	65.62
17.87	166.20	88.60	249.82	65.38
17.89	166.68	89.13	251.75	64.87
17.91	166.68	86.43	249.82	66.68
17.93	164.75	88.71	255.13	65.34
17.95	165.72	89.28	254.17	59.95
17.97	164.27	88.88	256.10	65.24
18.00	165.72	87.93	254.17	64.87
18.02	167.65	89.13	254.17	64.73
18.04	166.68	89.95	256.58	65.92
18.06	166.20	87.78	253.68	64.11
18.08	168.62	88.14	255.62	64.34
18.10	167.65	86.86	255.13	64.73
18.12	168.62	84.16	259.48	64.34
18.16	170.07	86.59	258.52	64.22

Days	Blank Corrected Reading	Corrected StdDev	Sample Corrected Reading	Corrected StdDev
18.22	174.42	87.53	260.45	65.50
18.25	174.90	86.86	262.38	66.40
18.27	175.87	87.96	262.87	63.69
18.29	174.42	85.33	262.87	64.87
18.31	177.80	87.10	267.22	64.64
18.33	176.83	87.61	269.15	63.37
18.35	177.32	83.81	265.77	62.92
18.37	181.18	84.83	268.67	66.18
18.39	181.67	86.86	269.15	64.98
18.41	186.02	85.08	270.60	63.72
18.43	184.08	85.43	268.67	64.73
18.45	177.32	88.14	269.63	66.18
18.47	182.15	87.20	269.15	65.68
18.50	177.80	83.13	267.22	63.81
18.52	177.80	86.86	267.70	65.02
18.54	175.87	86.94	263.35	62.27
18.56	175.38	86.18	266.25	64.37
18.58	179.73	84.65	266.73	65.02
18.60	177.32	83.92	267.70	65.52
18.62	180.22	84.23	266.73	66.00
18.64	177.80	86.43	265.28	62.41
18.66	179.25	87.73	265.28	63.94
18.68	178.28	87.49	268.18	65.52
18.70	176.83	86.38	266.73	66.33
18.72	179.73	86.18	268.18	66.27
18.75	178.28	85.70	269.63	66.27
18.77	177.80	85.69	269.63	65.10
18.79	179.73	86.82	268.18	68.08
18.81	179.25	90.05	264.80	68.49
18.83	174.90	85.08	266.25	63.57
18.85	176.83	86.86	263.83	66.40
18.87	175.87	85.27	267.22	64.86
18.89	175.38	86.62	265.77	66.67
18.91	176.35	87.05	263.83	66.14
18.93	177.32	87.71	264.80	64.98
18.95	176.83	86.18	269.15	64.98
18.97	181.18	84.59	272.05	64.73
19.02	188.92	86.36	275.92	66.79

Days	Blank Corrected Reading	Corrected StdDev	Sample Corrected Reading	Corrected StdDev
19.08	188.43	84.83	280.27	65.62
19.10	190.37	86.81	281.72	68.34
19.12	191.33	82.78	280.27	68.08
19.14	195.20	84.59	283.65	66.27
19.16	196.65	83.39	282.68	66.91
19.18	194.72	84.80	282.20	66.03
19.20	193.75	86.26	283.17	68.34
19.22	195.20	84.80	284.62	67.05
19.25	201.00	86.14	284.13	65.89
19.27	198.10	86.82	287.03	67.20
19.29	198.10	85.92	287.52	66.03
19.31	198.10	85.52	287.52	67.84
19.33	196.65	85.47	285.58	67.09
19.35	192.78	85.27	284.13	68.36
19.37	191.33	83.46	277.37	68.72
19.39	191.33	89.28	278.82	69.50
19.41	191.82	85.03	278.82	68.34
19.43	188.92	87.26	278.82	66.91
19.45	184.57	81.95	273.50	67.33
19.47	184.08	84.65	270.60	65.34
19.50	178.77	86.85	270.12	67.56
19.52	180.22	84.35	271.57	68.26
19.54	180.22	84.80	268.18	68.60
19.56	176.35	83.68	263.83	69.64
19.58	175.38	84.83	266.25	67.43
19.60	172.97	83.73	266.73	66.68
19.62	177.32	83.30	265.28	65.52
19.64	173.45	86.43	262.38	67.43
19.66	172.00	85.47	260.93	67.05
19.68	171.52	86.18	263.83	65.38
19.70	172.48	88.65	263.35	65.52
19.72	176.35	87.05	263.83	66.79
19.75	172.97	83.81	261.42	65.92
19.77	173.93	83.23	265.28	68.60
19.79	171.52	82.34	262.38	65.10
19.81	172.97	84.36	263.83	68.72
19.83	178.28	84.13	265.77	68.48
19.87	173.45	81.92	267.22	67.05

Days	Blank Corrected Reading	Corrected StdDev	Sample Corrected Reading	Corrected StdDev
19.93	180.22	83.55	269.63	67.84
19.95	181.67	81.44	269.63	65.38
19.97	184.08	83.25	267.70	68.72
20.00	179.73	81.52	270.12	67.05
20.02	183.12	82.34	272.53	71.75
20.04	181.67	82.56	271.08	68.49
20.06	181.18	82.78	270.60	68.21
20.08	182.15	82.57	268.18	65.38
20.10	186.50	84.13	272.05	69.29
20.12	181.67	81.44	269.15	66.99
20.14	181.18	85.05	272.05	65.68
20.16	182.63	83.97	271.57	71.05
20.18	182.63	85.70	273.02	66.83
20.20	183.60	79.43	271.57	67.33
20.22	179.73	83.39	273.50	70.60
20.25	181.67	82.38	272.53	66.54
20.27	186.50	85.33	272.05	67.43
20.29	185.05	82.59	275.43	66.40
20.31	184.57	84.57	271.57	66.54
20.33	184.08	83.23	273.50	68.49
20.35	181.67	82.81	270.60	70.66
20.37	183.12	83.92	271.57	69.64
20.39	178.77	83.01	269.15	68.26
20.41	174.42	87.21	265.28	69.24
20.43	160.40	107.74	260.45	55.55
20.45	124.63	92.52	241.12	62.78
20.47	143.00	90.39	250.78	67.43
20.50	149.77	85.02	250.78	66.45
20.52	153.15	88.65	252.72	68.72
20.54	154.60	85.72	248.37	67.62
20.56	152.67	87.12	248.37	65.39
20.58	154.12	84.35	244.50	67.43
20.60	157.02	83.01	247.88	68.21
20.62	158.95	84.35	244.98	64.34
20.64	157.02	82.11	247.88	67.20
20.66	157.02	83.47	244.02	65.82
20.68	152.67	85.98	244.50	66.18
20.72	155.57	84.93	244.02	67.33

Table A.19. BOD of Blank and Sample with 100 mg MM12 (Continued)

