Reprinted from: Release of nonindigenous biological control agents: Permit application information supplement. January 1995.

Published by: USDA-APHIS-PPQ-BATS.

Oberea erythrocephala: Permit application information supplement

R. W. HANSEN

USDA/APHIS/PPQ, Bozeman, MT

(Article begins on the following page.)

Release of nonindigenous biological control agents: Permit application information supplement

USDA-APHIS-PPQ-BATS

Oberea erythrocephala (Coleoptera: Cerambycidae)

1. Proposed action

Field release of the root-boring beetle, *Oberea erythrocephala* (Schrank) (Coleoptera: Cerambycidae), for the biological control of the exotic weed, leafy spurge (*Euphorbia esula* L.), in the United States

2. Details of proposed action

2.1 Purpose of the release(s)

Releases of Oberea erythrocephala will be used to initiate or augment populations at field insectary sites (FIS) in various states. Once these FIS populations are successfully established and are deemed sufficiently large, O. erythrocephala will be collected and distributed to leafy spurge-infested areas throughout the state.

oviposition Due to feeding and activities, Oberea erythrocephala adults may cause limited defoliation or induce the upper part of a leafy spurge shoot to wither and die; this damage has little or no impact on the weed. However, O. erythrocephala larvae feed within spurge roots, and this damage may kill leafy spurge plants. 0. erythrocephala larvae may cause mortality directly, by destroying root tissues and disrupting water and nutrient transport and storage, or indirectly, by providing entry sites for soil-borne pathogenic fungi.

Thus, the primary role of *Oberea erythrocephala* in the leafy spurge biocontrol program is to cause mortality among leafy spurge plants, and to reduce the competitiveness of those plants that do survive. This, in turn, should lead to increased competitiveness and abundance of native and forage plants.

2.2 Need for release

Leafy spurge is a perennial herbaceous plant native to Europe and Asia. Since its accidental introduction beginning in the nineteenth century (Dunn 1985), leafy spurge has become a widespread and economically-important weed in the northern United States and in Canada. The major economic impact of leafy spurge is based on reduced cattle production on infested rangelands (Leistritz *et al.* 1992). The weed also has an adverse ecological impact by displacing native plants (Belcher and Wilson 1989) and, perhaps, by degrading wildlife habitats (Wallace *et al.* 1992).

A variety of tactics may be employed in managing weedy plants. Chemical (herbicides) and cultural (e.g. sheep grazing, cultivation) control techniques may be effective against leafy spurge in some situations, but their widespread utilization is limited by logistical, economic, or environmental constraints. Biological control, however, may offer an opportunity for large-scale, cost-effective leafy spurge management, especially in remote areas and in low-value grazing lands.

2.3 Specific location of the rearing facility and release site(s)

Oberea erythrocephala adults will be collected from an established field insectary site in Stillwater County, Montana. In 1995, these insects will be provided for field release in some or all of the following states: Colorado, Idaho, Iowa, Michigan, Minnesota, Montana, Nebraska, New Mexico, Nevada, North Dakota, Oregon, South Dakota, Utah, Washington, Wisconsin, and Wyoming. Releases may also be made in other states in 1995 and future years.

2.4 Number to be released

Generally, about 100 Oberea erythrocephala adults will be provided to each state in 1995. The exact number will be determined after assessing the O. erythrocephala populations at the Montana collection sites in summer 1995.

2.5 Timing of release

Oberea erythrocephala adults will be collected in mid- to late July 1995, depending on local weather conditions. Field release in the various states will be accomplished within two days of collection in Montana.

2.6 Methods used for release

Releases of *Oberea erythrocephala* adults will be made at open (uncaged) field insectary sites.

3. Biology of target organism

3.1 Common name, scientific name, and taxonomic classification

Common name: Leafy spurge Scientific name: *Euphorbia esula* L.

