# DEVELOPING METHODOLOGY TO PREDICT AND INCREASE GRAIN PROTEIN

## CONTENT IN SPRING WHEAT

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# Developing Methodology to Predict and Increase Grain Protein Content in Spring Wheat

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## **MASTER OF SCIENCE**

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#### ABSTRACT

A challenge for hard red spring wheat (HRSW) (*Triticum aestivum* L. emend Thell.) producers is to obtain both high yields and market-required grain protein content (GPC). The ability to accurately predict HRSW yield with the Decision Support System for Agrotechnology Transfer (DSSAT) crop model early in the growing season may help producers determine probable GPC and lead to management decisions on whether to apply supplemental nitrogen (N) to enhance protein. A management decision HRSW producers may consider in high yielding environments is a late-season foliar N application to increase GPC. A second objective of this research was to test methods to improve the efficiency of a foliar N application. Improving the efficiency of a late-season foliar N application coupled with the ability to predict high yielding environments using DSSAT, can provide producers with effective management tools to determine the optimum situation in which supplementing GPC will have the most economic success.

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### PREFACE

This thesis was written as two manuscripts that will be submitted for publication in the appropriate scientific journals. The 'Introduction' provides a general review of this study and how both chapters are related to the main issue, developing methodology to predict and increase grain protein content in spring wheat. Following the Introduction, the thesis is divided into two manuscripts which contain Introduction, Materials and Methods, Results and Discussion, Conclusion, and References Cited sections that are specific to the chapter. The references for the 'General Introduction' can be found in the 'General References Cited' section.

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#### **INTRODUCTION**

Hard red spring wheat (HRSW) (*Triticum aestivum* L. emend Thell.) producers not only depend on quantity of grain in terms of yield, but also need to achieve market-standard grain protein content (GPC) in order to achieve maximum economic returns. Previous research has indicated a negative relationship between yield and GPC within a given cultivar and at a given level of fertility, which poses a challenge for HRSW producers as quality discounts at point of sale result when protein levels do not reach the market-standard (Alkier, 1972). However, a premium can usually be obtained when GPC levels exceed the market-standard (Jones and Olson-Rutz, 2012). Environmental and agronomic challenges influence maximum yield, which may encourage some HRSW producers to make in-season management decisions in order to regain the desired yield if the factors are negative, or may prompt some to conduct an in-season nitrogen (N) application if yield is likely to be high and the probability of a price premium for high GPC seems likely.

The ability to accurately predict yield may assist producers in determining probable GPC; however, environmental factors that influence crop growth and yield are highly unpredictable between years. A crop simulation model (CSM), such as the Decision Support System for Agrotechnology Transfer (DSSAT) is a prediction tool that uses environmental and agronomic interactions to determine soil impacts, crop growth, and yield components (Boote et al., 2015). The DSSAT-CSM has been used extensively to predict yield with preseason weather and agronomic inputs such as planting, fertilizer, and irrigation options (Bannayan et. al, 2003). Implementing DSSAT-CSM with in-season weather information has not been as widely studied for HRSW production, but its use may prove be an effective predictive tool to help producers make improved management decisions.

A management decision HRSW producers may consider in high yielding environments is an application of foliar N, such as urea (46-0-0) solution (US) or urea ammonium nitrate (UAN, 28-0-0), to increase GPC. Increasing GPC has been successfully practiced through this additional N application between ZGS 45 and ZGS 73 with the most effective at ZGS 69 (Zadoks et al., 1974); however, the efficiency of N absorption into the plant has been limited (Finney et al., 1957; Jones and Olson-Rutz, 2012). Therefore, developing methods to improve the efficiency of this foliar N application may prove to be economically feasible.

Developing techniques to improve N use efficiency from a late-season foliar N application coupled with improved yield prediction through the use of DSSAT-CSM may result in improved protein management for producers. These tools can assist producers in determining when supplemental N would have the greatest economic benefit.

#### References

- Alkier, A.C., G.J. Racz, and R.J. Soper. 1972. Effects of foliar- and soil-applied nitrogen and soil nitrate-nitrogen level on the protein content of Neepawa wheat. Can. J. Soil Sci. 52:301-309.
- Bannayan, M., N.J. Crout, and G. Hoogenboom. 2003. Application of the CERES-wheat model for within-season prediction of winter wheat yield in the United Kingdom. Agron. J. 95:114-125.
- Batchelor, W.D., K.J. Boote, A.J. Gijsman, L.A. Hunt, G. Hoogenboom, J.W. Jones, C.H. Porter,
  J.T. Ritchie, U. Singh, and P.W. Wilkens. 2015. Decision Support System for
  Agrotechnology Transfer (DSSAT) Version 4.6 (www.DSSAT.net). DSSAT Foundation,
  Prosser, Washington.

- Finney, K.F., H.C. Freyer, W. Meyer, and F.W. Smith. 1957. Effect of foliar spraying of Pawnee wheat with urea solution on yield, protein content, and protein quality. Agron. J. 49:341-347.
- Jones, C., and K. Olson-Rutz. 2012. Practices to increase grain protein. Misc. Publ. Montana State Univ., Bozeman, MT.
- Zadoks, J.C., T.T. Chang, and C.F. Konzak. 1974. A decimal code for the growth stages of cereals. Weed Res. 14:415-421.

# ARTICLE 1: RELIABILITY OF PREDICTING SPRING WHEAT YIELD WITH DSSAT USING EARLY SEASON WEATHER DATA

#### Abstract

A crop simulation model (CSM) can be a predictive tool to help hard red spring wheat (HRSW) (*Triticum aestivum* L. emend Thell.) producers make in-season crop management decisions, such as a late-season foliar N application to increase grain protein content (GPC) in high yielding environments. The objective of this research was to evaluate the Decision Support System for Agrotechnology Transfer (DSSAT) CSM in predicting HRSW yield at various points in the growing season to determine how early the model could be implemented to accurately predict final grain yield. Historic weather data was used in three approaches (distribution, historical average, analogue) to forecast weather for the remainder of the season from Zadoks growth stage (ZGS) 14, 45, and 61 for five locations throughout North Dakota. Across environments the strength of relationship between observed and simulated anthesis date and yield was significant at  $p \le 0.01$ . The optimum approach to simulate grain yield with forecasted weather was through a distribution style from ZGS 45 or 61.

#### Introduction

Weather events are often unpredictable and vary considerably between and within years. These events have a tremendous impact on the success of agricultural producers due to the environmental impact on yield potential. In order to obtain the desired grain quality, HRSW producers often make in-season management decisions based on the weather. However, generalized weather patterns are frequently used, which can make these decisions risky due to lack of specific accuracy (Hansen et al., 2000). Crop models, such as DSSAT, are software applications that use environmental and agronomic interactions to predict soil processes, vegetative growth, and yield outcomes (Batchelor et al., 2015). The DSSAT has the potential to predict these outcomes with defined weather and agronomic inputs, which allows users to make improved crop management recommendations (Hansen et al., 2000). In this paper the abbreviation 'DSSAT' will refer to the software application of the CSM used in this study.

In HRSW production, GPC determined at the point of sale must meet the market-standard level to avoid discounts for low GPC or receive a premium if GPC is greater. For a given cultivar, a negative relationship exists between yield and GPC at a given level of soil fertility. A management technique to increase GPC is through a foliar application of N fertilizer between the ZGS 45 and 73 growth stages (Finney et al., 1957). However, this application is not economical unless crop yields coupled with the premium increase or discount results in revenue that is greater than the cost of the additional N application.

A decision to make the additional application of N carries risk due to the unknown yield of the current crop, since future weather events throughout the season are unpredictable. Yield and protein prediction can be done through multiple strategies, but do not take into consideration the impact of future weather. The DSSAT has predictive capabilities by using updated weather information during the growing season supplemented with estimated future weather. Therefore, the use of DSSAT may be beneficial to agricultural producers by more accurately simulating crop growth, allowing for proactive in-season management practices to achieve maximum economic return.

#### **Crop Simulation Models and DSSAT Overview**

Crop simulation models estimate crop growth and yield throughout the growing season by simulating the multiple ecological and agronomic interactions that impact growth. These include daily weather data, soil characteristics, and crop management practices (Batchelor et al., 2003). The DSSAT is a computer program developed in 1985 and first released in 1986 for maize (*Zea mays* L.), wheat, soybean (*Glycine max* L.), and peanut (*Arachis hypogea* L.) crops (Batchelor et al., 2003). Further upgraded versions included an expansion of 42 crops and improved model calibration. The DSSAT simulates crop development, ontogeny, and phenology through heat unit accumulation and using carbon, N, and water balance principles within a defined area (Clarke et al., 2010).

The DSSAT incorporates models for broadleaf and cereal crops within a single program. The CROPGRO model computes the growth of various broadleaf crops and is further described by Boote et al. (1998). The CERES-maize, wheat, and barley (*Hordeum vulgare* L.) models compute the growth of cereal crops. Crop developments in the CERES models are regulated by growing degree days (GDD), which are computed internally from daily maximum and minimum temperatures uploaded into the program. Clarke et al. (2010) details the function of CERESwheat model with a base temperature for GDD calculations of 0°C.

The soil program within DSSAT integrates water, temperature, carbon, and N within the soil profile. A soil series is represented by a one-dimensional profile that is vertically layered and horizontally homogenous (Batchelor et al., 2003). Decomposition of organic matter and plant availability of N is determined by two primary soil organic modules, CENTURY (Parton et al., 1988; Parton et al., 1994) and PAPRAN (Seligman and Van Keulen, 1981). Soil water content is determined by soil layer characteristics and changes as water is supplied by precipitation or

irrigation. The lower limit and saturated water content within each layer is computed and subsequent water exceeding the drained upper limit percolates to the soil layer below and continues downward for each layer, depending on the initial amount of water added. Surface water runoff is computed using a modified runoff curve number from the USDA-Soil and Conservation Service (Williams, 1990). Soil evaporation is computed by an approach used by Priestly and Taylor (1972). Nutrient availability and movement use similar functions as soil water.

The DSSAT accounts for genotypic and phenotypic variations within a specific crop to environmental conditions. This allows for more accurate representations of cultivar-environment interactions (Batchelor et al., 2003). To represent the genetic makeup of a crop and cultivar, data files within the program have set parameters called genetic coefficients. The genetic coefficients are determined for each cultivar using crop, weather, and soil data (Batchelor, et al., 2003). Crop species may require different coefficients. Wheat genetic coefficients include vernalization, photoperiod response, grain filling duration, interval between leaf tip appearances, kernel number per unit canopy weight at anthesis, kernel size, and non-stressed mature tiller weight.

A strength of DSSAT is that expensive and time-consuming agronomic experiments can be simulated in a relatively short amount of time; however, limitations exist within models. Model performance is limited by the accuracy of the input variables, which can be affected by spatial and temporal variability, costs of measuring data, and technical knowledge. Model performance is only as good as the data quality input (Antle et al., 2016). Misuse as well as misrepresentation or misunderstanding of the tool can affect the usefulness of the model; therefore, limitations of the model output must be understood.

#### **Crop Modeling Techniques**

Crop model development began from a need for improving decisions for multiple environments from a single system that combined soil, climate, crops, and management information (Batchelor et al., 2003). The crop modeling software has been used extensively by researchers as a supplement to experiments and more recently has been used to support management decisions for producers (Boote et al., 1996). The DSSAT has specifically been used for research on the impact of fertilization, irrigation, pest management, climate variability, and site-specific farming on crop production. A new concept in the use of DSSAT by consultants is to forecast yield to help agricultural producers make management decisions (Bannayan et al., 2003).

Spring wheat producers might directly benefit from the use of a CSM if yield can be more accurately predicted for their region. As the growing season progresses, yield prediction is increasingly more accurate, whereas early season weather provides limited guidance for later season conditions and yield prediction. Bannayan et al. (2003) used the CERES-wheat model to predict wheat yield during the growing season with updated weather conditions as the growing season progressed. Results indicated forecasting grain yield improved significantly from early vegetative growth stages through the appearance of the flag leaf, with no significant improvement after that point.

Crop models have been used to forecast yield on both a small and large-scale through different approaches (Bannayan et al., 2003). In these approaches, the weather throughout the entire growing season is unknown, so past weather data are often used as an indication of probable future weather patterns. One technique in forecasting is to use past weather represented by daily averages calculated over multiple years. This approach uses a generalized climate since

it does not take into account the variability that can exist between years and will give a single predicted outcome of crop production (Barnett, 2004). A range of crop production expectations for a given year can be achieved when multiple past weather years are used for the same forecast. This range will give a distribution of outcomes with a maximum, median, and minimum expectation. The ability to accurately predict yield early in the growing season would be beneficial to producers and researchers as this would allow them the opportunity to make inseason management decisions more successfully (Batchelor et al., 2001).

#### **Predicting Yield and Grain Protein**

Spring wheat producers can effectively estimate grain yield prior to harvest through multiple techniques including kernel counts (Wiersma and Ransom, 2005), empirical based models (Balaghi et al., 2008), or proximal canopy sensing (Arnall et al., 2006). However, these techniques have limitations. For example, kernel counts are not taken until late in the growing season when it is too late to improve yield with additional inputs. Empirical based models and canopy sensing indicate the current condition of the crop, but cannot take into consideration the impact of future weather. Crop simulation models may be a tool that can assess the current condition of the crop and use estimated future weather to give an accurate assessment of yield. In turn, producers can use this information to make late season management decisions.

A foliar application of N fertilizer during the fruiting period in HRSW may be a management option to increase GPC (Baltensperger et al., 2008). However, this application may not be economical if the yield is not high enough to realize an economic benefit from the additional application. The premium received for protein levels above the market threshold of 140 g kg<sup>-1</sup> for HRSW and 120 g kg<sup>-1</sup> for hard red winter wheat is not fully realized until after

harvest due to the uncertainty of GPC of the current crop. The level of GPC is only determined after harvest, as there are currently no techniques to predict protein levels in-season.

Grain yield and GPC have an inverse relationship for a given cultivar and at a certain level of soil fertility. Therefore, understanding the impact of weather events on grain yield may indicate whether GPC levels are likely to be high or low. Weather variables such as temperature, solar radiation, soil moisture, and nutrient availability directly impact grain yield. Wheat response to N applications can depend on water and nutrient availability with low response to N in environments with low soil moisture from lack of rainfall or high temperatures. The DSSAT has the ability to depict the effect of weather events on the soil and crop to provide an estimation of end season yield. If high yields are estimated by the crop model, with insufficient N, then the simulations may provide support for a late season foliar N application to increase GPC.

#### Objective

The objective of this research was to determine if DSSAT could be used to predict HRSW yield in order to support a decision on the use of a late season foliar N application. If so, a further objective was to determine the best strategy to predict yield with estimated future weather through multiple modeling approaches at various HRSW growth stages.

#### **Materials and Methods**

#### **Crop Model Experiment Setup**

In order for simulations to be conducted within the DSSAT, hypothetical experiments had to be created with the experiment builder called the 'Crop Management' program. The program combines management inputs to enable of the simulation of soil and crop behavior. Required inputs for the experiment builder include cultivar, soil series, weather data, planting characteristics, fertility and water conditions, simulation execution and harvest dates, and model

functions (water and nutrients on/off). In this study, the DSSAT version 4.6 was used to simulate separate HRSW experiments using inputs for the years 2005-2016 at Carrington, Hettinger, Langdon, Minot, and Williston, ND.

All input variables were the same for each experiment except planting date, soil type, and weather data. Planting dates for each location and all years are presented in Table 1.1. The required planting information included seeding rate, seeding depth, and row width, which were set at 290 seeds m<sup>-2</sup>, 4 cm, and 18 cm, respectively. Other required information includes initial field conditions such as previous crop with N credit. The previous crop was not known; therefore, wheat was set as the previous crop since it provided no N credit. Simulation start date and nutrient and water options were also set the same across all locations. The simulation start date was set to the day of planting and water set to run using recorded precipitation. Fertilization information was not available for all years so the assumption was made that experiments were fertilized to the recommended level and not limited throughout the year so nutrient options were turned off.

Simulations were executed for each year and location for a total of 60 simulations (5 locations, 12 years). Anthesis date (days after planting, DAP) and grain yield were extracted to be used for statistical analysis. Additional simulations were executed within the 'Sensitivity Analysis' program with the various formatted weather files. The sensitivity analysis enables the user to change a single variable such as planting population, planting date, weather year, soil type, or cultivar in a previously created experiment. Then, the experiment can be simulated again rather quickly with the new variable.