Days	Blank Corrected Reading	Corrected StdDev	Sample Corrected Reading	Corrected StdDev
20.79	160.40	85.12	244.02	66.56
20.81	158.47	84.16	247.40	65.39
20.83	160.88	82.80	248.85	67.05
20.85	156.53	83.05	249.33	62.41
20.87	159.43	83.68	249.33	67.56
20.89	160.88	85.69	251.27	65.89
20.91	162.33	82.13	247.88	63.37
20.93	161.85	84.81	248.37	66.27
20.95	162.33	84.39	247.88	64.59
20.97	164.75	87.98	248.85	67.43
21.00	162.82	82.85	247.88	67.95
21.00	164.27	82.85	249.82	67.95
21.02	164.27	79.82	249.82	67.43
21.04	161.85	81.00	250.78	68.48
21.06	167.65	79.67	251.75	66.03
21.08	168.13	80.55	255.62	67.20
21.10	170.55	81.52	255.62	66.83
21.12	172.00	82.14	257.07	66.79
21.14	171.03	85.03	257.07	66.45
21.16	171.52	79.88	258.52	66.14
21.18	173.45	79.67	260.45	68.21
21.20	176.83	80.10	262.38	67.98
21.22	171.52	78.31	258.03	67.73
21.25	175.38	78.79	261.90	64.11
21.27	171.03	82.56	261.42	66.27
21.29	176.83	82.38	260.93	71.24
21.31	172.00	79.21	261.90	69.37
21.33	175.38	80.79	260.93	67.95
21.35	176.83	81.25	257.07	66.91
21.37	172.00	82.38	260.45	68.08
21.39	174.90	78.77	261.90	67.52
21.41	174.42	83.25	259.48	65.39
21.43	173.93	79.25	264.80	67.43
21.45	174.90	78.76	260.93	64.64
21.47	177.80	80.34	260.45	68.34
21.50	172.97	80.55	261.42	63.69
21.52	173.93	84.81	262.87	65.63
21.56	172.97	79.43	261.90	65.82

Days	Blank Corrected Reading	Corrected StdDev	Sample Corrected Reading	Corrected StdDev
21.62	174.90	82.34	263.83	67.56
21.64	175.87	77.64	261.42	69.64
21.66	174.90	80.79	261.90	66.14
21.68	177.32	82.34	263.35	68.16
21.70	176.83	81.67	261.90	66.45
21.72	175.38	76.79	262.87	67.56
21.75	173.93	79.46	262.87	66.35
21.77	172.97	81.89	262.38	64.75
21.79	174.90	83.46	260.93	68.08
21.81	175.38	81.89	261.90	66.14
21.83	170.55	82.59	263.35	64.86
21.85	174.42	80.34	259.97	66.40
21.87	175.87	82.13	261.42	66.03
21.89	173.45	80.86	265.77	65.39
21.91	175.87	77.54	260.45	66.45
21.93	173.45	82.13	261.42	66.56
21.95	174.90	78.79	260.45	65.10
21.97	175.38	78.09	265.28	62.13
22.00	175.38	80.32	261.42	67.31
22.02	175.38	81.05	264.80	62.78
22.04	175.38	82.85	262.38	64.86
22.06	175.87	76.75	258.52	67.05
22.08	174.90	83.01	262.38	67.43
22.10	175.87	81.00	263.83	65.89
22.12	171.52	82.14	260.93	66.67
22.14	175.38	80.55	260.93	69.00
22.16	174.90	79.65	265.28	64.73
22.18	173.45	81.00	260.93	66.40
22.20	174.90	82.14	262.87	66.56
22.22	173.93	78.54	263.35	70.28
22.25	175.38	80.10	259.97	65.18
22.27	175.87	83.01	258.03	66.45
22.29	177.80	81.00	261.42	65.28
22.31	173.45	80.55	262.87	69.24
22.33	173.93	77.86	258.52	64.01
22.35	174.42	79.21	260.93	64.98
22.37	177.32	81.89	261.90	65.39
22.41	174.42	80.77	262.87	67.31

Days	Blank Corrected Reading	Corrected StdDev	Sample Corrected Reading	Corrected StdDev
22.47	179.73	81.00	263.83	65.09
22.50	174.90	82.34	263.35	67.95
22.52	177.32	81.05	264.32	69.24
22.54	176.83	81.00	263.35	66.14
22.56	179.25	81.80	266.25	69.24
22.58	177.80	78.79	263.35	65.62
22.60	176.83	81.33	265.77	67.43
22.62	168.13	77.35	273.50	69.37
22.64	163.78	77.02	265.28	75.38
22.66	157.02	75.37	273.02	76.35
22.68	157.50	75.37	271.08	74.96
22.70	157.50	74.71	270.12	73.50
22.72	151.70	75.52	263.35	74.32
22.75	149.77	74.15	267.70	76.37
22.77	154.60	74.54	269.15	76.56
22.79	153.15	72.34	268.67	75.58
22.81	152.67	74.54	268.18	73.90
22.83	150.25	75.87	267.70	75.44
22.85	148.32	78.15	267.70	69.88
22.87	144.45	76.02	265.77	71.45
22.89	143.48	74.42	261.42	71.69
22.91	142.03	75.52	263.83	73.50
22.93	142.52	72.31	266.25	72.98
22.95	145.90	74.85	262.87	73.15
22.97	147.83	73.58	266.25	75.71
23.00	146.87	73.15	262.87	76.02
23.02	149.77	69.12	266.73	78.04
23.04	151.70	76.18	265.77	70.06
23.06	154.60	76.40	260.45	68.36
23.08	163.30	79.43	259.97	65.92
23.10	165.23	77.42	255.13	67.64
23.12	167.65	79.65	252.72	61.89
23.14	166.20	80.79	254.65	64.59
23.16	167.17	81.10	253.20	64.34
23.18	166.20	81.33	256.58	62.13
23.20	168.13	79.43	258.52	65.39
23.22	166.68	80.10	253.20	66.14
23.27	169.10	80.13	258.03	66.27

Table A.19. BOD of Blank and Sample with 100 mg MM12 (Continued)