Phylum: Magnoliophyta Subtribe: Euphorbiinae Class: Magnoliopsida Genus: Euphorbia Subclass: Rosidae Subgenus: esula Order: Euphorbiales Section: esula Family: Euphorbiaceae Subsection: esulae Subfamily: Euphorbioideae Species: esula Tribe: Euphorbieae

The taxonomic status of leafy spurge, and the family Euphorbiaceae in general, has not yet been fully resolved. There is a great deal of morphological and biochemical variation among Eurasian and introduced North American populations of *Euphorbia esula* (Crompton *et al.* 1990, Evans *et al.* 1991, Holden and Mahlberg 1992, Manners and Davis 1984). This has led to some classifications of the weed as a complex of numerous species and interspecific hybrids (Ebke and McCarty 1983, Radcliffe-Smith 1985). However, there is ample evidence that leafy spurge can be treated as a single, though highly variable, species (Crompton *et al.* 1990, Evans *et al.* 1991, Harvey *et al.* 1988), a classification that will be followed in this document.

3.2 General life history

Leafy spurge is a long-lived herbaceous perennial plant whose aboveground stems die back each fall but whose welldeveloped root system and adventitious buds persist from year to year (Best *et al.* 1980). Buds initiate elongation during the fall but remain below the soil surface throughout the winter (Messersmith *et al.* 1985). Bud elongation resumes in early spring and the aboveground portions of the plant (shoots) become apparent in April or May, depending on location.

Vegetative shoots develop rapidly and reach their full height in June or July. Mature stem heights are variable, depending on soil and weather conditions, but generally range from 0.25 to 1.5 m. Stems may be branched or unbranched. Leafy spurge leaves are usually green or grayish-green in color, linear or lanceolate in shape, and from 2-8 cm long and 2-10 mm wide (Best *et al.* 1980). Leaves are arranged alternately along the stem.

Leafy spurge possesses specialized, unisexual flowers in a compound umbellate arrangement (Selleck *et al.* 1962). On some shoots, flower buds begin to develop several weeks after the shoots first appear, with peak flowering occurring in June and July (Best et al. 1980). Flowers may also appear later during the growing season, depending on weather conditions (Raju 1985). Because of the sticky pollen, asynchrony in maturity of adjacent male and female flowers, and the presence of nectaries, leafy spurge appears to be pollinated primarily by insects (Best et al. 1980, Messersmith et al. 1985). Seeds develop and mature from July through September, and are then explosively expelled up to 5 m from the parent plant; generally, from 30-150 seeds are produced by each flowering shoot (Selleck et al. 1962). Leafy spurge seeds may be dispersed over longer distances by flowing water (Selleck et al. 1962), birds (Blockstein et al. 1987) or grazing mammals (Lacey et al. 1992). Spurge seeds are also transported by humans in gravel, soil, hay, or farm equipment (Messersmith et al. 1985).

Leafy spurge seeds usually germinate in the early spring (Best et al 1980). Though viability decreases over time, some dormant seeds may germinate after 10 or more years in the soil (Bowes and Thomas 1978, Selleck et al. 1962). Typically, spurge seedlings develop a single shoot during the first growing season but do not flower (Messersmith et al. 1985). Seedling shoots are usually shorter and less robust than those originating from root buds, reaching heights of 20 cm or less. During the initial growing season, seedlings begin to develop an extensive root system that may extend up to about 1 m and that possesses many root buds (Messersmith et al. 1985). In general, seedlings serve to initiate new leafy spurge patches; exposed mineral soil associated with disturbance (e.g. cultivation, trails, roads, overgrazing) seem best suited for leafy spurge seedling and, hence, patch establishment (Belcher and Wilson 1989). Only a small percentage of newly-germinated seedlings are able to survive in established patches (Best et al. 1980).

Established spurge patches possess a network of lateral and vertical roots that serve perenniating, reproductive, and nutrient and water storage functions (Stroh *et al.* 1990). Most of the root biomass is located in the upper 15 cm of the soil (Selleck *et al.* 1962), but some vertical roots may reach depths of 9 m or more (Best *et al.* 1980). Two types of adventitious buds are formed on the root system (Messersmith *et al.* 1985): **crown** buds are formed on the root system root crown, at the base of a current-year shoot, while **root**

buds may form almost anywhere along the lateral and vertical roots. Generally, one or more crown buds will elongate at the same location each year, while the number and location of elongating root buds varies greatly from year to year and from plant to plant (Messersmith *et al.* 1985). Root buds may be formed "spontaneously" or in response to root injury (Messersmith *et al.* 1985). Most root buds occur near the soil surface, but some may be found more than 3 m deep along vertical roots (Stroh *et al.* 1990).