Year	Langdon	Carrington	Hettinger	Minot	Williston
2005	16 May	21 April	1 May†	1 May†	28 April
2006	09 May	20 April	1 May†	1 May†	05 May
2007	24 April	27 April	1 May†	1 May†	27 April
2008	29 April	15 April	1 May†	1 May†	23 April
2009	19 May	06 May	1 May†	07 May	23 April
2010	20 April	23 April	12 April	11 May	23 April
2011	17 May	06 May	2 May	11 May	13 May
2012	24 April	17 April	28 March	25 April	25 April
2013	16 May	13 May	23April	15 April	10 May
2014	14 May	02 May	22 April	15 May	09 May
2015	28 April	10 April	10 April	23 April	24 April
2016	02 May	12 April	01 April	02 May	03 May

Table 1.1. Planting dates needed to run model simulations for five locations in North Dakota in 2005-2016.

<sup>†</sup> Planting date not available. An assumed planting date of 1 May was used to run model simulations.

### Weather Data and Forecasting Approach

Daily minimum and maximum air temperature, rainfall, and solar radiation were collected from the North Dakota Agricultural Weather Network (NDAWN) for the period 1991-2016 for Carrington, Hettinger, Langdon, Minot, and Williston, ND (NDAWN, 2017a). The weather data were uploaded into the WeatherMan program within DSSAT. WeatherMan compiles the weather data with weather station details to be used within the experiment builder

(Batchelor et al., 2015). The NDAWN weather station information is presented in Table 1.2.

from the North Dakota Agricultural weather Network.				
Location	Latitude	Longitude	Elevation	
	°N	°W	m	
Carrington	47.51	-99.13	475	
Hettinger	46.01	-102.64	840	
Langdon	48.76	-98.35	492	
Minot	48.18	-101.29	542	
Williston	48.13	-103.74	649	

Table 1.2. Location, coordinates, and elevation of weather station locations obtained from the North Dakota Agricultural Weather Network.

The accuracy of crop growth can be predicted by updating DSSAT with measured weather data throughout the growing season. This concept was applied by Bannayan et al., 2003 for forecasting winter wheat yield. A similar approach was used in this study. Weather data had to be in the proper format before being uploaded into WeatherMan. Weather files were formatted with historic daily measured data from January 1 through ZGS 14, 45, and 61, then from that date supplemented with historical measured data throughout the remainder of the year. This procedure was done in three different ways (analogue, distribution, historic average), referred to as modeling approaches. The dates of these growth stages were estimated using the NDAWN wheat GDD calculator (NDAWN, 2017b), which predicts wheat growth stages through a method developed by Bauer et al. (1984).

The historic weather data was formatted in three different ways to forecast weather for the remainder of the season in order to determine the best technique to simulate yield with DSSAT. These were referred to as 'modeling approaches' and are described as analogue, distribution, and historical average. An analogue represents a previous weather year with a similar weather pattern as the current weather year up to the point in the season where future weather data is needed. The weather analogue program (WAP) within DSSAT uses historic weather conditions at a given location and estimates a previous year (analogue year) that is best representative of the current year. The WAP estimated an analogue year from measured weather data up to each growth stage. The weather data from the selected analogue year was then used after each growth stage through the remainder of the year to give a single outcome. The distribution modeling approach used daily historic weather data from 1991-2016 after each growth stage through the remainder of the year. This provided a distribution of outcomes with higher yields in favorable weather years, and poor yield in years when weather was suboptimal for HRSW growth. The historical average modeling approach used daily averages (1991-2016) after each growth stage through the remainder of the year. Figure 1.1 further illustrates how weather data were formatted.



Figure 1.1. Depiction of the different modeling approaches (distribution, historical average, analogue) using historic weather data to forecast the weather for the rest of the season and model wheat growth.

#### **Genetic Coefficients Development**

There were no genetic coefficients for any of the currently grown HRSW cultivars in North Dakota. Therefore, genetic coefficients for the cultivar, Glenn, were estimated using the generalized likelihood uncertainty estimation (GLUE) program within DSSAT. The GLUE program estimates the seven genetic coefficients using an established HRSW cultivar within the model as a template. The program then alters the coefficients of the template cultivar to match the crop measurements of the desired cultivar and give a new output with the estimated genetic coefficients. Crop measurements required to run GLUE include key phenological dates and yield or yield components (Hunt, 1993).

Glenn is a HRSW cultivar developed by North Dakota State University HRSW breeding program and released in 2005 (North Dakota Crop Improvement and Seed Association, 2005). Glenn was selected because many crop measurements needed for the GLUE program to estimate genetic coefficients were available. Information on planting date, harvest date, and grain yield were obtained from North Dakota Research Extension Center (REC) cultivar trials that included Glenn. Emergence date and anthesis date were estimated with the NDAWN wheat GDD calculator (NDAWN, 2017b). These data were collected for 2005-2016 at Carrington, Hettinger, Langdon, Minot, and Williston, ND. Yield trial data has previously indicated to be useful for calibrating crop models (Piper et al., 1998). Additional phenological measurements were collected from plots established in 2016. These plots were seeded at a rate of 2.9 million viable seeds per ha<sup>-1</sup> with a no-till 3P605NT drill (Great Plains Mfg. Inc., Salina, KS) at Prosper (47.00°N, -97.11°W) and Casselton (46.88°N, -97.23°W), ND. Twenty-five individual tillers were collected in order to obtain kernel and tiller weight. The genetic coefficients estimated for Glenn by the GLUE program are summarized in Table 1.3.

Table 1.3. Genetic coefficients for Glenn estimated by the generalized likelihood uncertainty estimator using data from North Dakota Research Extension Center cultivar trials.

P1V†	P2D‡	P5§	G1¶	G2#	G3††	PHINT‡‡
% day <sup>-1</sup>	% reduction hr <sup>-1</sup>	°C day <sup>-1</sup>	# g <sup>-1</sup>	mg kernel <sup>-1</sup>	g	°C day <sup>-1</sup>
30	40	475	18	35	2.0	76

† PIV- Vernalization coefficient.

<sup>‡</sup> P2D- Photoperiod coefficient.

§ P5- Thermal time from linear fill through maturity.

¶ G1- Kernel number per unit stem + spike weight at anthesis.

# G2- Kernel growth rate.

†† G3- Tiller death coefficient.

**‡‡PHINT-** Thermal time between leaf tip appearances .

### Soil Data

The DSSAT requires details of the soil where the simulations are performed. Site-specific

soil information for each location where yield data were obtained for cultivar calibration was not

available. Cultivar trials were conducted on or near REC's in Carrington, Hettinger, Langdon,

Minot, and Williston, ND so a predominant soil series was determined for each using the Web

Soil Survey (USDA-NRCS, 2017). The predominant soil series was chosen to give a good

representation of the soil type in the area. A soil profile cannot be created within DSSAT without specific soil profile characteristics, but the soil program within DSSAT has established default soil profiles with soil layer characteristics. An established soil series was chosen from DSSAT that best represented the predominant soil series for each location. The DSSAT soil profile selected may have varied slightly from the site-specific soil, but these profiles were fairly representative of the soil at the location. Soil series information for each location is reported in Table 1.4.

		U		
Location	Soil	Soil Taxonomy§	Slope	DSSAT Soil
	Series‡			Series
			%	
Carrington	Heimdal-	Coarse-loamy, mixed, superactive, frigid Calcic Hapludolls	0-5	Silty Loam
Hettinger	Shambo	Fine-loamy, mixed, superactive, frigid Typic Haplustolls	0-5	Loam
Langdon	Svea	Fine-loamy, mixed, superactive, frigid Pachic Hapludolls	0-5	Silty Loam
Minot	Forman	Fine-loamy, mixed, superactive, frigid Calcic Argiudolls	0-5	Clay Loam
Williston	Williams	Fine-loamy, mixed, superactive, frigid Typic Argiustolls	0-5	Loam

Table 1.4. North Dakota REC<sup>†</sup> locations with the predominant soil series, soil taxonomy, and slope with the corresponding DSSAT soil series.

† Research extension center.

‡ Soil data obtained from (USDA-NRCS, 2017).

§ Soil taxonomy listed on individual lines based on hyphenated soil series name.

#### **Statistical Analysis**

Model evaluation and verification was done using PROC REG in SAS 9.3 (SAS Institute,

Cary, NC) to determine the strength of relationship between observed and simulated anthesis

date and yield using full season weather data. Regression analysis was also used to determine the

strength of relationship between simulated anthesis date and grain yield using full season

weather data to simulated anthesis date and grain yield using forecasted weather with each

modeling approach from each growth stage. Root mean square error (RMSE) and coefficient of

variation (CV) were determined to explain model performance and the strength of the relationship was determined at the 90%, 95%, and 99% ( $p\leq0.10$ ,  $p\leq0.05$ ,  $p\leq0.01$ ) levels of significance.

Model accuracy was determined by computing the difference of anthesis date and grain yield between simulations using full season weather data and simulations forecasting weather using each modeling approach and from each growth stage. A smaller difference indicated better model predictions. The means of the differences were determined with PROC ANOVA in SAS. Means were separated using Fisher's protected least significant difference at the 95% (p≤0.05) level of confidence.

#### **Results and Discussion**

#### **Model Evaluation**

The DSSAT-CSM had to be properly validated before conducting experiments. Validation was performed by determining the accuracy between simulated crop growth and observed crop growth. In this study, the 'observed' data included anthesis date, which was determined using the NDAWN wheat GDD calculator (NDAWN, 2017b), and Glenn yield data from North Dakota REC cultivar trials. The NDAWN wheat GDD calculator determines anthesis date through a method developed by Bauer et al. (1984), which showed that wheat development could be accurately estimated using GDD with a base temperature of 0°C. Observed data was used to describe these variables in these comparisons. Anthesis date and yield were used to evaluate model accuracy in order to determine the ability of DSSAT to simulate the vegetative and reproductive development of Glenn. The validation procedure is performed in order to understand the expected error between simulations from the crop model and observed data (Batchelor et al., 2008).

Simulated anthesis date was consistently later than the observed data across all locations. The RMSE between simulated and observed anthesis date ranged between 6 days to 1 day across locations. A lower RMSE indicates better model performance. Overestimation of anthesis date has occurred in other DSSAT applications and may be due to misrepresentation of some of the genetic coefficients that govern the rate of vegetative growth (Boote et al., 2001). Another explanation may be the use of NDAWN to determine anthesis date. Glenn is an earlier maturing cultivar and may mature slightly earlier than what was determined by the NDAWN wheat GDD calculator. Combined across all locations, the relationship between simulated and observed anthesis date was significant at  $p \le 0.01$  ( $r^2 = 0.70$ ) (Table 1.5). This was similar across all locations except Williston, which resulted in a significant relationship between simulated and observed anthesis date at  $p \le 0.05$ .

Table 1.5. Comparison between simulated and observed anthesis date for five locations in North Dakota and combined across all locations and years in 2005-2016.

	Carrington		Hettinger		Langdon		Minot		Williston		Combined	
	Sim.†	Obs.‡	Sim.	Obs.	Sim.	Obs.	Sim.	Obs.	Sim.	Obs.	Sim.	Obs.
					d	ays afte	after planting					
Average§	71	68	74	71	70	63	69	63	69	65	71	66
RMSE¶	2		1		6		3		6		2	
CV#	3		2		9		4		9		3	
$r^2$ ††	0.92***		0.96**	** 0.73		***	0.66***		0.39**		0.70***	

<sup>†</sup> Simulated data from model simulations.

‡ Observed data determined using the NDAWN wheat GDD calculator (NDAWN, 2017b).

§ Averaged anthesis date for simulated and observed data in 2005-2016.

¶ Root mean square error.

# Coefficient of variation.

<sup>††</sup> Coefficient of determination between simulated and observed.

\*, \*\*, \*\*\* Significant at (P≤0.10), (P≤0.05), and (P≤0.01) respectively.

In this study, configuring DSSAT to simulate grain yield is critical, but also difficult due

to various environmental and agronomic variables that can affect yield throughout the growing

season. Simulated yield significantly related to observed yield combined across all locations with

a RMSE of 814 kg ha <sup>-1</sup> and $r^2$ =0.46 (Table 1.6). Previous research has shown the inability to
consistently simulate grain yield. Boote et al. (2002) realized the relationship between simulated
and observed yield ranged from $r^2 = 0.33$ to 0.74 with simulations using different cultivars in
North Carolina. Similar results occurred in this research because the relationship between
simulated and observed yield ranged between $r^2 = 0.16$ to 0.61 across locations (Table 1.6). The
relationship between simulated and observed yield at Carrington and Williston were significant
at p $\leq$ 0.01, while Hettinger and Minot were significant at p $\leq$ 0.05. Simulated yield at Langdon was
not well related to observed yield with $r^2=0.16$ . For these locations, the RMSE between
simulated and observed grain yield was 705, 548, 806, 733, and 617 kg ha <sup>-1</sup> , respectively (Table
1.6). The variability between locations may be due to differences in the management practices
between locations that can impact environment x cultivar interactions and cannot be accurately
accounted for in the model (Adiku et al., 2017).

	Carrington		Hettinger		Langdon		Minot		Williston		Combined	
	Sim.†	Obs.‡	Sim.	Obs.	Sim.	Obs.	Sim.	Obs.	Sim.	Obs.	Sim.	Obs.
	kg ha-1											
Average§	4023	4028	4068	3626	4594	5082	4520	4225	3426	3165	4126	4025
RMSE¶	705		806		617		733		548		814	
CV#	17		22		12		17		17		20	
$r^2$ ††	0.52***		0.52**		0.16		0.50**		0.61***		0.46***	

Table 1.6. Comparison between simulated and observed yield for five locations in North Dakota and combined across all locations and years in 2005-2016.

<sup>†</sup> Simulated data from model simulations.

‡ Observed data obtained from North Dakota Research Extension Center cultivar trials.

§ Averaged yield for simulated and observed data in 2005-2016.

¶ Root mean square error.

# Coefficient of variation.

†† Coefficient of determination between simulated and observed.

\*, \*\*, \*\*\* Significant at (P≤0.10), (P≤0.05), and (P≤0.01) respectively.

Overall, the ability to simulate anthesis date and grain yield with DSSAT was deemed

adequate across all locations. Therefore, it appeared reasonable to apply the model to predict

yield from various growth stages using historic weather data, while understanding the expected error from model simulations. The growth and development processes of crop models and how they respond to environmental conditions can explain the error between the observed data and model simulations. These plant processes are determined by algorithms that respond to the environmental variables (temperature, N status, and moisture) input within the model, which may not adequately account for a plant response when environmental thresholds are exceeded (Bannayan et al., 2003). Model simulations can be improved by better adjusting soil and genetic components. The soil profiles used in this study represented a generalized profile and may account for some error from model simulations. Also, the genetic coefficients for Glenn were an estimation by the GLUE program. Improved cultivar response can be achieved when coefficients are developed using more intensive physical measurements during plant development.

#### Assessment of Model Simulations using Different Forecasting Methods

Historic weather data was used in the respective year in order to conduct simulations. Measured weather data was logged from the observed planting date through each growth stage, then supplemented with historic weather data to forecast the weather for the remainder of the season. In order to forecast weather, the weather data were formatted in three modeling approaches to determine the best method for predicting crop growth. The three modeling approaches were referred to as distribution, historical average, and analogue, which are previously described. In this study, simulated anthesis date and grain yield using full season weather data (FSWD) were compared to simulated anthesis date and grain yield with forecasted weather using the three modeling approaches (regardless of growth stage at which forecasting began) for all locations and years of model simulation. The smallest difference between simulated anthesis date and grain yield using FSWD and forecasted weather indicated better

model performance. Therefore, the optimum approach for using historic weather data to forecast weather for the remainder of the season was determined. The distribution approach used multiple weather years, which provided a range of outcomes. Therefore, the mean of these simulated values were used for comparisons.

There were significant differences for simulated anthesis date and grain yield when comparing the simulations using each modeling approach. Combined across all locations, the average difference between simulated anthesis date using FSWD and simulated anthesis date using the distribution, historical average, and analogue modeling approaches were 1.0, 0.9, and 1.5 days, respectively (Table 1.7). Data indicated the strength of relationship between anthesis date using FSWD and simulated anthesis date using each modeling approach varied across all locations. However, simulations using distribution and historical average approaches consistently agreed the best with simulations using FSWD with  $r^2 = 0.87$  and 0.90 for distribution and historical average modeling approaches, respectively ( $p \le 0.01$ ) (Table 1.8).