Days	Blank Corrected Reading	Corrected StdDev	Sample Corrected Reading	Corrected StdDev
23.33	165.72	77.86	257.07	66.26
23.35	171.52	79.82	254.65	66.56
23.37	170.55	79.46	259.00	66.14
23.39	170.07	82.11	260.45	67.95
23.41	174.42	79.92	259.97	68.36
23.43	171.03	79.21	259.97	68.07
23.45	171.52	82.39	262.87	66.14
23.47	173.45	79.67	261.90	65.62
23.50	173.93	78.50	259.97	66.03
23.52	173.93	80.79	262.87	66.79
23.54	172.48	76.59	262.87	68.72
23.56	172.97	76.30	261.42	70.01
23.58	175.38	79.72	261.90	66.67
23.60	175.87	78.33	265.28	67.05
23.62	177.32	78.33	265.28	66.27
23.64	174.42	77.46	265.28	65.75
23.66	175.87	77.46	264.80	66.03
23.68	175.87	79.65	261.42	67.05
23.70	172.00	81.44	260.93	67.43
23.72	171.03	80.13	259.48	66.45
23.75	170.55	78.76	257.07	65.39
23.77	172.00	76.08	257.55	64.87
23.79	171.52	80.17	258.52	67.52
23.81	167.65	78.59	255.13	67.09
23.83	162.82	80.13	252.23	65.82
23.85	161.85	80.34	256.10	63.58
23.87	165.72	82.34	253.68	64.08
23.89	156.53	77.97	246.92	65.50
23.91	161.85	79.67	249.33	62.01
23.93	165.23	79.68	250.78	64.64
23.95	162.82	78.14	247.40	65.39
23.97	164.75	77.64	248.85	67.19
24.00	161.85	75.87	249.82	65.28
24.02	165.23	79.44	248.37	64.22
24.04	166.20	79.43	253.20	63.69
24.06	166.68	78.09	252.72	64.86
24.08	169.58	77.64	254.17	65.62
24.12	172.00	79.67	259.00	65.10

Table A.19. BOD of Blank and Sample with 100 mg MM12 (Continued)

Days	Blank Corrected Reading	Corrected StdDev	Sample Corrected Reading	Corrected StdDev
24.18	172.48	77.20	258.52	71.05
24.20	174.42	78.20	258.03	65.50
24.22	173.93	75.87	259.48	68.89
24.25	172.48	77.73	260.45	67.09
24.27	174.42	79.88	260.93	67.43
24.29	172.97	79.68	260.93	65.09
24.31	174.42	79.44	261.90	66.27
24.33	174.90	81.44	259.48	64.01
24.35	173.93	79.46	260.93	65.92
24.37	172.97	76.08	262.38	64.98
24.39	175.87	78.54	259.00	64.86
24.41	175.38	78.09	261.42	65.92
24.43	176.83	78.09	259.48	65.92
24.45	177.32	77.87	260.93	63.69
24.47	177.80	77.86	258.52	65.39
24.50	178.77	79.21	263.35	67.20
24.52	175.87	76.98	262.87	67.31
24.54	175.38	77.42	263.35	63.58
24.56	178.28	80.59	260.93	66.80
24.58	177.32	77.42	262.38	65.10
24.60	179.73	78.77	268.18	67.43
24.62	172.97	78.77	262.38	65.24
24.64	172.97	80.19	257.07	65.38
24.66	157.98	110.70	251.75	44.84
24.68	107.23	88.37	221.30	57.87
24.70	143.48	82.71	238.22	56.35
24.72	152.18	81.48	241.12	60.08
24.75	160.88	79.65	247.88	58.39
24.77	164.75	76.75	251.75	60.08
24.79	161.85	77.87	250.78	64.01
24.81	161.85	80.55	252.23	60.84
24.83	166.20	78.54	252.23	66.35
24.85	169.10	77.66	253.68	64.22
24.87	168.13	76.88	253.20	64.30
24.89	170.07	77.86	255.62	64.75
24.91	170.07	77.19	255.13	64.87
24.93	168.13	76.97	259.97	65.24
24.97	171.03	79.21	259.97	63.58

Table A.19. BOD of Blank and Sample with 100 mg MM12 (Continued)

Days	Blank Corrected Reading	Corrected StdDev	Sample Corrected Reading	Corrected StdDev
25.04	170.07	77.64	256.58	65.39
25.06	173.45	77.18	260.45	66.79
25.08	172.97	76.79	258.52	66.56
25.10	175.38	76.79	261.90	67.98
25.12	173.93	76.79	259.97	67.20
25.14	173.93	76.75	259.97	63.28
25.16	173.45	80.34	259.48	67.69
25.18	171.52	79.88	260.45	66.14
25.20	170.55	77.87	259.00	65.28
25.22	170.55	78.59	259.97	65.75
25.25	172.97	79.21	259.00	62.41
25.27	170.07	78.33	256.58	68.48
25.29	170.07	79.72	257.55	65.92
25.31	171.52	79.53	256.58	67.95
25.33	172.00	76.21	257.07	68.07
25.35	173.93	79.06	257.07	66.56
25.37	175.38	78.39	260.45	68.26
25.39	172.00	77.54	258.03	63.47
25.41	172.00	81.25	259.00	63.20
25.43	172.00	75.26	263.35	66.14
25.45	174.90	79.21	259.00	68.26
25.47	172.97	83.10	259.48	68.16
25.50	173.45	75.20	261.90	67.09
25.52	178.77	77.42	262.38	66.35
25.54	176.83	78.54	266.73	65.82
25.56	176.83	78.12	261.90	67.20
25.58	180.70	74.79	265.77	65.63
25.60	181.18	79.46	260.93	64.86
25.62	172.97	79.44	262.87	64.22
25.64	170.55	82.47	259.00	69.12
25.66	163.78	103.46	252.72	53.00
25.68	116.42	83.52	225.17	58.45
25.70	155.08	81.03	247.40	61.89
25.72	159.43	78.77	248.85	62.20
25.75	159.92	78.79	249.82	62.74
25.77	162.33	80.32	248.85	64.01
25.79	165.23	76.33	252.23	64.98
25.83	171.03	74.53	255.62	65.28

Days	**Blank** Corrected Reading	Corrected StdDev	**Sample** Corrected Reading	Corrected StdDev
25.89	171.52	73.39	260.93	62.92
25.91	167.65	78.09	254.17	60.20
25.93	170.07	76.99	255.62	61.36
25.95	170.55	77.42	257.07	62.01
25.97	170.55	78.77	258.03	63.69
26.00	170.07	76.75	259.97	61.77
26.02	168.62	75.85	255.62	62.20
26.04	167.17	74.96	255.62	66.40
26.06	174.42	78.20	259.97	65.82
26.08	171.03	75.66	257.55	65.63
26.10	171.03	78.54	260.93	66.72
26.12	172.48	77.87	263.35	64.11
26.14	173.45	76.12	258.52	66.72
26.16	173.45	80.10	263.35	63.94
26.18	175.87	78.09	260.93	64.59
26.20	178.77	77.46	261.90	66.99
26.22	174.42	78.98	264.80	66.03
26.25	179.25	77.92	265.77	67.90
26.27	178.28	77.67	269.15	71.20
26.29	180.70	75.86	268.67	72.76
26.31	176.35	75.21	270.12	73.04
26.33	175.38	72.25	274.47	73.61
26.35	174.90	72.47	274.95	77.73
26.37	178.28	72.47	273.02	77.76
26.39	176.83	72.86	275.92	74.28
26.41	176.83	72.10	279.30	77.82
26.43	181.18	70.95	277.85	79.62
26.45	179.73	70.28	278.82	78.72
26.47	180.70	72.73	279.30	78.09
26.50	181.67	71.69	279.78	78.94
26.52	186.98	72.34	283.17	80.24
26.54	183.12	73.26	281.23	74.09
26.56	185.53	67.33	284.62	78.64
26.58	183.12	70.66	283.17	78.36
26.60	180.70	71.31	281.23	76.02
26.62	178.28	70.80	285.58	76.08
26.64	175.87	70.60	278.82	76.88
26.68	122.22	75.20	252.23	74.02