Only a subset of crown and root buds formed in a given year actually produce aboveground shoots, through a system of hormonal control and, perhaps, competition for water and nutrients (Raju 1985). Removal of shoots (e.g. mowing or grazing) usually causes activation of some dormant crown and root buds, and the production of new shoots; new shoot density often exceeds that observed before the treatment (Messersmith *et al.* 1985). Leafy spurge root fragments as small as 2 cm long will produce new shoots and root systems and, hence, new plants, provided they remain buried in soil (Messersmith *et al.* 1985).

Established leafy spurge plants expand vegetatively, through elongation of the horizontal roots and the formation of adventitious buds (Best et al. 1980). As some of these buds produce shoots in subsequent years, new root crowns are formed. The root connections among the new root crowns and the parent root system eventually disintegrate, resulting in independent, "daughter" plants. Through lateral expansion of the root systems, leafy spurge patches expand up to about 1 m in radius each year (Selleck et al. 1962, Stroh et al. 1990). At a given location, the rate of radial increase in patch size remains fairly constant from year to year (Selleck et al. 1962).

3.3 Pest status

Leafy spurge occurs in at least 30 US states and nine Canadian provinces (Dunn 1979), but is a significant economic problem primarily in Minnesota, Montana, Nebraska, North Dakota, South Dakota, Wyoming, Alberta, Manitoba, and Saskatchewan. Currently, about 660,000 ha are infested in Montana, North Dakota, South Dakota, and Wyoming, and the affected area doubles in size about every 10 years (Leitch *et al.* 1994).

Leafy spurge, like other members of the Euphorbiaceae, contains a milky latex throughout all parts of the plant. This latex is composed of a complex of chemicals, including a variety of terpenoid compounds (Mahlberg 1989). Some latex chemicals from leafy spurge have skin irritant and tumor-inducing properties in mammals (Seip and Hecker 1982, Upadhyay *et al.* 1978). Paradoxically, other compounds from *E. esula* latex may have antileukemic properties; plants in the family Euphorbiaceae have long been used to treat cancers and tumors in traditional medicine (Kupchan *et al.* 1975).

Due to its latex chemistry, leafy spurge can induce a variety of digestive maladies or, in sufficient quantities, may cause death when consumed by cattle (Kronberg *et al.* 1993). However, spurge is rarely eaten by cattle, who instead avoid spurge-infested pasture despite the presence of palatable grasses (Hein and Miller 1992, Lym and Kirby 1987). Significant (>50%) reductions in forage utilization by cattle result when leafy spurge achieves 10% or more of plant cover (Hein and Miller 1992). Interestingly, domestic sheep and goats are able to consume leafy spurge with no visible detrimental effects (Landgraf *et al.* 1984).

Leafy spurge is an aggressive competitor that, because of its expansive root system and dense shoot growth, is able to outcompete rangeland grasses and forbs for available water, nutrients, and light. In addition, leafy spurge appears to exert an allelopathic effect on other plants, possibly through chemicals leached from decomposing leaf, stem, and root tissues (Steenhagen and Zimdahl 1979). Generally, the abundance of grasses and other forbs is significantly reduced in established leafy spurge patches, and some species may disappear altogether (Belcher and Wilson 1989, Nowierski and Harvey 1989).

Leafy spurge infestations significantly reduce the abundance of native prairie plants (Belcher and Wilson 1989). This reduction in native plant diversity may have a negative impact on wildlife populations (Wallace *et al.* 1992).

The primary economic impacts of leafy spurge are based on reductions in available forage and, hence, reduced cattle production on infested rangeland. In Montana, North Dakota, South Dakota, and Wyoming, Leitch et al. (1994) estimate that direct and secondary losses due to lost cattle production approach \$120 million а year. An \$10 non-agricultural additional million in (e.q. recreational and watershed) losses (Wallace et al. 1992) bring the total losses due to leafy spurge to about \$130 million annually in the four-state area (Leitch et al. 1994).