The ability to adequately simulate grain yield using the three modeling approaches varied greatly. The average difference of simulated grain yield using FSWD and simulated grain yield using forecasted weather with each modeling approach was 486, 695, and 648 kg ha<sup>-1</sup>, for distribution, historical average, and analogue approaches, respectively (Table 1.7). The strength of relationship between simulated grain yield using FSWD and each modeling approach was  $r^2=0.64$ , 0.61, and 0.49 for distribution, historical average, and analogue aperoaches, respectively (Table 1.9). Data indicated simulating grain yield using the distribution and historical average approaches were the most effective since the relationship between simulated grain yield with FSWD and these two approaches were significant at p $\leq$ 0.01. However, simulations using the analogue modeling approach were not as effective since the same relationship was only

significant at  $p \le 0.05$ . The analogue approach may be less effective due to the inability for shortrange weather to match a weather year accurately. This can most likely be attributed to the lack of measured data available since it has only been available over roughly the last 30 yrs., as well as the inability to account for any variability between years (Van den Dool, 1988).

In this study, the optimum technique to simulate crop growth using historic weather data to forecast the weather for the remainder of the season was through a distribution approach. Overall, all three modeling approaches simulated anthesis date satisfactorily when compared to simulated anthesis date using FSWD. The greatest RMSE between simulated anthesis date using FSWD was found using the analogue modeling approach; however, variations between the modeling approaches were minor. Simulating grain yield with the distribution approach resulted in the lowest RMSE from simulated grain yield using FSWD with 348 kg ha<sup>-1</sup> combined across all locations (Table 1.9). In general, crop development was better represented using the distribution approach because the variability of weather conditions that affect plant growth are taken into account. A historical average or analogue approach used a single data set and may not have effectively taken into account the possible variability of weather for the remainder of the season. This was similar to observations in previous research (Thorton et al., 1997; Bannayan et al., 2003).

•omemea aeross an	iotanomo ana	J <b>ea</b> ls III <b>2</b> 000 <b>2</b> 010	
		Anthesis Date	Yield
		Days	kg ha <sup>-1</sup>
Distribution		1	486
Historical Avg.		0.9	695
Analogue		1.5	648
	LSD <sub>0.05</sub>	0.2	105

Table 1.7. Average difference<sup>†</sup> for anthesis date and grain yield between simulations using full season weather data and forecasted weather using each modeling approach combined across all locations and years in 2005-2016

<sup>†</sup> Averages calculated by determining the difference between simulations using FSWD and simulations using each modeling approach, combined across all locations and years.

Table 1.8. Comparison between simulated anthesis date using full season weather data and simulated anthesis date using three modeling approaches (distribution, historical average, analogue) in 2005-2016 for five locations.

	Carrington		Hettinger		Langdon		Minot		Williston		Combined	
	RMSE	r <sup>2</sup>	RMSE	r <sup>2</sup>	RMSE	r <sup>2</sup>	RMSE	r <sup>2</sup>	RMSE	r <sup>2</sup>	RMSE	$r^2$
	Days		Days		Days		Days		Days		Days	
Distribution	1.0	0.95***	1.3	0.92***	1.3	0.88***	1.2	0.80***	0.7	0.97***	1.0	0.87***
Historical Ave.	0.9	0.96***	1.3	0.93***	1.6	0.81***	1.4	0.74***	0.5	0.98***	0.9	0.90***
Analogue	1.3	0.92***	1.9	0.84***	2.3	0.61***	1.1	0.83***	0.8	0.95***	1.0	0.87***

† Zadoks Growth Stage

\*,\*\*,\*\*\* Significant at (P≤0.10), (P≤0.05), and (P≤0.01) respectively

Table1.9. Comparison between simulated grain yield using full season weather data and simulated grain yield using three modeling approaches (distribution, historical average, analogue) in 2005-2016 for five locations.

unde modeling upprodenes (distribution, instorieur d'erage, undrogde) in 2003-2010 for interioedions.												
	Carrington		Hettinger		Langdon		Minot		Williston		Combined	
	RMSE	r <sup>2</sup>	RMSE	$r^2$	RMSE	r <sup>2</sup>						
	kg ha <sup>-1</sup>		kg ha <sup>-1</sup>		kg ha <sup>-1</sup>		kg ha <sup>-1</sup>		kg ha <sup>-1</sup>		kg ha <sup>-1</sup>	
Distribution	463	0.70***	858	0.42**	539	0.81***	433	0.70***	470	0.76***	348	0.64***
Historical Ave.	527	0.60***	905	0.35**	718	0.66***	526	0.56***	510	0.72***	360	0.61***
Analogue	570	0.54***	1006	0.20	1059	0.26***	772	0.06	422	0.81***	411	0.49**

<sup>†</sup>Zadoks Growth Stage

\*,\*\*,\*\*\* Significant at (P≤0.10), (P≤0.05), and (P≤0.01) respectively
#### Assessment of Model Simulations from Multiple Growth Stages

Measured weather data were logged up to three growth stages (ZGS 14, 45, 61) for the respective year and forecasted weather data was supplemented from that date through the remainder of the growing season (regardless of modeling approach). Simulated anthesis date and grain yield using FSWD were compared to simulated anthesis date and grain yield with forecasts made from ZGS 14, 45, and 61 for all locations and years of model simulation. The ability to effectively predict grain yield early in the season can assist producers in making a range of more informed management decisions (Bannayan et al., 2003).

In theory, simulations that forecasted anthesis date from ZGS 61 should have had zero deviation from the simulated anthesis date using FSWD. Simulated anthesis date resulted in an RMSE of 0.3 days with forecasted weather from ZGS 61 compared to simulated anthesis date using FSWD indicating some error in the model (Table 1.10). However, the relationship between simulated anthesis date using FSWD to forecast anthesis date from ZGS 61 was very high ( $r^2$ =0.99), indicating satisfactory performance. The maximum difference between simulated anthesis date using FSWD and forecasted anthesis date from ZGS 14 was 2.0 days (Table 1.11). This was significantly greater compared to the difference in anthesis date when forecasting from ZGS 45 and 61 with 1.0 and 0.5 days, respectively (Table 1.11). Overall, the deviation from simulated anthesis date using FSWD was minor. Combined across all locations, anthesis date was significant between simulated anthesis date using FSWD and forecasted anthesis date using FSWD and forecasted anthesis date using FSWD and forecasted anthesis date using FSWD was minor. Combined across all locations, anthesis date was significant between simulated anthesis date using FSWD and forecasted anthesis date using FSWD and forecasted anthesis date from ZGS 14 and 45 at p≤0.01 with  $r^2$ =0.65 and 0.94, respectively (Table 1.10).

	Carri	ngton	Hetti	inger	Lan	gdon	М	inot	Wil	liston	Com	bined
	RMSE	r <sup>2</sup>	RMSE	r <sup>2</sup>	RMSE	r <sup>2</sup>	RMSE	r <sup>2</sup>	RMSE	r <sup>2</sup>	RMSE	r <sup>2</sup>
	Days		Days		Days		Days		Days		Days	
ZGS14	2.0	0.81***	2.5	0.73***	2.5	0.53***	2.0	0.47***	1.0	0.92***	1.7	0.65***
ZGS 45	1.1	0.94***	1.1	0.95***	1.7	0.79***	1.3	0.77***	0.4	0.98***	0.7	0.94***
ZGS 61	0.5	0.99***	0.7	0.98***	1.0	0.92***	0.8	0.91***	0.6	0.97***	0.3	0.99***

Table 1.10. Comparison between simulated anthesis date using full season weather data and simulated anthesis date with forecasted weather from three growth stages in 2005-2016 for five locations.

† Zadoks Growth Stage

\*,\*\*,\*\*\* Significant at (P≤0.10), (P≤0.05), and (P≤0.01) respectively

Table 1.11. Average difference<sup>†</sup> for anthesis date and grain yield between simulations using full season weather data and forecasted weather from ZGS 14, 45, and 61.

	Anthesis Date	Yield
	Days	kg ha <sup>-1</sup>
ZGS 14‡	2.0	890
ZGS 45	1.0	610
ZGS 61	0.5	331
LSD <sub>0.05</sub>	0.2	115

\* Averages calculated by determining the difference between simulations using FSWD and simulations using each modeling approach. Then combined across all locations and averaged.
‡ Zadoks growth stage. In contrast to the prediction of anthesis date, DSSAT was weaker in predicting grain yield. Generally, predicting yield from any growth stage resulted in an overestimation compared to simulations using FSWD. The maximum deviation of yield resulted when forecasts were made from ZGS 14 and the deviation in yields were reduced at later growth stages. The average difference between grain yield using FSWD and forecasted grain yield from ZGS 14, 45 and 61 was 890, 610, and 331 kg ha<sup>-1</sup>, respectively (Table 1.11). A significant relationship was found for simulations with forecasts from ZGS 45 ( $p\leq0.05$ ) and 61 ( $p\leq0.01$ ); however, not from ZGS 14 (Table 1.12). The strength of the relationship between simulated grain yield using FSWD and forecasted grain yield using FSWD and forecasted grain yield from ZGS 14, 45, and 61 was  $r^2=0.24$ , 0.41, and 0.86, respectively (Table 1.12). Previous research reported yield predication within 15% when grain yield was forecasted from simulations conducted halfway through the growing season (Thorton et al., 1997). Similar results were obtained with this study with forecasted grain yield from ZGS 45 within 7% of full season grain yield and within 5% when forecasted from ZGS 61 (Data not shown).

Across all locations, model performance improved as measured data was input as the growing season progressed. Anthesis date was significant ( $p \le 0.01$ ) with the relationship between simulated using FSWD and forecasted from ZGS 14. Improved performance with forecasted anthesis date at ZGS 45 suggested forecasts would have the greatest accuracy when conducted at this growth stage. However, forecasted anthesis date resulted in an average difference from simulated anthesis date using FSWD of only 2 days, indicating only minor differences when simulations are conducted with forecasted weather earlier in the growing season. The relationship between simulated yield using FSWD and simulated yield using forecasted weather also improved as measured weather data was input later into the growing season. In this study, data indicated grain yield should not be simulated with forecasted weather from ZGS 14 since

the relationship was not significant with simulations using FSWD. The impact weather will have on grain yield is unknown early in the growing season, since yield is not yet determined in the plant. Predicting yield from ZGS 14 may be unlikely for this reason. However, as crop development progresses, grain yield is further determined; therefore, weather will have less impact and yield prediction will be more accurate (Wiersma and Ransom, 2005).

#### **Assessment of Forecasted Crop Growth**

In this study, data indicated there was no significant interaction when supplementing each of the modeling approaches (distribution, historical average, analogue) to forecast anthesis date or grain yield from each growth stage (ZGS 14, 45, 61). The strength of relationship between simulated anthesis date using FSWD and simulated anthesis date using each modeling approach to forecast from all growth stages varied across locations. However, simulations using distribution and historical average modeling approaches supplemented at ZGS 45 consistently provided the greatest relationship with simulations using FSWD for anthesis date (Table 1.13). Similar results were observed when predicting grain yield. The best relationship consistently occurred with simulations using the distribution and historical average approach when supplemented from ZGS 61 (Table 1.14). Simulated using these approaches from ZGS 61 was significant at  $p \le 0.01$  and when supplemented from ZGS 45 the relationship was still significant ( $p \le 0.05$ ). Simulating crop growth using the analogue modeling approach to forecast weather was the least effective and should not be used.

	Carri	ngton	Hetti	nger	Lang	gdon	Min	ot	Willi	ston	Con	nbined
	RMSE	r <sup>2</sup>	RMSE	r <sup>2</sup>	RMSE	$r^2$	RMSE	r <sup>2</sup>	RMSE	r <sup>2</sup>	RMSE	r <sup>2</sup>
	kg ha <sup>-1</sup>		kg ha <sup>-1</sup>		kg ha <sup>-1</sup>		kg ha <sup>-1</sup>		kg ha <sup>-1</sup>		kg ha <sup>-1</sup>	
ZGS14	789	0.11	1083	0.07	1226	0.00	790	0.01	773	0.35**	503	0.24
ZGS 45	418	0.75***	1041	0.14	850	0.52***	686	0.26*	550	0.67***	444	0.41**
ZGS 61	189	0.95***	710	0.60***	464	0.86***	360	0.80***	357	0.86***	215	0.86***

Table 1.12. Comparison between simulated grain yield using full season weather data and simulated grain yield with forecasted weather from three growth stages in 2005-2016 for five locations.

<sup>†</sup>Zadoks Growth Stage

\*,\*\*,\*\*\* Significant at (P≤0.10), (P≤0.05), and (P≤0.01) respectively

Table 1.13. Comparison between simulated anthesis date using full season weather data and simulated anthesis date using three approaches to forecast weather from three growth stages in 2005-2016 for five locations.

	Carr	ington	Hett	tinger	Lan	gdon	М	inot	Wil	liston	Com	bined
	RMSE	$r^2$	RMSE	$r^2$	RMSE	r <sup>2</sup>	RMSE	r <sup>2</sup>	RMSE	$r^2$	RMSE	r <sup>2</sup>
Variable	Days		Days		Days		Days		Days		Days	
Distribution ZGS† 14	1.9	0.80***	2.3	0.77***	1.9	0.72***	1.9	0.53***	1.3	0.86***	1.7	0.65***
Distribution ZGS 45	0.8	0.96***	1.1	0.95***	1.6	0.80***	1.2	0.79***	0.8	0.95***	0.8	0.92***
Distribution ZGS 61	0.6	0.98***	0.7	0.98***	0.7	0.96**	0.8	0.92***	0.7	0.96***	0.6	0.96***
Historical Ave. ZGS 14	2.1	0.79***	2.1	0.81***	2.7	0.47**	2.0	0.47**	1.2	0.89***	1.8	0.61***
Historical Ave. ZGS 45	0.8	0.97***	1.1	0.95***	1.8	0.76***	1.3	0.77***	0.5	0.97***	0.8	0.92***
Historical Ave. ZGS 61	0.4	0.99***	0.6	0.99***	0.7	0.96***	0.6	0.95***	0.5	0.98***	0.4	0.98***
Analogue ZGS 14	2.2	0.76***	3.3	0.55***	3.3	0.2	1.9	0.50**	1.2	0.8832***	1.8	0.58***
Analogue ZGS 45	2.2	0.75***	1.7	0.87***	2.2	0.65***	1.3	0.77***	0.7	0.96***	0.9	0.91***
Analogue ZGS 61	0.4	0.99***	1.1	0.95***	1.6	0.80***	1.1	0.84***	1.0	0.92***	0.7	0.94***

*†* Zadoks Growth Stage

\*,\*\*,\*\*\* Significant at (P≤0.10), (P≤0.05), and (P≤0.01) respectively

	Carri	ington	Hett	inger	Lan	gdon	Mii	not	Willi	ston	Con	nbined
	RMSE	$r^2$	RMSE	r <sup>2</sup>	RMSE	r <sup>2</sup>	RMSE	$\mathbf{r}^2$	RMSE	$r^2$	RMSE	$r^2$
Variable	kg ha <sup>-1</sup>		kg ha <sup>-1</sup>		kg ha <sup>-1</sup>		kg ha⁻¹		kg ha⁻¹		kg ha⁻¹	
Distribution ZGS† 14	816	0.05	1081	0.07	1218	0.01	741	0.13	794	0.32	513	0.21
Distribution ZGS 45	451	0.71***	1004	0.20	658	0.71***	608	0.42**	629	0.57***	440	0.42**
Distribution ZGS 61	217	0.93***	723	0.59***	390	0.90***	251	0.90**	358	0.86***	223	0.85***
Historical Ave. ZGS 14	797	0.10	1098	0.04	1219	0.01	744	0.13	742	0.40**	531	0.15
Historical Ave. ZGS 45	441	0.72***	990	0.22	765	0.61***	666	0.30	697	0.47**	437	0.43**
Historical Ave. ZGS 61	296	$0.88^{***}$	767	0.53***	484	0.84***	345	0.81***	474	0.76***	206	0.87***
Analogue ZGS 14	798	0.09	1099	0.04	1226	0.00	792	0.01	894	0.13	517	0.20
Analogue ZGS 45	497	0.65***	1099	0.04	1097	0.20	794	0.00	578	0.64***	494	0.27
Analogue ZGS 61	251	0.91***	716	0.59***	703	0.67***	617	0.40**	405	0.82***	305	0.72***

Table 1.14. Comparison between simulated grain yield using full season weather data and simulated grain yield using three approaches to forecast weather from three growth stages in 2005-2016 for five locations.