Days	Blank Corrected Reading	Corrected StdDev	Sample Corrected Reading	Corrected StdDev
26.75	167.17	74.60	267.70	73.04
26.77	162.82	76.40	265.77	70.81
26.79	167.65	74.05	264.80	69.33
26.81	168.62	73.57	261.90	68.36
26.83	169.10	72.95	255.62	64.37
26.85	167.17	74.06	254.65	65.10
26.87	169.58	75.87	252.23	65.18
26.89	163.30	76.08	250.78	64.01
26.91	166.68	74.96	252.23	64.98
26.93	165.23	75.41	250.30	63.30
26.95	165.23	76.31	251.27	62.94
26.97	166.20	75.41	249.82	64.75
27.00	165.23	77.66	249.33	62.84
27.02	167.17	74.74	253.68	63.37
27.04	164.75	74.74	251.27	64.22
27.06	169.10	74.52	254.65	63.58
27.08	168.62	74.06	253.68	64.22
27.10	168.13	77.66	253.20	64.55
27.12	168.13	75.85	252.72	64.01
27.14	168.62	74.79	252.23	65.72
27.16	166.68	74.96	249.82	67.09
27.18	168.62	77.20	250.78	66.80
27.20	170.07	76.40	252.23	66.99
27.22	169.10	77.18	254.65	65.41
27.25	168.62	78.59	253.68	66.56
27.27	165.72	75.46	254.65	66.79
27.29	168.62	74.60	255.62	64.01
27.31	168.62	75.26	255.13	62.65
27.33	170.55	74.79	255.62	64.46
27.35	172.48	73.75	258.03	64.55
27.37	174.42	76.97	257.07	66.26
27.39	174.42	75.26	256.10	62.88
27.41	170.55	75.07	258.03	62.93
27.43	173.93	77.66	259.97	63.83
27.45	175.38	75.93	255.62	60.69
27.47	173.93	78.20	256.58	63.83
27.50	176.83	78.50	259.97	67.35
27.54	176.35	75.93	261.90	66.26

Table A.19. BOD of Blank and Sample with 100 mg MM12 (Continued)

Days	**Blank** Corrected Reading	Corrected StdDev	**Sample** Corrected Reading	Corrected StdDev
27.60	174.90	79.06	259.48	62.94
27.62	173.93	78.76	261.42	60.50
27.64	170.55	76.21	258.03	62.49
27.66	173.93	75.46	257.55	63.28
27.68	171.52	77.42	258.03	61.77
27.70	169.58	75.87	254.17	63.47
27.72	170.55	77.66	254.65	61.48
27.75	165.23	74.33	251.27	66.26
27.77	167.17	76.08	251.75	64.98
27.79	163.78	73.09	251.75	65.82
27.81	166.68	75.74	248.85	63.47
27.83	164.27	75.66	250.30	63.12
27.85	165.23	75.40	249.82	64.01
27.87	164.75	76.53	252.23	65.50
27.89	166.68	76.99	253.68	65.63
27.91	167.65	76.97	255.13	65.50
27.93	170.07	74.89	255.13	64.64
27.95	170.07	77.54	259.00	62.41
27.97	170.55	74.53	260.93	57.96
28.00	165.23	77.26	258.52	61.87
28.02	169.10	73.16	256.58	62.65
28.04	169.58	72.99	254.65	60.93
28.06	164.75	73.83	250.78	60.69
28.08	170.07	77.87	250.78	62.65
28.10	166.20	76.12	254.17	62.41
28.12	166.20	74.89	256.58	64.46
28.14	164.27	76.99	254.17	63.37
28.16	167.65	74.53	254.17	65.10
28.18	168.13	75.26	253.68	66.56
28.20	169.10	73.94	253.20	67.69
28.22	169.58	73.39	256.10	64.64
28.25	169.10	76.03	251.75	67.52
28.27	170.07	74.96	252.23	64.46
28.29	169.58	77.84	252.72	65.00
28.31	170.55	75.26	251.27	64.55
28.33	169.10	77.73	252.72	66.56
28.35	168.62	80.78	254.65	67.95
28.39	170.55	75.26	256.10	67.43

Days	Blank Corrected Reading	Corrected StdDev	Sample Corrected Reading	Corrected StdDev
28.45	173.45	77.35	256.10	65.09
28.47	169.58	76.52	254.65	63.67
28.50	172.48	77.73	256.10	65.00
28.52	172.97	78.50	254.17	63.05
28.54	172.48	74.33	257.55	62.84
28.56	172.97	77.35	258.03	67.43
28.58	172.48	77.87	256.10	64.11
28.60	172.00	75.26	257.07	67.09
28.62	167.65	74.60	253.68	63.05
28.64	164.27	73.72	252.23	63.12
28.66	162.33	103.57	252.72	55.14
28.68	111.58	82.81	226.13	57.75
28.70	146.87	80.79	242.08	61.81
28.72	155.08	79.67	247.40	59.76
28.75	159.43	76.79	244.98	61.03
28.77	163.78	77.46	248.37	61.13
28.79	159.92	74.41	248.85	61.36
28.81	160.40	77.20	246.92	60.85
28.83	158.47	75.93	245.95	60.77
28.85	163.78	78.59	246.92	62.84
28.87	165.72	78.32	250.78	63.05
28.89	167.17	74.53	253.20	61.13
28.91	166.68	76.40	251.75	64.64
28.93	170.55	75.87	256.10	63.37
28.95	164.75	71.00	254.17	62.65
28.97	160.40	72.33	252.72	63.17
29.00	161.37	72.28	259.48	61.76
29.02	162.33	72.73	254.17	61.36
29.04	164.75	74.29	255.62	63.28
29.06	171.03	72.33	249.33	63.12
29.08	165.72	74.33	249.82	64.46
29.10	168.62	73.20	255.13	65.00
29.12	168.62	75.46	251.27	61.24
29.14	167.17	74.60	247.88	66.56
29.16	170.07	75.74	248.85	63.37
29.18	168.62	75.07	249.82	60.85
29.20	171.03	73.57	253.20	63.58
29.25	171.03	72.95	254.17	66.80

Table A.19. BOD of Blank and Sample with 100 mg MM12 (Continued)