4. Biology of organism to be released

4.1 Common name, scientific name, and taxonomic classification

Common name: none Scientific name: Oberea erythrocephala (Schrank)

Phylum: Arthropoda Superfamily: Chrysomeloidae Class: Insecta Family: Cerambycidae Subclass: Pterygota Subfamily: Lamiinae Division: Endopterygota Tribe: Saperdini Order: Coleoptera Genus: Oberea Suborder: Polyphaga Species:

erythrocephala

4.2 Taxonomic specialist(s) who identified organism

Oberea erythrocephala populations at the collection site (see 2.3, above) originated with insects collected from Europe. These insects were identified by R.E. White, USDA-ARS, Systematic Entomology Laboratory (SEL). Subsequent confirmation of *O. erythrocephala* adults from these FIS has been made by Dr. White.

4.3 Location of voucher specimens

Type material for Oberea erythrocephala is kept at the International Institute of Biological Control in Delémont, Switzerland and the US National Museum of Natural History (USNM) in Washington, DC. Voucher specimens from later redistribution collections are kept at the SEL and USDA-APHIS Bozeman Biological Control Facility.

4.4 Natural geographic range of organism

Oberea erythrocephala occurs throughout southern Europe, through Russia into southern Siberia, and in central Asia (Schroeder 1979).

4.5 Location where organism was originally collected

Insects initially released in the United States were collected in northern Italy and in Switzerland.

4.6 General life history

Oberea erythrocephala overwinters as a dormant larva within a leafy spurge root. Overwintered larvae resume development in the following spring; in late spring, larvae construct a pupal cell in the upper part of the root crown and molt to the pupal stage. Adult beetles chew through the remaining root crown tissue and exit the pupal cell, emerging from the soil in early to mid-summer. *O. erythrocephala* adults feed on leafy spurge leaves, flowers, and stem tissue. They are slender beetles about 10-12 mm long, with characteristic reddish-orange head and thorax and long, slender antennae. *Oberea erythrocephala* beetles are active fliers, and on warm days may be found moving about on or above the leafy spurge canopy. Adults are relatively long-lived, capable of surviving up to a month or longer under field conditions (Schroeder 1979).

Mating occurs on the upper part of leafy spurge shoots. After mating, an O. erythrocephala female girdles a leafy spurge stem, chewing completely around the upper part of the stem one or more times (Schroeder 1979). An egg niche is chewed into the spurge stem above the girdles, into which a single eqg is laid; after oviposition, this niche is usually sealed by dried spurge latex. Generally, only a single eqq is laid on each attacked shoot. O. erythrocephala eggs hatch in about two weeks, and young larvae tunnel downward through the stem and enter the root crown area. Most larval feeding occurs within the root crown and largest lateral roots, beginning in mid- to late summer and continuing until the onset of dormancy in the fall. Most of the internal root crown tissue is consumed by the time larval development is completed. Though only one Ο. erythrocephala larva typically survives in each attacked shoot, several larvae may complete development in leafy spurge plants with large root systems (Schroeder 1979).

In Europe, Oberea erythrocephala completes its life cycle in one year (Schroeder 1979), but two years are apparently required to complete development in most of the US and Canada . In North America, O. erythrocephala larvae continue feeding through a second summer, enter dormancy over a second winter, and complete development during the subsequent spring.

4.7 Host range in the field

Oberea erythrocephala appears to feed only on leafy spurge (Euphorbia esula), cypress spurge (E. cyparissias) and a few other closely-related Euphorbia spp. in its native Europe (Schroeder 1979). To date, introduced US populations of O. erythrocephala have been reported only from E. esula.

4.8 Host range in laboratory/greenhouse tests

Controlled greenhouse studies showed at least limited feeding and some oviposition by Oberea erythrocephala adults on a number of European Euphorbia spp. in several subgenera (Schroeder 1979, 1980). However, only a few European Euphorbia spp. in the subgenus Esula supported larval development and thus could be considered likely hosts (Schroeder 1979, 1980). No North American spurges in the subgenus Esula were included in these tests. Poinsettia (E. pulcherrima), an important horticultural plant, was not utilized by O. erythrocephala.