† Zadoks Growth Stage

\*,\*\*,\*\*\* Significant at (P $\leq$ 0.10), (P $\leq$ 0.05), and (P $\leq$ 0.01) respectively

In this study, supplementing the distribution and historical average approach to forecast grain yield provided similar results across all locations, therefore, either would be a suitable method to forecast weather and predict crop growth. However, the distribution approach may improve yield forecasting because a generalized average is not being used, but rather a frequency of the distribution can be determined from the range of outcomes. The information obtained from these outcomes can be used as a decision support system to make improved management decisions (Bannayan et al., 2003).

As plant development progresses throughout the growing season, the precision of the simulations will increase as the distribution of the simulated values narrows around the likely outcome. In this study, the precision of simulations using the distribution approach improved when anthesis date and grain yield were forecasted from ZGS 45 and 61. As plant development progressed the range between the projected outcomes decreased for anthesis date with an average range of 10, 6, and 2 days for ZGS 14, 45, and 61, respectively (Table 1.15). Similar results were obtained for grain yield with the range between the projected outcome decreased by 3305, 1910, and 988 kg ha<sup>-1</sup> for ZGS 14, 45, and 61, respectively (Table 1.16).

Overall, the DSSAT-CSM can effectively predict anthesis date within 1-2 days from ZGS 14 and 45. Therefore, the crop model may be a beneficial decision support system to effectively determine when anthesis date will occur. This would be beneficial for a producer to make management decisions that occur around this time, such as a foliar fertilizer or pesticide application. Also, DSSAT is most accurate in predicting grain yield from ZGS 45 and 61. Therefore, the model may be a useful management tool for producers to make an informed decision for an additional N application to increase GPC if predicted yield is high. However,

since grain yield cannot be accurately predicted until ZGS 45, a small time frame is available to

make the decision whether or not this additional application should be conducted.

Table 1.15. Mean and range of simulated anthesis date using the distribution approach to forecast weather from ZGS<sup>†</sup> 14, 45, and 61 for five locations in North Dakota combined across all years in 2005-2016.

	ZG	S 14	ZGS	S4 45	ZG	S 61	_	
	Mean‡	Range§	Mean	Range	Mean	Range	Obs.¶	Sim.#
				-days after p	olanting			
Carrington	72	68 - 77	71	69 - 74	71	70 - 73	68	71
Hettinger	75	70 - 80	75	72 - 78	74	74 - 75	63	70
Langdon	71	67 - 77	70	68 - 75	70	69 - 72	72	74
Minot	70	66 - 77	69	67 - 74	69	68 - 71	63	69
Williston	70	67 - 77	70	68 - 74	70	69 - 71	65	69
Combined	71	68 - 78	71	69 - 75	71	70 - 72	65	71
Avg. Range		10		6		2		

† Zadoks growth stage.

‡ Mean anthesis date in 2005-2016 from simulations using the distribution approach to forecast weather from the respective growth stage.

§ Average range of anthesis date in 2005-2016 from simulations using the distribution approach to forecast weather from the respective growth stage.

¶ Mean of anthesis date in 2005-2016 determined from NDAWN wheat GDD calculator.

# Mean of anthesis date in 2005-2016 from simulations using full season weather data.

Table 1.16. Mean and range of simulated grain yield using the distribution approach to forecast weather from ZGS<sup>†</sup> 14, 45, and 61 for five locations in North Dakota combined across all years in 2005-2016.

_	ZGS 14		ZGS4 45		2	ZGS 61		
	Mean‡	Range§	Mean	Range	Mean	Range	Obs.¶	Sim.#
				kg h	a <sup>-1</sup>			
Carrington	3951	2161 - 5559	4099	3359 - 4874	4034	3607 - 4456	4028	4023
Hettinger	3885	2000 - 5411	4012	3119 - 5059	4020	3596 - 4441	5082	4594
Langdon	4563	2668 - 5959	4679	3642 - 5683	4711	4131 - 5319	3717	4034
Minot	4409	2758 - 5691	4559	3646 - 5498	4562	4074 - 5084	4225	4520
Williston	3820	2285 - 5774	3672	2782 - 4985	3541	3120 - 4169	3165	3426
Combined	4125	2374 - 5679	4204	3310 - 5220	4174	3706 - 4694	4083	4068
Avg. Range		3305		1910		988		

† Zadoks growth stage.

‡ Mean yield in 2005-2016 from simulations using the distribution approach to forecast weather from the respective growth stage.

§ Average range of yield in 2005-2016 from simulations using the distribution approach to forecast weather from the respective growth stage.

¶ Mean of observed yield in 2005-2016 from North Dakota Research Extension Centers.

# Mean of simulated yield in 2005-2016 from simulations using full season weather data.

#### Conclusion

The ability of DSSAT-CSM to simulate plant development varied across locations, but overall the crop model simulated anthesis date and grain yield with adequate accuracy for HRSW in North Dakota. Simulation error can most likely be attributed to the use of generalized soil types and estimated genetic coefficients, which can be improved with increased physical soil and plant measurements.

The three approaches used to forecast weather for the remainder of the season were all able to forecast anthesis date similarly. However, the greatest precision was obtained when simulating grain yield with the distribution and historical average modeling approaches. The analogue approach was the least effective, which may be explained by the inability to account for the variability of weather between years with this approach. Yield simulations using the distribution and historical average modeling approaches were similar and could predict yield with the greatest accuracy compared to yield simulations using FSWD. However, simulations using the distribution approach may be more beneficial because a frequency of the distribution from the range of outcomes can be determined in addition to a generalized average.

Across all locations, model performance improved as measured weather data was input as the growing season progressed. The associated error with simulated anthesis date using forecasted weather from ZGS 14 and ZGS 45 was minimal. In this study, grain yield was not accurately simulated with forecasted weather from ZGS 14 and should not be attempted. This may be due to yield still undetermined within the plant and the weather for the remainder of the season being unknown, which can have a major impact on plant development. Simulated grain yield with forecasted weather from ZGS 45 and 61 had an average RMSE of 444 and 215 kg ha<sup>-1</sup>

from simulated yield using FSWD. Grain yield was predicted with the greatest accuracy with forecasted weather from ZGS 61.

The method implemented to update measured weather data during the growing season was time consuming and would be very difficult on a large scale. However, an updating system that is incorporated into the model that updates weather data on a daily or weekly basis might provide more accurate forecasting with DSSAT (Bannayan et al., 2003). The information obtained in these yield predictions can provide an effective decision support system that allows producers to make improved management decisions, especially during the later part of the growing season. The results from this study indicate that an informed decision can be better made with simulations conducted from ZGS 45 or later since yield is not accurately predicted until this time. Therefore, a management decision to apply a late-season foliar N application could be determined during the growing season. However, since predictions cannot be made until at least ZGS 45, this decision would have to be made relatively quickly in order to achieve the most success from the additional N application.

## References

- Adiku, S.G., B.S. Freduah, F. Gbefo, A.Y. Kamara, and D.S. MacCarthy. 2017. Using CERES-Maize and ENSO as decision support tools to evaluate climate-sensitive farm management practices for maize production in the northern regions of Ghana. Frontiers in Plant Sci. 8:1-13.
- Aggarwal, P.K., A. Bala, B. Banerjee, A. Bhatia, S. Chander, M.G. Daryaei, N. Kalra, H. Pathak, and S. Rani. 2006. Infocrop: A dynamic simulation model for the assessment of crop yields, losses due to pests and environmental impact of agro-ecosystems in tropical environments. II. Performance of the model. Agric. Syst. 89:47-67.

- Aggarwal, P.K., and N. Kalra. 1994. Analysing the limitations set by climatic factors, genotype, water and nitrogen availability on productivity of wheat. II. Climatically potential yields and optimal management strategies. Field Crops Res. 38:93-103.
- Ahuja, L.R., D.J. Lyon, L. Ma, D.C. Nielsen, and S.A. Saseendran. 2013. Simulated yield and profitability of five potential crops for intensifying the dryland wheat fallow production system. Agric. Water Management. 116:175-192.
- Antle, J.M., B.O. Basso, K.J. Boote, R.T. Conant, I. Foster, H.J. Godfray, M. Herrero, R.E.
  Howitt, S. Janssen, J.W. Jones, B.A. Keating, R. Munoz-Carpena, C.H. Porter, C.
  Rosenzweig, and T.R. Wheeler. 2016. Brief history of agricultural system models. Ag.
  Systems. 155: 240-254.
- Arnall, D.B., K. Girma, G.V. Johnson, W.R. Raun, J.B. Solie, M.L. Stone, et al. 2006.
  Relationship between coefficient of variation measured by spectral reflectance and plant density at early growth stages in winter wheat. J. Plant Nutr. 29:1983–1997.
- Balaghi, R., H. Eerens, M. Jlibene, and B. Tychon. 2008. Empirical regression models using NDVI, rainfall, and temperature data for the early prediction of wheat grain yields in Morocco. Int. J. Appl. Earth Obs. Geoinf. 10:438–452.
- Baltensperger, D., J. Blumenthal, K.G. Cassman, S. Mason, and A. Pavlista. 2008. Importance and effect of nitrogen on crop quality and health. Misc. Publ. University of Nebraska-Lincoln, Lincoln, NE.
- Bannayan, M., N.J. Crout, and G. Hoogenboom. 2003. Application of the CERES-wheat model for within-season prediction of winter wheat yield in the United Kingdom. Agron. J. 95:114-125.

- Barnett, D.N., M. Collins, G.S. Jones, J.M. Murphy, D.H. Sexton, D.A. Stainforth, and M.J. Webb. 2004. Quantification of modeling uncertainties in a large ensemble of climate change simulations. Nat. 429:768–772.
- Batchelor, W.D., K.J. Boote, A.J. Gijsman, L.A. Hunt, G. Hoogenboom, J.W. Jones, C.H. Porter,
  J.T. Ritchie, U. Singh, and P.W. Wilkens. 2015. Decision Support System for
  Agrotechnology Transfer (DSSAT) Version 4.6 (www.DSSAT.net). DSSAT Foundation,
  Prosser, WA.
- Batchelor, W.D., H.G. Booltink, B.J. van Alphen, J.O. Paz, J.J. Stoorvogel, and R. Vargas. 2001. Tools for optimizing management of spatially variable fields. Agric. Syst. 70:445-476.
- Batchelor, W.D., K.C. DeJonge, A.L. Kaleita, J.O. Paz, and K.R. Thorp. 2008. Methodology for the use of DSSAT models for precision agriculture decision support. Comput. Electron. Agric. 65:276-285.
- Batchelor, W.D., K.J. Boote, G. Hoogenboom, L.A. Hunt, J.W. Jones, C.H. Porter, et al. 2003. The DSSAT cropping system model. Europ. J. Agron. 18:235-265.
- Bauer, A, A.L. Black, and A.B. Frank. 1984. Estimation of spring wheat leaf growth rates and anthesis from air temperature. Agron. J. 829–835.
- Boote, K.J., J.W. Jones, and N.B. Pickering. 1996. Potential uses and limitations of crop models. Agron. J. 88:704–716.
- Boote, K.J., G. Hoogenboom, and J.W. Jones. 1998. Simulation of crop growth: CROPGRO model. *In*: R.B. Curry and R.M. Peart (ed.) Agricultural systems modeling and simulation. Marcel Dekker, New York, NY. p. 651-692.

- Boote, K.J., G. Hoogenboom, A. Irmak, J. Jones, Mavromatis, and D. Shinde. 2001. Developing genetic coefficients for crop simulation models with data from crop performance trials. Crop Sci. 41:40-51.
- Boote, K.J., G. Hoogenboom, J. Jones, T.K. Mavromatis, and G.G. Wilkerson. 2002.
   Repeatability of model genetic coefficients derived from soybean performance trials across different states. Crop Sci. 42:76-89.
- Boote, K.J., and L.A. Hunt. 1998. Data for model operation, calibration and evaluation. *In*: G.
  Hoogenboom, P.K. Thorton, and G.Y. Tsuji (eds.) Understanding options for agricultural production. Kluwer Academic Publishers/ICASA, Dordrecht, the Netherlands. p. 9-40.
- Bowen, W.T., J.E. Brink, J. Brock, G. Farmer, A.C. Ravelo, P.K. Thorton, and P.W. Wilkens.
  1997. Estimating millet production for famine early warning: an application of crop simulation modelling using satellite and ground-based data in Burkina Faso. Agric.
  For. Meteorol. 83:95-112.
- Clarke, T.R., A.N. French, D.J. Hunsaker, P.J. Pinter Jr., and J.W. White. 2010. Evaluation of the CSM-CROPSIM-CERES-Wheat model as a tool for crop water management. Trans. ASABE Publ. 53:87-102.
- Drury, C.F., G. Hoogenboom, T.Z. Li, and J.Y. Yang. 2015. Evaluation of the DSSAT-CSM for simulating yield and soil organic C and N of a long-term maize and wheat rotation experiment in the Loess Plateau of Northwestern China. Agric. Syst. 135:90–104.
- Finney, K.F., H.C. Freyer, W. Meyer, and F.W. Smith. 1957. Effect of foliar spraying of pawnee wheat with urea solution on yield, protein content, and protein quality. Agron. J. 49:341-347.

- Hansen, J.W., and J.W. Jones. 2000. Scaling-up crop models for climate variability applications. Agric. Syst. 65:43-72.
- Hunt, L.A. 1993. Designing improved plant types: A breeder's viewpoint. *In*: K. Metselaar, F.Penning de Vries and P. Teng (eds.) Systems approaches for agricultural development.Kluwer Academic Press, Boston, MA. p. 3-17.
- NDSU Foundation Seed. 2005. NDSU Agriculture Glenn Hard Red Spring Wheat. Available at http://ndsuresearchfoundation.org/files/pdf/Ag%20Brochures/Glenn\_brochure.pdf (accessed 15 Oct. 2017).
- North Dakota Agricultural Weather Network. 2017a. Available at https://ndawn.ndsu.nodak.edu/ weather-data-daily.html (accessed 19 Oct. 2017). Fargo, ND.
- North Dakota Agricultural Weather Network. 2017b. Available at https://ndawn.ndsu.nodak.edu/ wheat-growing-degree-days.html (accessed 19 Oct. 2017). Fargo, ND.
- Parton, W.J., C.V. Cole, and J.W. Steward. 1988. Dynamics of C, N, P and S in grassland soils: a model. Biogeochem. 5:109-131.
- Parton, W.J., C.V. Cole, D.S. Ojima, and D.S. Schimel.1994. A general model for soil organic matter dynamics: sensitivity to litter chemistry, texture and management. *In*: R.B. Bryant and R.W. Arnold (eds.) Quantitative modeling of soil forming processes. SSSA Spec.
  Publ. 39. ASA, CSSA and SSA, Madison, WI. P. 147-167.
- Piper, E.L., K.J. Boote, and J.W. Jones. 1998. Evaluation and improvement of crop models using regional cultivar trial data. Trans ASAE 14:435-446.
- Priestley, C.B., and R.J. Taylor. 1972. On the assessment of surface heat flux and evaporation using large-scale parameters. Monthly Weath. Rev. 100:81–92.

- Ritchie, J.T. 1998. Soil water balance and plant stress. *In*: G.Y. Tsuji et al., (ed.) Understanding options for agricultural production. Kluwer Acad., Norwell, MA.
- Seligman, N.C., and H. Van Keulen. (1981). PAPRAN: A simulation model of annual pasture production limited by rainfall and nitrogen. P. 192–221. *In*: M.J. Frissel and J.A. Van Veen (eds.) Simulation of nitrogen behavior of soil plant systems. PUDOC, Wageningen, the Netherlands.
- USDA-Natural Resources Conservation Service. 2017. Available at http://websoilsurvey.nrcs. usda.gov/ app/HomePage.htm (accessed 19 Oct. 2017). USDA-NRCE, Washington DC, PA.
- Van Den Dool, H.M. 1989. A new look at weather forecasting through analogues. Monthly Am. Meteorl. Soc. 117:2230-2247.
- Wiersma, J.J., and J.K. Ransom. 2005. The Small Grains Field Guide. North Dakota State Univ.Ext. Publ. A290 and Univ. of Minnesota Ext. Publ. 0788-S. Fargo, ND, and St. Paul, MN.
- Williams, J.R. 1990. Quantitative theory is soil productivity and environmental pollution. philosophical transactions: Biol. Sci. 329:421-428.
- Zadoks, J.C., T.T. Chang, and C.F. Konzak. 1974. A decimal code for the growth stages of cereals. Weed Res. 14:415-421.