Days	Blank Corrected Reading	Corrected StdDev	Sample Corrected Reading	Corrected StdDev
29.31	167.65	75.93	253.20	63.91
29.33	169.58	77.66	251.75	65.82
29.35	171.03	74.07	254.65	62.99
29.37	171.52	76.12	253.20	64.86
29.39	172.00	75.26	255.62	64.86
29.41	169.10	75.46	255.62	62.43
29.43	170.07	76.59	252.72	63.50
29.45	170.07	76.59	259.00	63.12
29.47	170.07	77.66	255.62	62.65
29.50	173.93	75.41	257.07	63.91
29.52	175.38	76.08	259.00	65.28
29.54	172.97	77.66	257.07	65.82
29.56	172.48	76.79	257.55	63.05
29.58	175.38	74.89	259.48	62.37
29.60	174.42	76.59	255.13	62.01
29.62	171.52	74.23	260.93	59.76
29.64	173.93	77.26	256.58	64.11
29.66	173.45	75.07	255.62	61.48
29.68	173.93	77.26	257.07	63.67
29.70	173.45	74.13	257.55	61.39
29.72	169.10	75.69	253.68	60.57
29.75	169.58	76.59	254.17	61.77
29.77	171.52	75.66	257.07	65.28
29.79	168.13	74.79	255.62	61.32
29.81	169.58	74.74	253.20	61.57
29.83	170.55	76.79	256.10	68.70
29.85	168.13	76.12	252.23	65.28
29.87	169.10	73.62	255.62	65.18
29.89	172.00	74.96	253.68	63.37
29.91	168.62	73.66	257.07	61.39
29.93	169.58	72.72	255.13	66.35
29.95	168.62	74.41	253.68	63.94
29.97	167.65	75.26	256.58	62.74
30.00	164.75	76.33	254.65	64.75
30.02	164.27	74.60	250.78	64.22
30.04	165.72	74.29	253.20	64.64
30.06	165.23	72.61	251.75	63.05
30.10	168.62	74.41	255.13	65.89

Table A.19. BOD of Blank and Sample with 100 mg MM12 (Continued)

Days	Blank		Sample	
	Corrected Reading	Corrected StdDev	Corrected Reading	Corrected StdDev
30.16	170.07	72.08	257.07	65.50
30.18	171.52	71.47	256.58	65.09
30.20	171.52	76.03	256.10	63.05
30.22	170.07	75.93	257.55	66.56
30.25	172.48	72.08	256.10	64.22
30.27	174.42	71.76	259.00	64.34
30.29	174.42	75.07	255.13	65.50
30.31	172.48	74.33	256.58	67.05
30.33	174.42	72.33	256.58	66.14
30.35	174.42	73.83	259.00	62.78
30.37	174.42	71.86	260.93	65.52
30.39	174.90	73.57	262.38	66.14
30.41	172.00	76.69	259.00	63.81
30.43	174.90	74.23	261.90	65.82
30.45	175.38	74.74	257.55	64.46
30.47	176.35	74.79	260.93	62.74
30.50	173.93	74.53	257.55	64.86
30.52	175.38	76.33	267.22	64.08
30.54	178.77	73.94	264.32	61.89
30.56	175.38	74.54	264.80	61.77
30.58	173.93	73.57	261.42	65.24
30.60	176.35	74.37	264.32	64.75
30.62	174.42	73.22	260.93	64.08
30.64	178.77	75.20	264.32	61.87
30.66	172.97	83.55	262.38	58.69
30.68	158.47	76.53	252.72	63.37
30.70	168.13	75.87	255.62	60.60
30.72	169.10	76.40	255.62	63.47
30.75	167.65	74.41	256.10	64.55
30.77	172.00	73.86	254.17	63.83
30.79	170.55	71.47	252.72	63.69
30.81	170.07	72.07	255.62	62.41
30.83	170.07	72.61	255.13	63.58
30.85	168.62	74.23	255.13	64.55
30.87	168.13	74.96	251.27	64.01
30.89	165.72	72.07	253.68	64.55
30.91	168.62	73.66	251.27	62.11
30.95	166.20	71.66	254.17	60.22

Table A.19. BOD of Blank and Sample with 100 mg MM12 (Continued)

Days	Blank Corrected Reading	Corrected StdDev	Sample Corrected Reading	Corrected StdDev
31.02	169.10	73.20	250.30	63.83
31.04	164.27	72.33	249.82	61.77
31.06	168.62	74.99	254.17	65.09
31.08	164.75	72.95	251.75	63.20
31.10	165.72	73.20	251.75	61.62
31.12	164.27	70.73	255.13	62.30
31.14	163.30	71.91	251.75	61.36
31.16	163.30	72.61	254.17	64.30
31.18	167.17	74.23	249.82	62.94
31.20	165.23	73.75	254.17	65.24
31.22	171.52	75.20	257.07	59.51
31.25	170.07	73.27	250.78	62.20
31.27	170.55	74.89	258.52	64.64
31.29	175.87	73.75	256.10	66.14
31.31	170.07	71.47	256.10	64.22
31.33	175.87	75.26	256.10	62.30
31.35	172.00	71.76	257.55	62.84
31.37	175.87	74.23	255.13	61.57
31.39	175.38	72.07	259.00	64.22
31.41	174.42	73.86	254.65	60.77
31.43	176.35	74.23	259.97	62.11
31.45	173.93	73.57	262.38	60.50
31.47	174.90	74.23	263.83	59.95
31.50	175.38	76.79	260.45	60.85

A.3.7. METAC Kinetics

Table A.20. P Removal by MM12 over Varying Times of Exposure

Time (min)	$C_0 = 0.1$ mg/L Normalized Conc.	Standard Deviation	$C_0 = 50$ mg/L Normalized Conc.	Standard Deviation	$C_0 = 5$ mg/L Normalized Conc.	Standard Deviation
1440	0.28	0.02	0.27	0.03	0.08	0.00
1080	0.28	0.02	0.26	0.01	0.08	0.00
720	0.26	0.03	0.26	0.00	0.08	0.02
360	0.24	0.02	0.26	0.00	0.08	0.02
180	0.23	0.00	0.26	0.00	0.08	0.00
120	0.22	0.02	0.27	0.02	0.08	0.00
60	0.22	0.02	0.27	0.01	0.08	0.02
30	0.21	0.02	0.26	0.01	0.09	0.02
15	0.20	0.00	0.27	0.00	0.08	0.00
10	0.22	0.02	0.26	0.00	0.08	0.01
8	0.21	0.02	0.26	0.00	0.08	0.01
6	0.20	0.00	0.26	0.00	0.08	0.00
4	0.22	0.02	0.26	0.02	0.08	0.01
0	1.00	0.00	1.00	0.00	1.00	0.00