Thus, the host plant range of *Oberea erythrocephala* appears restricted below the subgeneric level, and may only include leafy spurge and other Eurasian *Euphorbia* species in the subgenus *Esula*.

4.9 Specific references on the organism

- Schroeder, D. 1979. Investigations on Oberea erythrocephala (Schrank.) (Col.: Cerambycidae), а possible biocontrol aqent of leafv spurge, Euphorbia sp. (Euphorbiaceae) in Canada. Intl. Inst. of Biol. Contr., Delémont, Switzerland. Final report. 25 p.
- Schroeder, D. 1980. Investigations on Oberea erythrocephala (Schrank) (Col.: Cerambycidae), a possible biocontrol agent of leafy spurge, Euphorbia spp. (Euphorbiaceae), in Canada. Z. angew. Entomol. 90: 237-254.

4.10 List of known parasitoids or predators of organism

Several specialized hymenopteran parasitoids (Braconidae and Ichneumonidae) have been reported among European populations of Oberea erythrocephala (Schroeder 1979). However, these parasitoids do not occur in North America, and no native or introduced parasitoids have been reported among O. erythrocephala populations in the US.

5. Distribution of organism in the US

5.1 Current North American distribution

Oberea erythrocephala was approved for US release in November 1979. Through 1994, O. erythrocephala has been released in 11 US states: Colorado, Iowa, Minnesota, Montana, Nebraska, North Dakota, Oregon, South Dakota, Washington, Wisconsin, and Wyoming. Established populations are present in Colorado, Montana, and North Dakota. The status of populations in other states remains uncertain, but initial releases in most were made in 1993 and 1994. The insect also has a limited distribution in central and western Canada.

5.2 Expected North American range

There are no obvious climatic or ecological barriers to survival and establishment of *Oberea erythrocephala* in most or all of the spurge-infested areas of the US and Canada. Anecdotal evidence suggests that this agent may be poorly adapted to far-northern areas of the US, and recent distribution efforts have concentrated in the southern spurge-infested states. Of course, the ultimate North American range of this insect will reflect the extent of human redistribution activities.

6. Expected environmental impact of proposed release(s)

6.1 Human impacts

Oberea erythrocephala releases should have no impact on humans or on private property, exclusive of impacts on leafy spurge infestations.

6.2 Direct impacts

Under optimal site conditions, Oberea erythrocephala populations will, directly or indirectly, kill leafy spurge plants over large areas. Concurrently, the relative abundance of nontarget grasses and forbs will be expected to increase.

The host range of Oberea erythrocephala is limited to plant species in the subgenus Esula of the genus Euphorbia, including the target weed (leafy spurge) and cypress spurge (E. cyparissias), an introduced weed in eastern North America. The two federally-protected native spurges (Euphorbia garberi and E. deltiodes) are in the subgenus Chamaesyce (Pemberton 1985) and are not potential host plants for Oberea erythrocephala.

The potential host status of 21 North American species (occurring north of Mexico) in the subgenus *Esula* (Pemberton 1985) has not been evaluated. This group includes *E. purpurea* and *E. telephiodes*, two rare species being considered for protection (Pemberton 1985). Nine of the 21 native species in the subgenus *Esula* are annuals (Pemberton 1985) that could possibly be utilized "temporarily" by Oberea erythrocephala, but would not permit completion of the life cycle and, hence, population establishment; beetle larvae require plant roots yearround. The 12 perennial species in the subgenus could be considered possible O. erythrocephala hosts, though most occur in the southern US and are not sympatric with leafy spurge populations (Pemberton 1985).

6.3 Indirect effects

No native or exotic insects, birds, reptiles, or mammals are known to depend largely or exclusively on leafy spurge. Thus, *Oberea erythrocephala* releases should have no adverse indirect impacts.

6.4 Methods to prevent undesired environmental effects

No undesired environmental effects are anticipated.