# ARTICLE 2: TECHNIQUES TO IMPROVE GRAIN PROTEIN CONTENT OF HARD RED SPRING WHEAT WITH A LATE SEASON NITROGEN APPLICATION Abstract

A late-season foliar nitrogen (N) application may increase grain protein content (GPC) for hard red spring wheat (HRSW) (*Triticum aestivum* L. emend Thell.) in environments or cultivars with low protein potential. Two experiments were conducted to evaluate the effectiveness of a foliar N application through the combination of N solutions with a fungicide, urease inhibitor (N-(n-butyl) thiophosphoric triamide - NBPT), and various adjuvants using multiple droplet sizes. Urea ammonium nitrate (UAN) and urea solution (US) were applied at Zadoks growth stage (ZGS) 65 and 69. Applications resulted in greater leaf burn when combined with a fungicide, NBPT, or adjuvant. The best technique to increase GPC was through a foliar application of UAN or US at ZGS 69. The addition of a fungicide, NBPT, or adjuvants with the N solutions did not increase GPC. Therefore, the associated additional costs do not make these additions economically feasible. Spring wheat growers are advised to only consider a late-season foliar N application when environments are favorable for high yields and a profitable protein premium seems likely

#### Introduction

Spring wheat producers depend on grain yield and GPC in order to be economically successful. High grain yield and high GPC can be achieved if N availability and environmental conditions are favorable. However, attaining both high grain yield and GPC can be challenging due to environmental impacts, N availability, and cultivar selection. Protein in the grain has to reach a market threshold at point of sale, otherwise discounts are imposed leading to reduced producer profit. However, if GPC is above the market threshold a premium may be realized by the grower (Jones and Olson-Rutz, 2012). Spring wheat producers may be prompted to make inseason management decisions to increase GPC if environmental factors are positive for greater yield and N supply is perceived to be deficient. A management practice utilized to increase GPC when conditions point to reduced GPC is through a late season foliar application of N fertilizer.

Urea ammonium nitrate (28-0-0) or urea solution (ranging from 14 to 20 percent N by weight) are two possible N fertilizers that HRSW producers use to increase GPC. Increasing GPC has been successfully achieved with this additional N application between ZGS 45 and 73 with the most effective response occurring at ZGS 69 (Zadoks et al., 1974; Finney et al., 1957; Endres and Schatz, 1993). Even at the optimum timing, the efficiency of N absorption into the plant with this method application is limited (Finney et al., 1957; Alkier et al., 1972). Therefore, it may be possible to develop methods to improve the efficiency of this foliar application and could reduce costs and amount of residual N in the environment.

#### Wheat Nitrogen Use

Nitrogen is an essential nutrient in the production of chlorophyll and rubisco, which are essential for photosynthesis. Photosynthesis promotes the formation of compounds and carbohydrates needed by the plant for growth, storage, and energy (Lawlor et al., 1989). Nitrogen is also an essential component of amino acids, which are the building blocks of protein. Plant roots absorb inorganic N in the form of NO<sub>3</sub><sup>-</sup> and NH<sub>4</sub><sup>+</sup> from residual soil N, mineralization of organic matter, or fertilizer applications (Flaten and Grant, 1998). Environmental conditions, concentration of N in the soil solution, and the growth stage and growth rate of the plant can influence the amount of N uptake (Brown et al., 2005).

Grain yield and GPC are produced simultaneously during wheat development, but N is first be allocated towards grain yield before it is allocated to protein development in the kernel (Goos et al., 1982). Most of the N taken up during early vegetative growth is used in the formation of tillers, leaves, and spikes, which impact grain yield potential (Brown et al., 2005). Nitrogen is supplied to the developing kernel by remobilization from the vegetative biomass or uptake during or after heading. Nitrogen that is taken up around heading will usually influence GPC, but can only marginally influence yield because the number and size of kernels are mostly fixed at this time (Brown et al., 2005)

Environmental and agronomic factors, especially cultivar selection and nutrient and water availability, affect the rate and timing of N uptake (Fowler et al., 1990; Campbell, 1977). Therefore, an inverse relationship between grain yield and GPC result from a certain level of fertility. This can be explained by differences in N uptake and the plants' ability to utilize energy and nutrient reserves from the vegetative stage during kernel development (Brown et al., 2005). Inverse relationships between yield and grain protein has been reported in a number of crops including barley (*Hordeum vulgare* L.), oat (*Avena sativa* L.), corn (*Zea mays* L.), rice (*Oryza sativa* L.), sorghum (*Sorghum bicolor* L. Moench), and wheat (Baltensperger et al., 2008).

The proportion of N supplied to the grain is influenced by concentration of N in the vegetative tissues, cultivar grown, and environmental conditions, particularly soil moisture availability (Flaten and Grant, 1998). Yield potential is higher in favorable environments with adequate soil moisture that promotes N uptake during early plant development, but less N may be available during the grain filling period resulting in kernels with high starch content and low protein. Protein content can be high in environments with sufficient N, but inadequate moisture or environmental stress during early plant development can limit N assimilation during grain filling if the environmental conditions improve (Neidig and Snyder, 1924). High grain yield and

GPC can be obtained when N availability within an environment is greater than what is sufficient to support maximum yield potential for the seasonal environment (Bailey et al., 1992).

#### **Foliar Nitrogen Application**

A late season foliar N application has been found to increase GPC in low protein potential cultivars or environments. A potential increase of 5 to 10 g kg<sup>-1</sup> in protein has been realized with a 34 kg ha<sup>-1</sup> application of N between ZGS 45 and 73, when kernel development demands requires high levels of N (Finney et al., 1957; Schatz and Endres, 1993; Ransom et al., 2012). Studies have reported the most effective timing was directly post-anthesis (ZGS 69), with decreased effectiveness before and after this stage (Finney et al., 1957; Gooding and Davis, 1992; Schatz and Endres, 1993; Bly and Woodard, 2003). The optimum N rate for a postanthesis N application is 34 kg N ha<sup>-1</sup>, with larger rates only slightly increasing protein levels while decreasing yield (Freeman et al., 2002). Schatz (2012) observed a 10 g kg<sup>-1</sup> increase in protein across three HRSW cultivars when 34 kg ha<sup>-1</sup> of N was applied to the foliage postanthesis.

Urea ammonium nitrate or US are commonly used as N sources for foliar applications. A solution of UAN allows for N to be readily absorbed by the foliage in the form of  $NO_3^-$  and  $NH_4^+$ , as well as water-soluble organic N from urea (Christiaens et al., 2015). A US is made by dissolving dry fertilizer urea (46-0-0) in water, which can then be directly applied to the foliage using a sprayer. A US has been found to produce less burn to the foliage than UAN, but is more susceptible to ammonia volatilization from the activity of the urease enzyme present in the soil (Gooding and Davies, 1992).

Liquid forms of fertilizer N can cause leaf burning due to the salt content of the solution and a potential to form high concentrations of ammonia on the leaf tissue (Bremner, 1995). Since

US generally cause less burn than UAN and US are lower in salt and higher in potential ammonia, the greatest cause of leaf burn with these fertilizers is probably salt content. The degree of phytotoxicity depends on the growth stage, N concentration, and the cultivar (Gooding, 1988). The flag leaf in particular can be significantly affected by ammonia accumulation from a foliar application. Maintenance of the flag leaf before and during anthesis is needed for achieving high grain yield and GPC because it is the major source of photosynthates to create carbohydrates and amino acids during the grain filling period (Simpson et al., 1983). A management tactic to reduce phytotoxicity is to make the application in the cool of the day when humidity is high (Garcia and Hanway, 1976; Franzen, 2017). After anthesis, protection of the flag leaf does not appear to be as important as before or during anthesis (Schatz and Endres, 1993).

#### **Techniques to Improve Efficiency of Foliar Nitrogen Applications**

A late season foliar N application can be rapidly absorbed by the foliage, but efficiency is often low (Gooding and Davies, 1992). The foliar fertilizer is often absorbed through the leaf cuticle; however, irregularities of the leaf surface and waxes can prevent wetting of the cuticle, reducing absorption (Akin and Gray, 1984). A greenhouse study reported less than 1% of the N in the foliar application was supplied to the grain (Alkier et al., 1972). The remaining N is subject to environmental losses or washed off by precipitation and absorbed through the roots. However, N absorption through the foliage may be improved with the addition of an adjuvant or urease inhibitor, NBPT. Altman et al. (1983) reported N recovery in the grain was 44% for winter wheat when an adjuvant was added with a US applied directly to the foliage.

Adjuvant is the general term that includes surfactants, oils, and fertilizers, which vary in their chemical makeup, overall effect, and intended use (Zollinger, 2010). The purpose of an

adjuvant is to increase uptake of the active ingredient of the pesticide applied by decreasing surface tension and increasing droplet retention (Hanzen, 2000). The addition of an adjuvant to the N solution has been reported to improve uptake; however little exploration has been performed to test multiple types of adjuvants.

Surfactants help spray solutions absorb through the leaf cuticle by emulsifying, dispersing, sticking, and spreading on the leaf surface (Zollinger, 2010). The addition of a surfactant with N solutions can increase retention of the spray solution, more effectively transferring N to the grain (Altman et al., 1983; Brinck et al., 2000). Oil agents, or penetrants, are also common spray adjuvants. These include methylated seed oil (MSO) and various crop oil concentrates (COC). The oil in methylated seed oils are derived from plants, while the oil in COCs are derived from petroleum oil (Zollinger et al., 2017). A penetrant allows the spray solution to infiltrate leaf cuticles by breaking down waxy extracellular and lipid cellular barriers of the plant (Hanzen, 2000).

Droplet size can also impact pesticide absorption, retention, and deposition, which may improve the action of an N solution. Spray pressure, spray mixture, or nozzle type affects droplet size, which ranges from very fine (<145 microns) to ultra-coarse (> 650 microns) (Hofman and Solseng, 2004; Askew et al., 2013). The optimum size varies depending on intended use, volume, or liquid form being applied. Creech et al., (2016) reported that dicamba control of common lambsquarter (*Chenopodium album*) was improved using a medium droplet compared to a fine droplet. However, common sunflower (*Helianthus annuus* L.) was best controlled by dicamba with a very coarse droplet.

Another technique to improve the effectiveness of a foliar N application is through the use of a urease inhibitor in conjunction with a urea containing liquid fertilizer. A urease inhibitor

reduces the breakdown of the urea molecule by binding the active site of the urease enzyme (Deiana et al., 1999). The urease enzyme breaks the urea molecule into its component parts of NH<sub>3</sub> and CO<sub>2</sub>. The ammonia can then be lost to the environment if it is located near or at the soil surface, and especially if soil pH is greater than 7. The addition of a urease inhibitor to US has resulted in reduced N loss from ammonia volatilization (Bemner and Douglas, 1971, 1973). Brinck et al. (2000) reported improved N recovery by the crop when a urease inhibitor was added to a soil-applied US, but the urease inhibitor was not significantly beneficial when applied to foliage.

If the environment is favorable for disease development, wheat quality can also be negatively affected. A fungicide application at ZGS 65 in HRSW can prevent the graindamaging effects of fusarium head blight (*Fusarium graminearum*). Recommendations for fungicide and foliar N applications almost always urge growers to apply them separately in HRSW, even though the recommended application timing overlap. This is due to the phytotoxic effects of the liquid fertilizer on the leaf tissue, which the fungicide is intended to protect (Franzen, 2015). However, information on the effect of applying a fungicide with N solutions simultaneously is lacking. Spring wheat producers might be able to reduce costs if the fungicide and N solution could be applied together.

#### **Economic Return**

Wheat grain yield and GPC are the two most important constituents in generating an economic return in HRSW production. The standard market threshold for GPC are 140 g kg<sup>-1</sup> for HRSW and 120 g kg<sup>-1</sup> for hard red winter wheat in the upper Midwest. Price deductions are imposed if GPC falls below this market standard, while premiums may be realized if protein content is greater than the market standard. The price discounts and premiums vary depending on

the year (Bly and Woodard, 2003). For example, discounts ranged from \$22.05 Mg<sup>-1</sup> in September 2010 and \$9.18 Mg<sup>-1</sup> in September 2015 (Olson, 2015). When high protein HRSW is limited in supply, the premium for high protein may represent as much as 50% or more of the total market price of HRSW in some years (Brown et al., 2005).

A foliar N application carries an additional input cost to growers, but if the environment if favorable for high yields with insufficient N throughout the grain filling period, then an additional N application to increase GPC may be justified. The additional N application may be highly profitable if an increase in GPC reduces low-protein discounts or results in a protein premium payment greater than the cost of the application.

#### Objective

The objective of this research was to test techniques that might improve the effectiveness of increasing GPC with a foliar N application of N alone or in combination with a fungicide, a urease inhibitor, adjuvants, and using different droplet sizes.

#### **Materials and Methods**

## **General Information**

Field experiments were conducted near Casselton (46.88°N, -97.23°W), Fargo (46.93°N, -96.86°W), and Prosper (47.00°N, -97.11°W), ND in 2016 and repeated in 2017 at Ada, MN (47.35°N, -96.41°W), Casselton (46.88°N, -97.23°W) and Prosper (47.00°N, -97.11°W), ND. Soil series, taxonomy, and slope for each location are presented in Table 2.1. Soil samples were collected in the fall to determine the levels of plant-available phosphorous (P), potassium (K), and residual nitrate-N. Five random 2.5 cm core samples at a 0-30.5 and 30.5-60 cm depths were collected from the trial and combined prior to this analysis. A uniform application of dry urea (46-0-0) was applied at 75% of the recommended rate for each location and incorporated prior to

planting using a field cultivator. The North Dakota wheat nitrogen calculator was used to determine the recommended N rate (North Dakota Wheat Nitrogen Calculator, 2017). A full N rate was not applied in order to improve the probability of a protein response from a foliar N application. Table 2.2 indicates N level, previous crop, and N rate applied at each location in 2016 and 2017.

Table 2.1. Soil series, taxonomy and slope at Casselton, Fargo, and Prosper, ND, and Ada, MN in 2016-2017.

Location	Soil Series†	Soil Taxonomy‡	Slope
			%
Casselton	Bearden	Fine-silty, mixed, superactive, frigid Aeric Calciaquolls	0-2
Fargo	Fargo	Fine, smectitic, frigid Typic Epiquerts	0-2
Prosper	Bearden	Fine-silty, mixed, superactive, frigid Aeric Calciaquolls	0-2
Ada	Glyndon	Coarse-silty, mixed, superactive, frigid Aeric Calciaquolls	0-3

<sup>†</sup> Soil data obtained from (USDA-NRCS, 2016).

‡ Soil taxonomy listed on individual lines based on hyphenated soil series name.

Table 2.2. Previous crop	p with nitrogen (N	J) credit, soil N,	, P, and K test,	and N rate	applied
for locations in 2016-20	)17.				

Year	Location	Previous crop	Residual soil nitrate-N†	N credit	N rate	Р	K
				-kg ha <sup>-1</sup>		ppm	ppm
	Casselton, ND	Wheat	37	0	119	25	455
2016	Fargo, ND	Soybean‡	20	44	98	25	§
	Prosper, ND	Wheat	103	0	70	18	250
	Ada, MN	Soybean	16	50	135	11	107
2017	Casselton, ND	Wheat	83	0	95	11	368
	Prosper, ND	Wheat	108	0	78	17	370

† 2.5 cm core samples taken at 0-60 cm depth.

‡ Glycine max L.

§ Data not available.