A.3.8. METAC Behavior in Eutrophic Lake Samples

Table A.21. P Removal by MM12 from Eutrophic Lakes and Low Conc. Synthetic Lab Water

Source	Sample	Absorbance (nm)	Conc.	Dilution Factor	Adj. Conc ug/L	Average	StdDev	Sorbed Phosphate (mg/L)	Normalized	Normalized StdDev
	1	0.002	0.00	5.00	13.52	22.53	10.32	128.41	0.85	0.07
	2	0.005	0.01	5.00	33.79					
	3	0.003	0.00	5.00	20.28					
	C	0.022	0.03	5.00	148.69	150.94	17.01			
	C	0.02	0.03	5.00	135.17					
Synthetic	C	0.025	0.03	5.00	168.96					
	1	0.008	0.01	5.00	54.07	49.56	7.80	135.17	0.73	0.04
	2	0.006	0.01	5.00	40.55					
	3	0.008	0.01	5.00	54.07					
	KC	0.028	0.04	5.00	189.24	184.73	7.80			
	KC	0.026	0.04	5.00	175.72					
Katrina	KC	0.028	0.04	5.00	189.24					
	1	0.008	0.01	5.00	54.07	51.82	3.90	128.41	0.71	0.02
	2	0.008	0.01	5.00	54.07					
	3	0.007	0.01	5.00	47.31					
	HNC	0.026	0.04	5.00	175.72	180.23	3.90			
	HNC	0.027	0.04	5.00	182.48					
Holy Name	HNC	0.027	0.04	5.00	182.48					

Table A.21. P Removal by MM12 from Eutrophic Lakes and Low Conc. Synthetic Lab Water (Continued)

Source	Sample	Absorbance (nm)	Conc.	Dilution Factor	Adj. Conc ug/L	Average	StdDev	Sorbed Phosphate (mg/L)	Normalized	Normalized StdDev
	1	0.007	0.01	5.00	47.31	47.31	0.00	164.46	0.78	0.00
	2	0.007	0.01	5.00	47.31					
	3	0.007	0.01	5.00	47.31					
	AC	0.031	0.04	5.00	209.52	211.77	17.01			
	AC	0.034	0.05	5.00	229.79					
Ardmore	AC	0.029	0.04	5.00	196.00					
	1	0.009	0.01	5.00	60.83	51.82	10.32	135.17	0.72	0.06
	2	0.006	0.01	5.00	40.55					
	3	0.008	0.01	5.00	54.07					
	IC	0.025	0.03	5.00	168.96	186.99	15.61			
	IC	0.029	0.04	5.00	196.00					
Independence	IC	0.029	0.04	5.00	196.00					
	1	0.007	0.01	5.00	47.31	42.80	3.90	114.90	0.73	0.02
	2	0.006	0.01	5.00	40.55					
	3	0.006	0.01	5.00	40.55					
	HMC	0.022	0.03	5.00	148.69	157.70	7.80			
	HMC	0.024	0.03	5.00	162.21					
Half Moon	HMC	0.024	0.03	5.00	162.21					

A.3.9. METAC Temperature Study

Table A.22. P Removal Data by MM12 Under Varying Temperature Conditions

Temp (°C)	Normalized P Removal	Normalized StdDev
4	0.919	0.015
10	0.935	0.000
15	0.915	0.011
20	0.937	0.015
25	0.928	0.007
30	0.929	0.014
35	0.941	0.008

A.3.10. METAC Partial-Selectivity Study

Table A.23. Anion Removal by MM12 (Anion C_0 = 110 meq/L)

Anion	Pre-PO_4^{3-} Introduction		Post-PO_4^{3-} Introduction	
	Amount Sorbed (meq)	StdDev	Amount Sorbed (meq)	StdDev
NO_3^-	0.340	0.050	0.048	0.002
SO_4^{2-}	0.350	0.038	0.052	0.007
Cl^-	0.337	0.031	0.043	0.005

Table A.24. P Removal by MM12 after Anion Saturation (Anion C_0 = 110 meq/L) (P C_0 = 110 meq/L)

Anion Present	Post-Anion Introduction	
	P Sorbed mg (PO_4^{3-} meq)	StdDev mg (PO_4^{3-} meq)
NO_3^-	16.94 (0.535)	1.67 (0.053)
SO_4^{2-}	16.64 (0.526)	1.16 (0.037)
Cl^-	17.35 (0.548)	2.28 (0.072)

A.3.11. METAC Statistical Analysis

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```

One-way ANOVA: % Removal versus pH

```
Method

Null hypothesis        All means are equal
Alternative hypothesis  At least one mean is different
Significance level     α = 0.05

Equal variances were assumed for the analysis.


Factor Information

Factor  Levels  Values
pH          10  10, 11, 12, 3, 4, 5, 6, 7, 8, 9


Analysis of Variance

Source  DF  Adj SS  Adj MS  F-Value  P-Value
pH       9   999.3  111.03     7.69    0.000
Error   20   288.9   14.45
Total   29  1288.2


Model Summary

      S    R-sq  R-sq(adj)  R-sq(pred)
3.80088  77.57%     67.48%      49.53%


Means

pH  N    Mean  StDev        95% CI
10  3   29.27   1.92  ( 24.69,  33.85)
11  3  27.438  1.143  (22.861, 32.016)
12  3   27.69   2.41  ( 23.11,  32.27)
3   3   21.31   8.76  ( 16.73,  25.88)
4   3   19.00   5.50  ( 14.43,  23.58)
5   3   24.45   3.73  ( 19.87,  29.03)
6   3  35.849  1.303  (31.271, 40.426)
7   3   36.60   1.99  ( 32.02,  41.18)
8   3   35.74   1.97  ( 31.16,  40.32)
9   3   29.24   1.78  ( 24.66,  33.82)

Pooled StDev = 3.80088
```

Tukey Pairwise Comparisons

```
Grouping Information Using the Tukey Method and 95% Confidence
```

```
pH  N    Mean  Grouping
7   3   36.60  A
6   3   35.849 A
8   3   35.74  A
10  3    29.27 A B
9   3    29.24 A B
12  3    27.69 A B
11  3   27.438 A B
5   3    24.45   B
3   3    21.31   B
4   3    19.00   B
```

Means that do not share a letter are significantly different.