7. Literature cited

- Belcher, J. W. & S.D. Wilson. 1989. Leafy spurge and the species composition of a mixed-grass prairie. J. Range Manage. 42: 172-175.
- Best, K. F., G.G. Bowes, A.G. Thomas & M.G. Maw. 1980. The biology of Canadian weeds. 39. Euphorbia esula L. Can. J. Plant Sci. 60: 651-663.
- Blockstein, D. E., B.D. Maxwell & P.K. Fay. 1987. Dispersal of leafy spurge seeds (Euphorbia esula) by mourning doves (Zenaida macroura). Weed Sci. 35: 160-162.
- Bowes, G. G. & A.G. Thomas. 1978. Longevity of leafy spurge seeds in the soil following various control programs. J. Range Manage. 31: 137-140.
- Crompton, C. W., A.E. Stahevitch & W.A. Wojtas. 1990. Morphometric studies of the *Euphorbia esula* group (Euphorbiaceae) in North America. Can. J. Bot. 68: 1978-1988.
- Dunn, P. H. 1979. The distribution of leafy spurge (Euphorbia esula) and other weedy Euphorbia spp. in the United States. Weed Sci. 27: 509-516.
- Dunn, P. H. 1985. Origins of leafy spurge in North America, pp. 7-13. In A.K. Watson [ed.], Leafy spurge. Weed Sci. Soc. Am., Champaign, IL. 104 p.

- Ebke, D. H. & M.K. McCarty. 1983. A nursery study of leafy
 spurge (Euphorbia spp.) complex from North America. Weed
 Sci. 31: 866-873.
- Evans, J. O., J.M. Torrell, R.V. Valcarce & G.G. Smith. 1991. Analytical pyrolysis-pattern recognition for the characterisation of leafy spurge (*Euphorbia esula* L.) biotypes. Ann. Appl. Biol. 119: 47-58.
- Harvey, S. J., R.M. Nowierski, P.G. Mahlberg & J.M. Story. 1988. Taxonomic evaluation of leaf and latex variability of leafy spurge (Eurphorbia spp.) for Montana and European accessions. Weed Sci. 36: 726-733.
- Hein, D. G. & S.D. Miller. 1992. Influence of leafy spurge on forage utilization by cattle. J. Range Manage. 45: 405-407.
- Holden, A. N. G. & P.G. Mahlberg. 1992. Application of chemotaxonomy of leafy spurges (*Euphorbia* spp.) in biological control. Can. J. Bot. 70: 1529-1536.
- Kronberg, S. L., R.B. Muntifering, E.L. Ayers & C.B. Marlow. 1993. Cattle avoidance of leafy spurge: a case of conditioned aversion. J. Range Manage. 46: 364-366.
- Kupchan, S. M., I. Uchida, A.R. Branfman, R.G. Dailey Jr & B.Y. Fei. 1975. Antileukemic principles isolated from Euphorbiaceae plants. Science 191: 571-572.
- Lacey, J. R., R. Wallander & K. Olson-Rutz. 1992. Recovery, germinability, and viability of leafy spurge (Euphorbia esula) seeds ingested by sheep and goats. Weed Tech. 6: 599-602.
- Landgraf, B. K., P.K. Fay & K.M. Havstad. 1984. Utilization of leafy spurge (Euphorbia esula) by sheep. Weed Sci. 32: 348-352.
- Leistritz, F. L., F. Thompson & J.A. Leitch. 1992. Economic impact of leafy spurge (Euphorbia esula) in North Dakota. Weed Sci. 40: 275-280.
- Leitch, J. A., F.L. Leistritz & D.A. Bangsund. 1994. Economic effect of leafy spurge in the upper Great Plains: methods, models, and results. North Dakota St. Univ. Agric. Econ. Rep. 316. 8 p.