Two experiments were designed as randomized complete blocks with four replications to

develop best practices for stimulating a protein response following a foliar N application. The

first experiment (Experiment One) consisted of twelve treatments in 2016 and fourteen treatments in 2017 and was constructed to evaluate the effects of separate and combined applications of two N solutions with a fungicide at ZGS 65 and 69 (Table 2.3). Experiment Two consisted of fifteen treatments in 2016 and eighteen treatments in 2017 and was designed to identify the effects a foliar application of N fertilizer combined with four adjuvants using two droplets sizes at ZGS 69 (Table 2.4). Additional treatments in Experiment One included a urease inhibitor. Two dilution ratios and the use of streamer bars to deliver the foliar N fertilizer in a concentrated band instead of broadcast were included in Experiment Two.

Two N solutions, UAN and a US, were used to supply N at a rate of 34 kg N ha<sup>-1</sup>. In treatments receiving foliar fertilizer, N solutions were applied at a rate of 187 l ha<sup>-1</sup>. In both experiments, UAN was applied at a dilution ratio (volume:volume) of 50:50, unless stated otherwise. The US was prepared by mixing dry urea with lukewarm water to provide a solution that was 50% urea by weight (23-0-0). For treatments requiring a urease inhibitor, (NBPT) (Agrotain Advanced, Koch Agronomic Services, LCC, Wichita, KS), was added to the US at a label recommended rate of 2.10 ml kg<sup>-1</sup> and UAN at 1.05 ml kg<sup>-1</sup>. In treatments receiving fungicide, prothioconazole and tebuconazole at 126 g and 126 g ai ha<sup>-1</sup>, respectively, were applied with a non-ionic surfactant (NIS) (Activator 90, Loveland Products, Loveland, CO) at 0.25% v/v. In Experiment Two, adjuvants included an NIS (Activator 90, Loveland Products, Loveland, CO), MSO (Super Spread MSO, San Francisco, CA), methylated seed oil organosilicone surfactant (MSOOS) (Dyne-Amic, Helena Chemical Company, Collierville, TN), and petroleum oil concentrate (POC) (Herbimax, Loveland Products, Loveland, CO) at 0.5% v/v, 1.8 l ha<sup>-1</sup>, 438 ml ha<sup>-1</sup>, and 2.3 l ha<sup>-1</sup>, respectively. Foliar applications were made in Experiment One using XR TeeJet 11002 nozzles (TeeJet Technologies, Wheaton, IL), while Experiment

Two utilized streamer bars, and XR TeeJet 11002 and TT11002 nozzles which delivered droplet sizes of 226-325 and 326-400 microns, respectively. Treatments were applied using a  $CO_2$  pressurized hand-held backpack sprayer at 207 kPa and constant speed of 3.8 km h<sup>-1</sup> with the boom height 46 cm above the crop canopy.

#### **Planting and Plot Maintenance**

Experiments One and Two were seeded side by side at a seeding rate of 2.9 million viable seeds ha<sup>-1</sup> using a no-till 3P605NT drill (Great Plains Mfg. Inc., Salina, KS) with seven rows spaced 18 cm apart. Experimental units were 1.5 m wide by 5.2 m long with 0.3 m gaps between units. Alleys between replications were cut mid-season at a width of 1.5 m, leaving the total harvested area of each experimental unit to be 1.5 m by 3.7 m. Trials at Casselton and Fargo were planted on 13 April and Prosper on 14 April in 2016. In 2017, Ada, Casselton, and Prosper were planted on 17 April, 2 May, and 15 April, respectively. A uniform seed bed was prepared before planting using a field cultivator. The HRSW cultivar, Croplan 3419, was planted across all locations. Cropland 3419 is a 2014 cultivar release of Winfield United (Arden Hills, Minnesota) with high yield potential and intermediate protein potential.

Treatment	Nitrogen Solution	Tank Mix Addition	Stage†
1)‡ Unfertilized Control			
2) Fertilized Control			
3)‡ Fungicide			ZGS 65
4) Fungicide		NIS§	ZGS 65
5) Fungicide	UAN¶	NIS	ZGS 65
6) Fungicide	UAN	NIS, NBPT	ZGS 65
7)	UAN		ZGS 65
8) Fungicide	US#	NIS	ZGS 65
9) Fungicide	US	NIS, NBPT	ZGS 65
10)	US		ZGS 65
11) Fungicide		NIS	ZGS 65
	UAN		ZGS 69
12)	UAN		ZGS 69
13) Fungicide		NIS	ZGS 65
	US		ZGS 69
14)	US		ZGS 69

Table 2.3. Treatment structure of Experiment One including tank mix combinations of fungicide, nitrogen solution, and tank mix additions at the specified timing for all locations in 2016-2017.

<sup>†</sup> Stage = Zadoks growth stage (ZGS) 65 is Anthesis, ZGS 69 is Post-Anthesis.

<sup>‡</sup> Treatments included in 2017 only.

§ Nonionic surfactant.

¶ Urea ammonium nitrate (28-0-0).

# Urea (46-0-0) solution.

All locations were scouted throughout the season for pathogens and weed presence.

Pathogen pressure was minimal in 2016 and 2017 at all locations; however, a fungicide application of prothioconazole and tebuconazole at 126 g and 126 g ai ha<sup>-1</sup>, respectively, were applied at ZGS 65 in Experiment Two to reduce the influence of disease at anthesis, due to prediction of possible fusarium head blight by the NDSU small grains disease forecasting model (NDSU small grain disease forecasting model, 2017). Broadleaf and grass weeds were controlled with an application at ZGS 14 of fenoxaprop, pyrasulfotole, bromoxynil octanoate, and bromoxynil heptanoate at 56, 18, 74, and 73 g ai ha<sup>-1</sup>, respectively.

Treatment, N Solution	Tank Mix Addition	Droplet Size	Dilution Ratio†	Adjuvant Rate
1) <sup>‡</sup> Unfertilized Control				
2) Fertilized Control				
3) UAN§		1¶	50:50	
4) UAN		2#	50:50	
5) UAN	MSO††	1	50:50	1.8 l ha <sup>-1</sup>
6) UAN	MSO	2	50:50	1.8 l ha <sup>-1</sup>
7) UAN	POC‡‡	1	50:50	2.3 l ha <sup>-1</sup>
8) UAN	POC	2	50:50	2.3 l ha <sup>-1</sup>
9) UAN	NIS§§	1	50:50	0.5 % v/v
10) UAN	NIS	2	50:50	0.5 % v/v
11) UAN	MSOOS¶¶	1	50:50	438 ml ha <sup>-1</sup>
12) UAN	MSOOS	2	50:50	438 ml ha <sup>-1</sup>
13)## UAN	Urease Inhibitor	1	50:50	1.05 ml kg <sup>-1</sup>
14)## Urea solution	Urease Inhibitor	1		2.10 ml kg <sup>-1</sup>
15) UAN		1	60:40	
16) UAN		1	75:25	
17)‡ UAN		3†††	50:50	
18)‡ UAN		3	50:50	

Table 2.4. Treatment structure of Experiment Two, including dilution ratio of nitrogen (N) solution and rate of tank mix additions for the specified droplet size for all locations in 2016-2017.

<sup>†</sup> Dilution ratio of spray solution %fertilizer:%water.

*‡* Treatments included in 2017 locations only.

§ Urea ammonium nitrate (28-0-0).

¶ 226-325 micron droplet size produced by a XR11002 nozzle.

# 236-400 micron droplet size produced by a TT1102 nozzle.

*††* Methylated seed oil.

**‡**‡ Petroleum oil doncentrate.

§§ Non-Ionic Surfactant.

¶¶Methylated seed oil organosilicone surfactant.

## Treatments not included at Fargo location in 2016.

††† Streamer bars.

#### **Data Collection and Harvest Methods**

Plots were harvested using a Wintersteiger Classic plot combine (Wintersteiger Ag, Reid,

Austria). In 2016, Fargo was harvested on 26 July, and Casselton and Prosper were harvested on

29 July. In 2017, Ada, Casselton, and Prosper were harvested on 11, 22, and 7 Aug.,

respectively. After harvest, grain samples were cleaned using a Clipper Office Tester and

Cleaner (Seedburo Equipment Co., Chicago, IL) and dried (if necessary). Grain yield, moisture,

and test weight (TW) were recorded using a GAC 2100 moisture tester (DICKEY-John Corp.,

Minneapolis, MN) and GPC was analyzed using Perten Instruments DA 7250 (Perten Instruments, Springfield, IL). Yield was adjusted and expressed at 13.5% moisture.

Field measurements in 2016 included stand counts and the percent of flag leaf phytotoxicity (leaf burn), which was visually estimated seven days after foliar applications. Leaf burn was based on symptoms across the entire plot using a percent scale of 0% (no injury) to 100% (complete leaf necrosis). In 2017, field measurements also included normalized difference vegetative index (NDVI) and normalized difference red edge (NDRE) readings using a handheld CropCircle ACS 470 (Holland Scientific, Lincoln, NE) before each foliar application and seven days post application. Times of application, average temperature, wind speed, and wind direction were recorded for each application using the nearest NDAWN weather station.

#### **Statistical Analysis**

Data were statistically analyzed using the PROC GLM procedure in SAS 9.3 (SAS Institute, Cary, NC). Random variables included year, location, and replicate, while treatments were considered fixed. Experiment One and Experiment Two were analyzed separately by location and year. Bartlett's test for homogeneity error of variance was conducted to determine if environments could be combined. Application timing and N source were additional factors analyzed in experiment one. Adjuvant and droplet size were separately analyzed as a factorial arrangement in experiment two. Both NDVI and NDRE readings were measured, and the strength of relationship between the two measurements were strong ( $r^2=0.70$ ), therefore, only NDVI readings will be discussed. Main effects and interactions were tested using the appropriate error terms. A square root transformation was applied to percent leaf burn data prior to analysis to obtain a normal distribution of the data. The control was not included in leaf burn analysis because 0 values would provide an inaccurate assessment of means separation for treatments receiving a foliar application. Means were separated using Fisher's protected least significant difference (LSD) at the 95% level of confidence ( $\alpha$ =0.05).

# **Results and Discussion**

#### **Combined Analysis**

Barltett's test for homogeneity error of variance was not significant when comparing environments for both experiments, thus allowing for combining of environments in the ANOVA. In the combined analysis, the environment by treatment interactions (E x T) for GPC, leaf burn, and NDVI measurements were significant between environments in 2016 and 2017. Therefore, GPC, leaf burn, and NDVI measurements and will be discussed separately by environment, then combined across all environments within each year. Test weight and yield did not have significant E x T interactions and were combined across all environments in 2016 and 2017.

# Experiment One: Combined and Separate Applications of Fungicide with UAN and US *Leaf Burn*

Experiment One investigated the effects of combining the applications of a fungicide with two N solutions (UAN or US) applied at ZGS 65 and 69. Across all environments, leaf burning was observed with all treatments receiving a spray solution (fungicide or N solution), but was greatest with treatments receiving an N solution. Phytotoxicity of the leaf tissue following a foliar N application can be described as scorching, burning, or tipping (Gooding and Davies, 1992). Burning was the greatest when fungicide was combined with NIS and UAN and applied at ZGS 65. Less damage was observed when either UAN or US were applied alone at the same timing (Table 2.5). The addition of NBPT to the N solution had no effect on leaf burn. Treatments containing UAN consistently caused significantly greater leaf burn than US, regardless of timing (Table 2.6). Others have also found that the severity of leaf burn varied depending on the type of N solution used (Alkier et al., 1972; Gooding et al., 1992; Bremner, 1995). Some forms of N fertilizer, such as urea has a lower salt concentration compared to UAN. Thus, desiccation of leaf cells with urea through osmosis is reduced and caused less injury than UAN (Gary, 1977).

There was a significant E x T interaction for leaf burning. The severity of burning across environments can be explained by differences in weather conditions at the time of application. In 2016, locations were planted within one day of each other and the plants matured at the same time causing some applications to take place when weather conditions were not ideal. However, in 2017 maturity varied between locations so applications could be made when weather conditions were favorable. The recommended weather conditions to reduce the chance of burning are to apply the solution in the cool of the day and when humidity is high (Garcia and Hanway, 1976). High temperatures and low humidity can lead to less moisture in the leaf tissue, resulting in lower dilution of N compounds in the concentrated solution, causing the burn (Akin and Gary, 1984; Gooding and Davies, 1992).

In 2016, leaf burn was assessed by visually determining the percentage of the flag leaf damaged, seven days after application. Burning was assessed in 2017 from NDVI values measured with a handheld CropCircle ACS-470. The CropCircle is an active optical sensor that is capable of quantifying the leaf biomass through red NDVI values (Sharma, L.K. et al., 2015). Foliage damaged by the N solution was necrotic and less green, resulting in a lower red NDVI value measured by the sensor.

								2	
		<u>2016</u>				<u>2017</u>			
			Lea	af Burn‡		NDVI			
	Application		Farg						
Treatment	Timing	Timing Cass. o Pros. Comb.						Pros.	Comb.
	%%								
1) Control, no treatment§						0.79	0.72	0.73	0.75
2) Fungicide + NIS¶	ZGS 65	18 a#	23 a	15 a	18 a	0.74	0.74	0.74	0.74
3) Fungicide + NIS + UAN††	ZGS 65	53 c	55 c	38 bcd	48 f	0.60	0.59	0.62	0.60
4) Fungicide + UAN + NBPT‡‡	ZGS 65	40 bc	43 bc	38 bcd	40 ef	0.64	0.56	0.66	0.62
5) UAN	ZGS 65	30 ab	30 ab	28 abcd	29 bcde	0.74	0.74	0.70	0.72
6) Fungicide + NIS + US§§	ZGS 65	25 ab	30 ab	20 ab	25 abcd	0.75	0.64	0.70	0.70
7) Fungicide + US + NBPT	ZGS 65	30 ab	40 bc	25 abc	32 bcde	0.73	0.65	0.70	0.69
8) US	ZGS 65	23 a	20 a	23 ab	22 ab	0.76	0.69	0.71	0.72
9) Fungicide + NIS	ZGS 65	20 a	13 ha	13 cd	35 cdof	0.73	0.65	0.60	0.60
UAN	ZGS 69	20 a	45 00	45 Cu	55 cuer	0.75	0.05	0.09	0.09
10) UAN	ZGS 69	18 a	53 c	45 d	38 def	0.73	0.55	0.69	0.66
11) Fungicide + NIS	ZGS 65	10 .	25 ah	20 sh	24 aba	0.76	0.60	0.71	0.72
US	ZGS 69	10 a	55 ab	20 ab	24 abc	0.76	0.69	0.71	0.72
12) US	ZGS 69	18 a	28 ab	23 ab	23 abc	0.75	0.65	0.70	0.70
	Mean	26	36	29	30	0.72	0.66	0.70	0.69
	LSD <sub>0.05</sub>	15	17	16	12	0.06	0.08	0.02	0.06

Table 2.5. Effects of treatment applications on leaf burn measured by visual percent leaf burn (2016) and red NDVI<sup>+</sup> (2017) for all environments and combined within years.

*†* Normalized difference vegetative index.

‡ Means separation based on transformed values of % leaf burn, which are used to calculate the LSD values.

§ Control excluded % leaf burn analysis because no leaf burn occurred.

¶ Nonionic surfactant.

# LSD values valid for comparisons within locations.

†† Urea ammonium nitrate (28-0-0).

**‡‡** N-n-butyl thiophosphoric triamide, urease inhibitor.

§§ Urea (46-0-0) solution.

The average percent leaf burn (PLB) in 2016 was 26, 36, and 29% at Casselton, Fargo,

and Prosper, respectively (Table 2.5). Across all environments, the greatest amount of burning

was observed for treatments containing fungicide + UAN and fungicide + UAN + NBPT with 48

and 40%, respectively (Table 2.5). Similar results were observed in 2017, with these treatments

giving the lowest NDVI values with 0.60 and 0.62, respectively (Table 2.5). This is compared to

an average NDVI value of 0.78 for fertilized plots shortly before application (Data not shown).

Significantly less burning was observed in treatments with US when combined with a fungicide,

as well as US alone. In 2016, treatments containing fungicide alone, fungicide + US, and US

alone resulted in average PLB of 18, 25, and 22%, respectively (Table 2.5). The NDVI values for

these treatments in 2017 had similar results with 0.74, 0.70, and 0.72, respectively (Table 2.5).

Table 2.6. Effects of treatment applications on leaf burn influenced by nitrogen source and measured by visual percent leaf burn (2016) and red NDVI<sup>†</sup> (2017) for all environments and combined within years.