Tukey Simultaneous 95% CIs

Interval Plot of % Removal vs pH

One-way ANOVA: % Removal versus pH

```
Method
```

```
Null hypothesis          All means are equal
Alternative hypothesis   At least one mean is different
Significance level       α = 0.05
```

Equal variances were assumed for the analysis.

```
Factor Information
```

```
Factor  Levels  Values
pH          10   10, 11, 12, 3, 4, 5, 6, 7, 8, 9
```

```
Analysis of Variance
```

```
Source  DF  Adj SS   Adj MS  F-Value  P-Value
pH       9  429.94  47.7707    90.63    0.000
Error   20   10.54   0.5271
Total   29  440.48
```

```
Model Summary
```

```
       S    R-sq  R-sq(adj)  R-sq(pred)
0.725994  97.61%     96.53%      94.62%
```

```
Means
```

```
pH  N    Mean  StDev        95% CI
10  3  94.543  0.769  (93.668, 95.417)
11  3  95.130  0.437  (94.256, 96.004)
12  3  95.142  0.872  (94.268, 96.017)
3   3  81.348  0.822  (80.473, 82.222)
4   3  91.941  0.831  (91.067, 92.816)
```

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```
5   3  93.471  0.939  (92.596, 94.345)
6   3  93.236  0.467  (92.362, 94.111)
7   3  92.381  0.920  (91.507, 93.256)
8   3  92.016  0.450  (91.141, 92.890)
9   3  92.299  0.465  (91.425, 93.174)

Pooled StDev = 0.725994
```

Tukey Pairwise Comparisons

```
Grouping Information Using the Tukey Method and 95% Confidence

pH  N    Mean  Grouping
12  3  95.142  A
11  3  95.130  A
10  3  94.543  A
5   3  93.471  A B
6   3  93.236  A B
7   3  92.381    B
9   3  92.299    B
8   3  92.016    B
4   3  91.941    B
3   3  81.348       C

Means that do not share a letter are significantly different.
```

Tukey Simultaneous 95% CIs

Interval Plot of % Removal vs pH

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```
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Retrieving project from file: 'E:\Research\MS\METAC\Statistical
Analysis.MPJ'
```

One-way ANOVA: % Removal2 versus Water

```
Method

Null hypothesis         All means are equal
Alternative hypothesis  At least one mean is different
Significance level      α = 0.05

Equal variances were assumed for the analysis.


Factor Information

Factor  Levels  Values
Water        7  Adrmore, Ardmore, Half Moon, Holy Name, Independence, Katrina,
Synthetic
```

```
Analysis of Variance

Source  DF  Adj SS  Adj MS  F-Value  P-Value
Water    6   411.1   68.52     3.56    0.033
Error   11   211.8   19.26
Total   17   623.0


Model Summary

      S    R-sq  R-sq(adj)  R-sq(pred)
4.38843  65.99%     47.45%           *


Means

Water          N   Mean  StDev       95% CI
Adrmore        1  77.66      *  (68.00, 87.32)
Ardmore        2  77.66   0.00  (70.83, 84.49)
Half Moon      3  72.86   2.47  (67.28, 78.43)
Holy Name      3  71.25   2.17  (65.67, 76.83)
Independence   3  72.29   5.52  (66.71, 77.87)
Katrina        3  73.17   4.22  (67.59, 78.75)
Synthetic      3  85.07   6.84  (79.50, 90.65)


Pooled StDev = 4.38843
```

Tukey Pairwise Comparisons

```
Grouping Information Using the Tukey Method and 95% Confidence

Water          N   Mean  Grouping
Synthetic      3  85.07  A
Ardmore        2  77.66  A B
Adrmore        1  77.66  A B
Katrina        3  73.17  A B
Half Moon      3  72.86  A B
Independence   3  72.29    B
Holy Name      3  71.25    B


Means that do not share a letter are significantly different.
```

Tukey Simultaneous 95% CIs

Interval Plot of % Removal2 vs Water

One-way ANOVA: % RemovalT versus Temp

```
Method

Null hypothesis         All means are equal
Alternative hypothesis  At least one mean is different
Significance level      α = 0.05
```

```
Equal variances were assumed for the analysis.


Factor Information

Factor  Levels  Values
Temp         7  4, 10, 15, 20, 25, 30, 35


Analysis of Variance

Source  DF  Adj SS  Adj MS  F-Value  P-Value
Temp     6   16.07   2.678     2.07    0.123
Error   14   18.10   1.293
Total   20   34.17


Model Summary

       S    R-sq  R-sq(adj)  R-sq(pred)
1.13690  47.04%     24.34%       0.00%


Means

Temp  N    Mean  StDev         95% CI
4     3  91.880  1.490  (90.472, 93.288)
10    3   93.47   0.00  ( 92.06,  94.87)
15    3  91.530  1.119  (90.123, 92.938)
20    3  93.742  1.521  (92.335, 95.150)
25    3  92.838  0.723  (91.430, 94.246)
30    3  92.907  1.433  (91.499, 94.314)
35    3  94.087  0.829  (92.679, 95.494)

Pooled StDev = 1.13690
```

Tukey Pairwise Comparisons

```
Grouping Information Using the Tukey Method and 95% Confidence

Temp  N    Mean  Grouping
35    3  94.087  A
20    3  93.742  A
10    3   93.47  A
30    3  92.907  A
25    3  92.838  A
4     3  91.880  A
15    3  91.530  A

Means that do not share a letter are significantly different.
```

Tukey Simultaneous 95% CIs

Interval Plot of % RemovalT vs Temp

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One-way ANOVA: BOD versus Time

```
* NOTE * Cannot draw the interval plot for the Tukey procedure. Interval plots for
         comparisons are illegible with more than 45 intervals.


Method

Null hypothesis          All means are equal
Alternative hypothesis   At least one mean is different
Significance level       α = 0.05

Equal variances were assumed for the analysis.


Factor Information

Factor  Levels  Values
Time        12  10B, 10S, 15B, 15S, 20B, 20S, 25B, 25S, 30B, 30S, 5B, 5S


Analysis of Variance

Source  DF  Adj SS  Adj MS  F-Value  P-Value
Time    11   39235    3567     0.97    0.499
Error   24   88414    3684
Total   35  127649


Model Summary

      S    R-sq  R-sq(adj)  R-sq(pred)
60.6953  30.74%      0.00%       0.00%


Means

Time  N   Mean  StDev       95% CI
10B   3  125.0   70.8  ( 52.7, 197.3)
10S   3  178.7   53.6  (106.3, 251.0)
15B   3  119.0   73.7  ( 46.7, 191.3)
15S   3  184.7   55.5  (112.3, 257.0)
20B   3  141.3   68.9  ( 69.0, 213.7)
20S   3  203.0   56.6  (130.7, 275.3)
25B   3  132.7   66.3  ( 60.3, 205.0)
25S   3  193.7   54.9  (121.3, 266.0)
30B   3  128.3   64.5  ( 56.0, 200.7)
30S   3  188.0   54.7  (115.7, 260.3)
5B    3   99.0   54.3  ( 26.7, 171.3)
5S    3  160.0   48.4  ( 87.7, 232.3)

Pooled StDev = 60.6953
```

Tukey Pairwise Comparisons

```
Grouping Information Using the Tukey Method and 95% Confidence

Time  N   Mean  Grouping
20S   3  203.0  A
25S   3  193.7  A
```

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```
30S   3  188.0  A
15S   3  184.7  A
10S   3  178.7  A
5S    3  160.0  A
20B   3  141.3  A
25B   3  132.7  A
30B   3  128.3  A
10B   3  125.0  A
15B   3  119.0  A
5B    3   99.0  A
```

Means that do not share a letter are significantly different.

Interval Plot of BOD vs Time

One-way ANOVA: Sorption (mg) versus Polymer

```
* NOTE * Cannot draw the interval plot for the Tukey procedure. Interval plots for
         comparisons are illegible with more than 45 intervals.