- Lym, R. G. & D.R. Kirby. 1987. Cattle foraging behavior in leafy spurge (Euphorbia esula L.) infested rangeland. Weed Tech. 1: 314-318.
- Mahlberg, P. G. 1989. Chemotaxonomic affinities of Eurasian leafy spurges (*Euphorbia* spp.) in relation to a biological control program, pp. 145-151. In E.S. Delfosse [ed.], Proc. VII Intl Symp. Biol. Contr. Weeds. Rome, Italy, March 6-11, 1988. Inst. Sper. Patol. Veg., Rome, Italy.
- Manners, G. D. & D.G. Davis. 1984. Epicuticular wax constituents of North American and European Euphorbia esula biotypes. Phytochemistry 23: 1059-1062.
- Maw, E. 1981. Biology of some Aphthona spp. (Col.: Chrysomelidae) feeding on Euphorbia spp. (Euphorbiaceae), with special reference to leafy spurge (Euphorbia sp. near esula). MS Thesis. Univ. of Alberta, Edmonton, AB. 258 p.
- Messersmith, C. G., R.G. Lym & D.S. Galitz. 1985. Biology of leafy spurge , pp. 42-56. In A.K. Watson [ed.], Leafy spurge. Weed Sci. Soc. Am., Champaign, IL. 104 p.
- Nowierski, R. M. & S.J. Harvey. 1987. Vegetation composition at four rangeland sites infested by leafy spurge. Montana State Univ. MSU Biology Rept. 40. 21 p.
- Pemberton, R.W. 1985. Native plant considerations in the biological control of leafy spurge, pp. 365-390. In E.S. Delfosse, ed. Proc. VI Intl. Symp. Biol. Contr. Weeds. Agriculture Canada, Ottawa. 885 p.
- Radcliffe-Smith, A. 1985. Taxonomy of North American leafy
 spurge , pp. 14-25. In A.K. Watson [ed.], Leafy spurge.
 Weed Sci. Soc. Am., Champaign, IL. 104 p.
- Raju, M. V. S. 1985. Morphology and anatomy of leafy spurge , pp. 26-41. In A.K. Watson [ed.], Leafy spurge. Weed Sci. Soc. Am., Champaign, IL. 104 p.
- Rees, N. E. & N.R. Spencer. 1993. Biological control of leafy
 spurge, pp. 182-192. In L.F. James, J.O. Evans, M.H. Ralphs
 & R.D. Child [eds.], Noxious range weeds. Westview Press,
 Boulder, CO.
- Schroeder, D. 1979. Investigations on Oberea erythrocephala
 (Schrank.) (Col.: Cerambycidae), a possible biocontrol
 agent of leafy spurge, Euphorbia sp. (Euphorbiaceae) in
 Canada. Intl. Inst. of Biol. Contr., Delémont,
 Switzerland. Final report. 25 p.

- Schroeder, D. 1980. Investigations on Oberea erythrocephala
 (Schrank) (Col.: Cerambycidae), a possible biocontrol agent
 of leafy spurge, Euphorbia spp. (Euphorbiaceae), in Canada.
 Z. angew. Entomol. 90: 237-254.
- Seip, E. H. & E. Hecker. 1982. Skin irritant ingenol esters from Euphorbia esula. J. Medicinal Plant Res. 46: 215-218.
- Selleck, G. W., R.T. Coupland & C. Frankton. 1962. Leafy spurge in Saskatchewan. Ecol. Monogr. 32: 1-29.
- Steenhagen, D. A. & R.L. Zimdahl. 1979. Allelopathy of leafy spurge (Euphorbia esula). Weed Sci. 27: 1-3.
- Stroh, R. K., D. Bangsund & J.A. Leitch. 1990. Leafy spurge
 patch expansion. ND State Univ. Dept. of Agric. Econ.
 Staff Paper Series No. AE90001. 17 p.
- Upadhyay, R. R., F. Bakhtavar, M. Ghaisarzadeh & J. Tilabi. 1978. Cocarcinogenic and irritant factors of Euphorbia esula L. latex. Tumori 64: 99-102.
- Wallace, N. M., J.A. Leitch & F.L. Leistritz. 1992. Economic impact of leafy spurge on North Dakota wildland. North Dakota St. Univ. Agric. Exp. Sta. Agric. Econ. Rep. No. 281. 23 p.

R.W. Hansen USDA-APHIS-PPQ Forestry Sciences Lab Montana State Univ. Bozeman, MT 59717-0278

19 January 1995