	2016				2017				
	Leaf Burn‡				NDVI				
	Cass. Fargo Pros. Comb. Ada Cass. Pro							Comb.	
	%%								
Control, no treatment§					0.79 a	0.72 a	0.73 a	0.75 a	
Fungicide + NIS#	17 a¶	22 a	15 a	18 a	0.78 a	0.74 a	0.74 a	0.75 a	
UAN††	32 b	44 b	38 b	38 c	0.69 c	0.62 c	0.67 c	0.66 b	
Urea Solution <sup>‡</sup> ‡	22 a	30 a	22 a	25 b	0.75 b	0.66 b	0.70 b	0.71 a	

<sup>†</sup> Normalized difference vegetative index.

‡ Means separation based on transformed values of % leaf burn, which are used to calculate the LSD values.

§ Control excluded % leaf burn analysis because no leaf burn occurred.

¶ Nonionic Surfactant.

# LSD values valid for comparisons within locations.

†† Means derived from all treatments receiving urea ammonium nitrate (28-0-0), regardless of timing.

**‡** Means derived from all treatments receiving urea (46-0-0) solution, regardless of timing.

# Test Weight and Yield

Treatments did not differ significantly for TW and yield whether fungicide or N solutions

were combined or applied separately, and with or without NBPT. The combined average TW and

yield were 761 kg m<sup>-3</sup> and 5180 kg ha<sup>-1</sup>, respectively (Data not shown). Previous studies have

reported decreased yields when foliar N applications caused severe burning (Mullins and

Phillips, 2004). However, these results have not been consistent (Mullins and Phillips, 2004). In

this study, burning may not have been sufficient to negatively influence TW and yield.

The N source supplied from UAN or US, as well as the timing of application at ZGS 65 or 69, did not differ significantly for TW or yield. The average TW and yield for treatment applications of UAN or US were 760 and 761 kg m<sup>-3</sup>, and 5158 and 5185 kg ha<sup>-1</sup>, respectively (Data not shown). Treatments with N solutions applied at ZGS 65 or 69 (regardless of N source) had a TW of 760 and 761 kg m<sup>-3</sup> and yielded 5172 and 5170 kg ha<sup>-1</sup>, respectively.

An additional treatment in 2017 included an unfertilized control with zero N pre-plant and no foliar N application. Test weight and yield of the unfertilized control were significantly lower from all other treatments at Ada and Casselton (data not shown). However, the unfertilized control was not significant from all other treatments in Prosper. This may be due to well below normal rainfall at the beginning of the growing season (Table 2.7). The N in the fertilized plots was not effective because of lack of soil moisture and the plant roots could not effectively uptake N. Colman and Lazenby (1975) reported perennial ryegrass to have a low response to N fertilizer under low soil moisture conditions. The residual N at Prosper was high and therefore the applied N may not have affected the unfertilized plot greatly.

	Casselton‡			Prosper			F	argo	Ada	
	2016	2017	Normal	2016	2017	Normal	2016	Normal	2017	Normal
Month						-mm				
April	71	32	37	43	17	37	59	35	27	36
May	90	25	77	82	17	77	33	71	34	82
June	77	121	100	38	88	100	69	99	74	114
July	106	53	88	88	50	88	132	71	68	93
August	37	58	67	26	53	67	48	65	32	70
Total	381	289	369	277	224	369	340	341	235	395

Table 2.7. Monthly rainfall totals from planting to harvest in Casselton, Prosper, and Fargo, ND, and Ada, MN in 2016 and 2017, along with normal (1981-2010)<sup>†</sup>.

† Information collected from NDAWN, 2017.

‡ Weather information collected from Casselton Agronomy Farm, Casselton, ND.

# Grain Protein Content

The E x T interaction was significant for GPC. The average GPC in 2016 at Casselton, Prosper, and Fargo was 139, 129, and 134 g kg<sup>-1</sup>, respectively. In 2017, the average GPC was 140, 135, and 135 g kg<sup>-1</sup> at Casselton, Prosper, and Ada, respectively (Table 2.8). The major differences in GPC across environments can be explained by environmental conditions that impacted grain quality, such as temperature and rainfall. These weather conditions can impact N uptake from the soil, N absorption by the foliage, and redistribution of N within the plant, especially during grain filling (Altenbach et al., 2003; Jenner et al., 1991).

		Casselton		Prosper		Fargo	Ada	Comb.	
Treatment	Application Timing	2016	2017	2016	2017	2016	2017	2016-2017	
		g kg <sup>-1</sup>							
1) Control, No treatment		137	129	126	132	132	127	131	
2) Fungicide + NIS <sup>+</sup>	ZGS 65	138	136	130	132	131	128	132	
3) Fungicide + NIS + UAN ‡	ZGS 65	139	141	129	137	133	134	135	
4) Fungicide + UAN + NBPT§	ZGS 65	137	132	128	135	134	135	134	
5) UAN	ZGS 65	138	142	129	135	137	139	137	
6) Fungicide + NIS + US¶	ZGS 65	140	133	128	133	132	137	134	
7) Fungicide + US + NBPT	ZGS 65	139	141	127	135	132	137	135	
8) US	ZGS 65	139	139	128	135	133	137	135	
9) Fungicide + NIS	ZGS 65	142	146	122	138	127	138	130	
UAN	ZGS 69	142	140	155	150	157	150	159	
10) UAN	ZGS 69	144	148	131	139	137	138	139	
11) Fungicide + NIS	ZGS 65	140	145	131	138	133	138	138	
US§	ZGS 69	140	145	151	150	155	150	150	
12) US	ZGS 69	141	150	132	135	132	136	138	
	Range#	7	21	7	7	6	12	8	
	Mean	139	140	129	135	134	135	136	
	$LSD_{0.05}$	4	8	3	3	5	3	3	

Table 2.8. Grain protein content influenced by all treatments in experiment one for individual and combined environments.

† Non Ionic Surfactant.

<sup>‡</sup> Urea Ammonium Nitrate (28-0-0).

§ N-n-butyl thiophosphoric triamide, urease inhibitor.

¶ Urea (46-0-0) solution.

# Range = Difference between maximum and minimum values across all treatments for that location.

Significant differences in GPC were also found between treatments. The greatest increase between the control and foliar N applications occurred at Casselton in 2017 with 21 g kg<sup>-1</sup> (Table 2.8). In this study, combining a fungicide with either UAN or US did not significantly influence GPC compared the N solution alone at the same timing. The average GPC for treatments with applications at ZGS 65 were 135, 134, 137, 134, 135, and 135 g kg<sup>-1</sup> for fungicide + NIS + UAN, fungicide + NIS + UAN + NBPT, UAN alone, fungicide + NIS + US, fungicide + NIS + US + NBPT, and US alone, respectively (Table 2.8). Gooding et al. (2009) reported similar results with no impact in grain N from applications of US combined with propiconazole (250 a.i ha<sup>-1</sup>). However, GPC consistently increased with foliar N applications at ZGS 69, indicating application timing may have a greater influence on GPC.

Data indicated that applications of either UAN or US at ZGS 69 had the highest GPC across all environments. Protein levels were significantly lower in four out of the six environments when foliar N applications (regardless of N solution) occurred at ZGS 65 compared to ZGS 69. Across all environments, average GPC was 135 and 138 g kg<sup>-1</sup> when foliar N applications occurred at ZGS 65 and 69, respectively (Table 2.9). Findings in this study are contrasting to previous research that has indicated foliar N applications at ZGS 61 had the greatest influence in GPC and responses from a foliar N application decreased as the kernel developed (Finney et al., 1957; Bly and Woodard, 2003; Endres and Schatz, 1993). The N source (UAN or US), regardless of timing, did not significantly influence GPC.

Overall, a combined fungicide and N solution application will not influence GPC compared to a foliar application of N solution alone at the same timing. However, these data indicated that the timing of application may have a greater influence on the effectiveness of this
application. Therefore, foliar applications of fungicide and N solutions should be done separately

to achieve the greatest increase in GPC.

,				1			
	Casse	elton	Pros	sper	Fargo	Ada	Combined
	2016	2016 2017		2017	2016	2017	2016-2017
				g kg	g <sup>-1</sup>		
Control, No Treatment	137 a†	129 a	127 ab	132 a	132 a	127 a	131 a
Fungicide + NIS‡	138 a	130 a	130 a	132 a	131 a	130 a	133 ab
ZGS 65§	139 b	138 b	128 b	135 b	134 a	137 b	135 b
ZGS 69	142 c	147 c	132 c	138 c	134 a	137 b	138 c

Table 2.9. Grain protein content influenced by timing of application, regardless of N solution, for individual and combined environments in experiment one.

† Non Ionic Surfactant.

‡ LSD values valid for comparisons within locations.

§ Zadoks growth stage at which N solution applications occurred, regardless of N source.

## Experiment Two: Adjuvant and Droplet Size Combinations with UAN

# Leaf Burn

Experiment Two investigated the effects of foliar applications of UAN combined with different adjuvants and droplet size combinations at ZGS 69. Across all environments, foliar N applications caused leaf burning. However, the severity of burning was greater than in experiment one, with the flag leaf completely desiccated in some environments. The E x T interactions were significant for PLB and NDVI. The interactions can be explained by weather conditions at the time of application similar to experiment one, since applications occurred at the same time for both experiments.

The adjuvant x droplet interaction was not significant for leaf burning in 2016 and 2017. The droplet size can affect the proportion of spray solution in contact with the leaf surface. Chan et al. (2009) reported the coverage of the spray solution on the leaf surface increased exponentially as droplet diameter increased. Therefore, a larger droplet may increase the potential for burning if it retained. However, in this study, foliar N applications with either medium or coarse droplets did not significantly increase burning across all environments.

The average PLB in 2016 was 47, 49, and 45% at Casselton, Fargo, and Prosper, respectively (Data not shown). Different UAN and adjuvant combinations resulted in different amounts of burning across environments in 2016 and 2017. The greatest burning resulted from foliar treatments of UAN and NIS with average an PLB of 60% in 2016 and NDVI value of 0.62 in 2017 (Table 2.10). This is compared to an average NDVI value of 0.75 for fertilized plots shortly before application. Foliar applications of UAN and all adjuvants resulted in greater burning compared to UAN alone. The average PLB in 2016 was 33, 55, 57, 60, and 55% for UAN, UAN + MSO, UAN + POC, UAN + NIS, and UAN + MSOOS, respectively (Table 2.10). The NDVI values for these treatments had similar results in 2017 with 0.70, 0.65, 0.65, 0.62, and 0.63, respectively (Table 2.10). Burning has occurred in previous research with foliar applications of N solutions combined with an adjuvant. Kaiser (2017) found leaf burning to increase in corn from foliar UAN applications with the addition of an MSO adjuvant.

Overall, the addition of an adjuvant, regardless of formulation, increased burning over UAN applications alone. The addition of an adjuvant to a UAN solution most likely increases burning due to the action of the adjuvant that allows the solution to increase coverage, "stick" to the leaf surface, or dissolve the leaf cuticle. Due to these actions, the accumulation of N on the leaf surface increases resulting in leaf desiccation, especially under favorable weather conditions (Poulton et al., 1990).

		<u>20</u>	<u>16</u>		<u>2017</u>				
		Visual Le	<u>*</u> 1	A 1	<u> </u>		0 1		
	Cass.	Fargo	Pros.	Comb.	Ada	Cass.	Pros.	Comb.	
		%	,						
UAN§ Medium Droplet	38	25	33	32	0.72	0.68	0.70	0.70	
UAN Coarse Droplet	35	38	28	33	0.75	0.69	0.68	0.71	
$UAN + MSO\P Medium Droplet$	33	65	78	58	0.73	0.56	0.68	0.65	
UAN + MSO Coarse Droplet	33	55	70	53	0.69	0.58	0.67	0.65	
UAN + POC# Medium Droplet	75	66	33	58	0.69	0.56	0.70	0.65	
UAN + POC Coarse Droplet	75	63	30	56	0.70	0.55	0.68	0.65	
UAN + NIS†† Medium Droplet	68	60	65	64	0.68	0.55	0.67	0.63	
UAN + NIS Coarse Droplet	60	50	58	56	0.69	0.53	0.60	0.61	
UAN + MSOOS ** Medium Droplet	63	48	65	58	0.67	0.55	0.65	0.62	
UAN+ MSOOS Coarse Droplet	48	48	60	52	0.71	0.56	0.68	0.65	
UAN Alone	36 a	31 a	30 a	33 a	0.73	0.69	0.69	0.70	
UAN MSO	33 a	60 b	74 b	55 ab	0.71	0.57	0.67	0.65	
UAN POC	75 c	64 b	31 a	57 ab	0.70	0.56	0.69	0.65	
UAN NIS	64 bc	55 ab	61 b	60 b	0.68	0.54	0.63	0.62	
UAN MSOOS	55 ab	48 ab	63 b	55 ab	0.69	0.55	0.66	0.63	
Medium Droplet	55	53	55	54	0.70	0.58	0.68	0.65	
Coarse Droplet	50	51	49	50	0.71	0.58	0.66	0.65	
Adjuvant x Droplet LSD <sub>0.05</sub>	NS	NS	NS	NS	NS	NS	NS	NS	
Adjuvant LSD <sub>0.05</sub>	NA§§	NA	NA	NA	0.02	0.05	NS	0.05	
Droplet LSD <sub>0.05</sub>	NS	NS	NS	NS	NS	NS	NS	NS	

Table 2.10. Effects of adjuvant, droplet, and adjuvant x droplet interaction on leaf burn measured by visual percent leaf burn (2016) and red NDVI<sup>+</sup> (2017) for all environments and combined within years in experiment two.

*†* Normalized difference vegetative index.

‡ Means separation based on transformed values of % leaf burn, which were used to calculate the LSD values. LSD values valid for comparisons within locations

§ Urea ammonium nitrate (28-0-0).

¶ Methylated seed oil.

# Petroleum oil concentrate.

*†*† Nonionic surfactant.

**‡**‡ Methylated seed oil organisilicone surfactant.

§§ Means separation represented by letters in table, LSD values valid for comparisons within locations.

## Test Weight and Yield

A relationship between TW and yield has been reported in previous research, and when

yield is reduced, often a decrease in TW is also realized (Lopez-Bellido et al., 2003). The

adjuvant x droplet interaction and droplet size (regardless of adjuvant) was not significant for

TW or yield. In this study, data indicated TW and yield were not significantly different between foliar applications of UAN and the control. However, foliar applications of UAN combined with an adjuvant significantly reduced TW and yield.

The combined average TW and yield for all treatments across all environments was 762 g kg<sup>-1</sup> and 5334 kg ha<sup>-1</sup> (Data not shown). The average TW for UAN, UAN + MSO, UAN + POC, UAN + NIS, and UAN + MSSOS was 765, 760, 758, 760, and 763 g kg<sup>-1</sup>, respectively (Table 2.11). The differences in TW between UAN and adjuvant combinations can be explained by the severity of burning that occurred from these foliar applications. The burning causes plant stress, especially during the grain-filling period when these applications occur. This stress, along with the reduced photosynthesis of the flag leaf can reduce starch accumulation, thus lowering TW (Altenbach et al., 2003).

Yield decreased only marginally with foliar applications of UAN and adjuvants. A foliar application of UAN and NIS reduced yield the greatest and was significantly lower yielding than UAN alone. The yield for these treatments was 5133 and 5476 kg ha<sup>-1</sup>, respectively (Table 2.11). Foliar applications of UAN and the other three adjuvants were not significantly lower yielding than UAN alone. Yield is also negatively affected if the plant is stressed during the grain filling period. These stresses can include high temperatures, lack of water or N, and phytotoxicity. These stresses reduce the duration of grain filling, limiting starch deposition (Tewolde et al., 2006). The severe phytotoxicity stress may explain the reduction in grain yield (Figure 2.1).

	Test Weight	Yield
	kg m <sup>-3</sup>	kg ha⁻¹
UAN† Medium Droplet	766	5526
UAN Coarse Droplet	765	5426
UAN + MSO <sup>‡</sup> Medium Droplet	760	5360
UAN + MSO Coarse Droplet	761	5247
UAN + POC§ Medium Droplet	758	5326
UAN + POC Coarse Droplet	759	5182
UAN + NIS¶ Medium Droplet	761	5093
UAN + NIS Coarse Droplet	758	5173
UAN + MSOOS# Medium Droplet	764	5256
UAN+ MSOOS Coarse Droplet	761	5310
UAN	765	5476
UAN MSO	760	5304
UAN POC	758	5254
UAN NIS	760	5133
UAN MSOOS	763	5283
Medium Droplet	762	5312
Coarse Droplet	761	5268
Adjuvant x Droplet LSD <sub>0.05</sub>	NS	NS
Adjuvant LSD <sub>0.05</sub>	4	165
Droplet LSD <sub>0.05</sub>	NS	NS

Table 2.11. Average test weight and yield for adjuvant, droplet, and adjuvant x droplet interactions combined across environments in 2016 and 2017 in experiment two.