Method

Null hypothesis          All means are equal
Alternative hypothesis  At least one mean is different
Significance level       α = 0.05

Equal variances were assumed for the analysis.


Factor Information

Factor    Levels  Values
Polymer       12  2500 MM, 2500 MM12, 2500 MM4, 2500 MN, 3000 MM, 3000 MM12, 3000 MM4,
3000
                  MN, 3500 MM, 3500 MM12, 3500 MM4, 3500 MN


Analysis of Variance

Source   DF   Adj SS   Adj MS  F-Value  P-Value
Polymer  11  1426.05  129.641    31.59    0.000
Error    24    98.48    4.103
Total    35  1524.53


Model Summary

      S    R-sq  R-sq(adj)  R-sq(pred)
2.02568  93.54%     90.58%      85.47%


Means

Polymer      N    Mean  StDev       95% CI
2500 MM      3  11.710  0.867  ( 9.296, 14.123)
2500 MM12    3   22.20   2.21  ( 19.78,  24.61)
2500 MM4     3   16.12   2.24  ( 13.71,  18.54)
2500 MN      3   9.847  0.888  ( 7.434, 12.261)
```

```
3000 MM     3  11.250  0.689  ( 8.837, 13.664)
3000 MM12   3  25.940  1.164  (23.526, 28.354)
3000 MM4    3   17.80   1.94  ( 15.39,  20.22)
3000 MN     3   9.622  0.845  ( 7.208, 12.036)
3500 MM     3  11.365  1.107  ( 8.951, 13.779)
3500 MM12   3  28.278  1.636  (25.864, 30.692)
3500 MM4    3   18.59   5.14  ( 16.17,  21.00)
3500 MN     3   9.959  1.079  ( 7.545, 12.373)

Pooled StDev = 2.02568
```

Tukey Pairwise Comparisons

```
Grouping Information Using the Tukey Method and 95% Confidence

Polymer     N    Mean  Grouping
3500 MM12   3  28.278  A
3000 MM12   3  25.940  A B
2500 MM12   3   22.20    B C
3500 MM4    3   18.59      C D
3000 MM4    3   17.80      C D
2500 MM4    3   16.12        D E
2500 MM     3  11.710          E F
3500 MM     3  11.365          E F
3000 MM     3  11.250          E F
3500 MN     3   9.959            F
2500 MN     3   9.847            F
3000 MN     3   9.622            F

Means that do not share a letter are significantly different.
```

Interval Plot of Sorption (mg) vs Polymer

One-way ANOVA: Normalized P Removal versus Interference

```
* NOTE * Cannot draw the interval plot for the Tukey procedure. Interval plots for
         comparisons are illegible with more than 45 intervals.


Method

Null hypothesis        All means are equal
Alternative hypothesis  At least one mean is different
Significance level      α = 0.05

Equal variances were assumed for the analysis.


Factor Information

Factor        Levels  Values
Interference      19  Cl 100, Cl 50, Cl 500, Control, HA 10, HA 2, HA 5, HCO3 10, HCO3
5,
                      HCO3 50, NO3 10, NO3 100, NO3 50, NOM 10, NOM 5, NOM 50, SO4
100, SO4
                      1000, SO4 50
```

```
Analysis of Variance

Source        DF    Adj SS    Adj MS   F-Value  P-Value
Interference  18   3.63853  0.202141   699.25    0.000
Error         38   0.01099  0.000289
Total         56   3.64951


Model Summary

         S    R-sq  R-sq(adj)  R-sq(pred)
0.0170024  99.70%     99.56%      99.32%


Means

Interference  N     Mean    StDev          95% CI
Cl 100        3   0.4505   0.0327  ( 0.4306,  0.4703)
Cl 50         3  0.54942  0.01150  (0.52955, 0.56930)
Cl 500        3  0.18654  0.01622  (0.16666, 0.20641)
Control       3   0.9243   0.0182  ( 0.9044,  0.9442)
HA 10         3  0.94200  0.00402  (0.92212, 0.96187)
HA 2          3  0.94226  0.00400  (0.92239, 0.96214)
HA 5          3  0.93911  0.00406  (0.91924, 0.95898)
HCO3 10       3  0.90242  0.00942  (0.88255, 0.92229)
HCO3 5        3  0.89075  0.00499  (0.87087, 0.91062)
HCO3 50       3  0.94455  0.00483  (0.92468, 0.96442)
NO3 10        3  0.90561  0.00821  (0.88574, 0.92548)
NO3 100       3  0.60352  0.00469  (0.58365, 0.62339)
NO3 50        3   0.7463   0.0249  ( 0.7264,  0.7661)
NOM 10        3  0.94226  0.00400  (0.92239, 0.96214)
NOM 5         3   0.9253   0.0180  ( 0.9055,  0.9452)
NOM 50        3  0.88180  0.01083  (0.86192, 0.90167)
SO4 100       3   0.5321   0.0420  ( 0.5122,  0.5519)
SO4 1000      3  0.14383  0.01244  (0.12395, 0.16370)
SO4 50        3   0.7469   0.0207  ( 0.7270,  0.7668)

Pooled StDev = 0.0170024
```

Tukey Pairwise Comparisons

```
Grouping Information Using the Tukey Method and 95% Confidence

Interference  N     Mean  Grouping
HCO3 50       3  0.94455  A
NOM 10        3  0.94226  A B
HA 2          3  0.94226  A B
HA 10         3  0.94200  A B
HA 5          3  0.93911  A B
NOM 5         3   0.9253  A B C
Control       3   0.9243  A B C
NO3 10        3  0.90561  A B C
HCO3 10       3  0.90242  A B C
HCO3 5        3  0.89075    B C
NOM 50        3  0.88180      C
SO4 50        3   0.7469        D
NO3 50        3   0.7463        D
NO3 100       3  0.60352          E
Cl 50         3  0.54942            F
SO4 100       3   0.5321            F
```

```
Cl 100       3   0.4505             G
Cl 500       3   0.18654                H
SO4 1000     3   0.14383                H
```

Means that do not share a letter are significantly different.

Interval Plot of Normalized P Removal vs Interference

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Retrieving project from file: 'E:\Research\MS\METAC\Statistical
Analysis.MPJ'
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