† Urea Ammonium Nitrate (28-0-0).

‡ Methylated seed oil.

§ Petroleum oil concentrate.

¶ Nonionic surfactant.

# Methylated seed oil organisilicone surfactant.



Figure 2.1. Effect of leaf burn on yield in 2016 (left) and 2017 (right) caused by foliar N applications. Leaf burn measured by visual % leaf burn in 2016 and normalized difference vegetative index in 2017.

### **Grain Protein Content**

There were significant E x T interactions for GPC. In 2016, the average GPC for Casselton, Prosper, and Fargo was 139, 132, and 133 g kg<sup>-1</sup>, respectively, and in 2017 was 136, 138, and 135 g kg<sup>-1</sup> for Casselton, Prosper, and Ada (data not shown). Differences in GPC across environments may be due to weather conditions at the time of application and immediately following the foliar N application. High temperatures and low moisture can affect the efficiency of this application (Terman, 1979). The average temperature during the grain filling period was normal, with minimal variation between locations (data not shown). However, in 2016, average precipitation during the same time was normal, but was below normal in 2017 (Table 2.7). Environments with no response to the late season foliar N application may be due to sufficient N in the soil profile above what is needed to produce yield and GPC under the conditions in that particular year.

Significant differences in GPC were found between treatments in four out of the six environments in 2016 and 2017. An average increase of 8 g kg<sup>-1</sup> over the control occurred across all environments (Table 2.12). The adjuvant x droplet interaction was not significant for GPC.

Mercer (2007) suggested that decreasing droplet size would lead to an increase in the uptake of the active ingredient and therefore increase grain N. Findings in this study indicated GPC was only marginally different between droplet sizes and was not significant across all environments.

Different UAN and adjuvant combinations (regardless of droplet size) resulted in different GPC, but was not significantly different compared to UAN alone in five out of the six environments. The greatest increase in GPC occurred with a foliar application of UAN and POC at Prosper in 2016 with 136 g kg<sup>-1</sup> compared to UAN alone with 133 g kg<sup>-1</sup> (Table 2.12). However, this was not consistent across environments. Grain protein content may not have been increased with the addition of an adjuvant due to phytotoxicity caused by the N solution. After foliar N applications, the solution is either absorbed through the leaf stomata or run off leaf surfaces and absorbed by the roots and redistributed to the grain. The function of the adjuvants are to increase retention of the droplets, improve the interface between the leaf surface and N compounds, or increase penetration of the N compounds. However, the rate or amount of uptake of the N compounds may have been so great that it resulted in localized cell death. The consequence of this resulted in the inability of the N to translocate out of the leaf, reducing the effectiveness of the applications (Brian, 1972; Merritt, 1982; Knoche et al., 1992; Forster et al., 1997).

Overall, combined data confirmed that a late season foliar UAN application at ZGS 69 can reliably increase GPC. Findings in this study suggest the addition of adjuvants used in this experiment (regardless of formulation) will not increase GPC greater than an application of UAN alone. Also, droplet size will not have an impact on the efficiency of this late season application. This is contrasting to previous research that has reported an increase in GPC while using smaller droplet sizes with an adjuvant and UAN (Wyatt, 2013). Different responses in GPC to various

adjuvants added to late season foliar N applications across environments suggests that additional research needs to be conducted to help determine favorable environmental conditions to use an adjuvant with UAN.

<b>t</b>	Casse	elton	Pros	sper	Fargo	Ada	Comb.
	2016	2017	2016	2017	2016	2017	2016-2017
UAN† Medium Droplet	140	139	133	139	136	136	137
UAN Coarse Droplet	138	138	133	141	130	134	136
UAN + MSO <sup>+</sup> , Medium Droplet	140	138	130	138	132	134	136
UAN + MSO Coarse Droplet	140	136	132	138	134	133	136
UAN + POC§ Medium Droplet	140	136	135	138	130	135	135
UAN + POC Coarse Droplet	138	142	136	138	132	134	137
$UAN + NIS \P Medium Droplet$	138	135	130	140	134	134	135
UAN + NIS Coarse Droplet	139	137	132	138	134	135	136
UAN + MSOOS# Medium Droplet	138	132	132	139	132	136	135
UAN+ MSOOS Coarse Droplet	138	138	133	138	133	136	136
UAN Alone	139	138	133	140	133	135	136
UAN MSO	140	137	131	138	133	134	136
UAN POC	139	139	136	138	131	134	136
UAN NIS	139	136	131	139	134	135	135
UAN MSOOS	138	135	133	138	133	136	135
Medium Droplet	139	136	132	139	133	135	136
Coarse Droplet	139	138	133	139	132	134	136
Adjuvant x Droplet LSD <sub>0.05</sub>	NS	NS	NS	NS	NS	NS	NS
Adjuvant LSD <sub>0.05</sub>	NS	NS	4	NS	NS	NS	NS
Droplet LSD <sub>0.05</sub>	NS	NS	NS	NS	NS	NS	NS

Table 2.12. Grain protein content influenced by adjuvant, droplet, and adjuvant x droplet interactions for individual and combined environments in experiment two.

† Urea Ammonium Nitrate (28-0-0).

‡ Methylated seed oil.

§ Petroleum oil concentrate.

¶ Nonionic surfactant.

# Methylated seed oil organisilicone surfactant.

# Additional Treatments

In 2016 and 2017, additional foliar N applications included dilution ratios (UAN:Water)

of 60:40 and 75:25, as well as NBPT combined with UAN and US. These treatments showed no

effect on leaf burn, TW, yield, or GPC, with the exception of UAN and NBPT. Foliar applications of UAN and NBPT significantly reduced TW compared to UAN alone and the control (data not shown). Similar GPC was achieved with UAN:Water dilution ratio of 50:50, 60:40, and 75:25. This suggests that a single volume of the N solution can cover a larger area, and the same N rate can be applied using less total volume, ultimately saving costs for a producer. Foliar applications of UAN and US with NBPT were not significantly different from all treatments receiving a foliar N application.

Experiment Two included an unfertilized control in 2017, similar to experiment one. Parallel results were obtained in experiment two and TW and yield were significantly lower compared to fertilized treatments at Ada and Casselton, but no significant difference was found at Prosper. This can be explained for similar reasons described in experiment one with greater levels of residual N and low soil moisture at the beginning of the growing season in Prosper.

Additional treatments in 2017 included foliar applications of UAN at ZGS 65 and 69 using streamer bars rather than broadcast nozzles. A streamer bar is adapted to a spray nozzle and delivers a uniform stream of the N solution. This is contrasting to broadcast nozzles that deliver the N solution in a pattern to the foliage. A benefit of streamer bars is that foliar applications can be made in windy conditions and less contact of the N solution with the foliage, unlike broadcast nozzles (Arnall et al., 2009). In this study, the severity of leaf burning was significantly decreased when UAN was applied with streamer bars compared to UAN applications with broadcast nozzles. A single stream applies the spray solution allowing less contact to be made with the foliage, thus causing less burning. Test weight and yield were not significantly different between foliar applications of UAN with streamer bars and UAN applications using broadcast nozzles (data not shown). Streamer bars may be an effective method

to apply UAN in environments with sufficient soil moisture and adequate rainfall following application. However, without sufficient moisture or rainfall to move N into the soil solution, the plant roots cannot absorb and redistribute N to the grain.

### Conclusion

The severity of leaf burn from a late-season foliar N application can vary depending on the weather conditions at the time of application, as well as the N solution. An application of UAN will often increase leaf burning over US. This has been reported in previous research due to the increased toxicity of ammonia in UAN. Also, combining a fungicide with either UAN or US will increase leaf burning compared to applications done separately. The addition of an adjuvant (regardless of formulation) will most likely cause severe leaf burning when combined with UAN due to increased coverage and absorption of the N compounds. In this study, droplet size did not impact leaf burning, TW, yield, or GPC.

Test weight and grain yield are often significantly related. Previous research has indicated foliar N applications to have variable effects on grain yield. In this study, a foliar application at ZGS 65 of N solution combined with a fungicide to have no substantial effect on TW or yield. However, the addition of an adjuvant, regardless of formulation, reduced TW and yield compared to UAN alone. This is likely due to an increase in leaf burn realized with the additional adjuvant. Damage to the foliage can decrease TW and yield due to reduced starch accumulation during grain filling.

A late-season foliar N application significantly increased GPC. In this study, an increase between 6 to 21 g kg<sup>-1</sup> was observed between treatments receiving supplemental N and the check for both experiments. The combination of a fungicide with N solution will not have a negative impact on GPC, but rather the timing of this application will more effectively influence GPC.

The greatest increase was realized with an application at ZGS 69, with N source from either UAN or US. The addition of an adjuvant or NBPT will not significantly increase GPC beyond that of UAN alone, therefore, the additional cost of these inputs would not be recommended for producers.

Overall, a benefit from a late season foliar N application can be realized due to an increase in GPC, especially in years that weather conditions favor N uptake and redistribution of N to the grain. However, no benefit in increasing GPC is realized with the use of an adjuvant or NBPT and the additional cost would not be worth the expense for the producer. The additional treatments such as increased dilution ratios and the use of streamer bars to apply the N solution may have some benefits to producers and future research should investigate the effectiveness of these further.

#### References

- Akin, G.W., and R.C. Gray. 1984. Foliar fertilization. *In*: Hauck, R.D. (ed.) Nitrogen in crop production. Am. Soc. Agron. Madison, WI. p. 579-584.
- Alkier, A.C., G.J. Racz, and R.J. Soper. 1972. Effects of foliar- and soil-applied nitrogen and soil nitrate-nitrogen level on the protein content of Neepawa wheat. Can. J. Soil Sci. 52:301-309.
- Altenbach, S.B., R. Chan, F.M. DuPont, E.L. Johnson, K.M. Kothari, and D. Lieu. 2003. Temperature, water and fertilizer influence on the timing of key events during grain development in US spring wheat. J. Cereal Sci. 37:9-20.
- Altman, D.W., W.E. Kronstad, and W.L. McCuistion. 1983. Grain protein percentage, kernel hardness, and grain yield of winter wheat with foliar applied urea. Agron. J. 75: 87-91.

- Arnall, B., J. Edwards, and H. Zhang. Methods for applying topdress nitrogen to wheat. Ext. Publ. PSS-2261. Oklahoma Coop. Ext. Serv., Stillwater, OK.
- Askew, S.D., R. Grisso, L. Hipkins, P. Hipkins, and D. Mccall. 2013. Nozzles: selections and sizing. Ext. Publ. 442-032. Virginia Coop Ext., Blacksburg, VA.
- Bailey, L.D., L.E. Gauer, D.T. Gehl, and C.A. Grant. 1992. Effects of nitrogen fertilization on grain protein content, nitrogen uptake, and nitrogen use efficiency of six spring wheat (*Triticum aestivum* L.) cultivars, in relation to estimated moisture supply. Can. J. Plant Sci. 72:235-241.
- Baltensperger, D., J. Blumenthal, K.G. Cassman, S. Mason, and A. Pavlista. 2008. Importance and effect of nitrogen on crop quality and health. Misc. Publ. University of Nebraska-Lincoln, Lincoln, NE.
- Bly, A.G., and H.J. Woodard. 2003. Foliar nitrogen application timing influence on grain yield and protein concentration of hard red winter and spring wheat. Agron. J. 95:335-338.
- Bremner, J.M. 1995. Recent research on problems in the use of urea as nitrogen fertilizer. Fert. Res. 42:321-329.
- Bremner, J.M., and L.A. Douglas. 1973. Effects of some urease inhibitors on urea hydrolysis in soils. Proc. Soil Sci. Soc. Am. 32:225-226.
- Bremner, J.M., and L.A. Douglas. 1971. Inhibition of urease activity in soils. Soil Biol. Biochem. 3:297:301.
- Brian, R.C. 1972. Uptake and movement of paraquat in cocksfoot and wheat as influenced by surfactants. Pestic. Sci. 3:121-132.

- Brinck, J., L. Grant, and F. Tiberg. 2000. Adsorption and surface-induced self-assembly of surfactants at the solid-aqueous interface. Curr. Opin. in Colloid and Interface Sci. 4:411-419.
- Brown, B., N. Christensen, B. Pan, J. Stark, and M. Westcott. 2005. Nitrogen management for hard wheat protein enhancement. Ext. Publ. PNW-578. Pacific Northwest Ext. Publ., Moscow, ID.
- Campbell, C.A., H.R. Davidson, and F.G. Warder. 1977. Effects of fertilizer N and soil moisture on yield, yield components, protein content and N accumulation in the aboveground parts of spring wheat. Can. J. Soil Sci. 57:311-327.
- Chan, K.C., J. Frantz, M.E. Reding, Y. Yu, and H. Zhu. 2009. Evaporation and coverage area of pesticide droplets on hairy and waxy leaves. Biosystems Engineering 104:324-334.
- Christiaens, R., A. Pandey, and O. Walsh. 2015. Liquid N fertilizer evaluation in spring wheat.*In*: Davenport, J., M. Galen, and J. Walworth (eds.) Proceedings of the Western NutrientManagement Conference, Reno, NV. 5-6 March 2015. Reno, NV. 11:181-187.
- Colman, R.L., and A. Lazemby. 1975. Effect of moisture on growth and nitrogen response by *Lolium perenne* L. Plant Soil. 42:1-13.
- Creech, C.F., R.S. Henry, J.D. Luck, and J.G. Moraes. 2016. The impact of spray droplet size on the efficacy of 2, 4-D, Atrazine, Chlorimuron-Methyl, Dicamba, Glufosinate, and Saflufenacil. Weed Technol. 3:573-586.
- Deiana, S., C. Gessa, B. Manunza, and M. Pintore. 1999. The binding mechanism of urea, hydroxamic acid and N-(N-butyl)-phosphoric triamide to the urease active site. A comparative molecular dynamics study. Soil Biol. Biochem. 31:789–796.

- Endres, G., and B. Schatz. 1993. Foliar N applied post-anthesis to enhance wheat grain protein. Mis. Publ. North Dakota State Univ., Carrington Res. Ext. Center., Carrington, ND.
- Forster, W.A., J.A. Zabkiewicz, R.J. Murray, and S.M. Zedaker. 1997. Contact phytotoxicity of triclopyr formulations on three plant species in relation to their uptake and translocation. *In*: Proceedings 50<sup>th</sup> New Zealand Plant Protection Conference, Lincoln, New Zealand 125-128.
- Finney, K.F., H.C. Freyer, W. Meyer, and F.W. Smith. 1957. Effect of foliar spraying of pawnee wheat with urea solution on yield, protein content, and protein quality. Agron. J. 49:341-347.
- Flaten, D., and C. Grant. 1998. Fertilizing for protein content in wheat. *In*: D.B. Fowler, W.E.
  Geddes. A.M. Johnston, and K.R. Preston (eds.) Wheat production and marketing: Proc of the wheat protein symposium. Saskatoon, Saskatchewan, Canada. Printcrafters Inc., Winnipeg. MB. p. 151-168
- Franzen, D. 2017. Protein Enhancement Recipe. Crop and Pest Report for June 22, 2017. Misc.Publ. North Dakota State Univ. Fargo, ND.
- Franzen, D. 2015. Nitrogen application for spring wheat and durum protein increase for June 18, 2015. Misc. Publ. North Dakota State Univ. Fargo, ND.
- Franzen, D. 2010. North Dakota fertilizer recommendation tables and equations. Ext. Publ. SF-882:1-16. North Dakota State Univ. Ext. Serv., Fargo, ND.
- Freeman, K.W., G.V. Johnson, R.W. Mullen, W.R. Raun, W.E. Thompson, C.W. Woolfolk, and K.J. Wynn. 2002. Influence of late-season foliar nitrogen applications on yield and grain nitrogen in wheat. Agron. J. 94:429-434.

- Fowler, D.B., J. Brydon, B.A. Darroch, M.H. Entz, and A.M. Johnston. 1990. Environment and genotype influence on grain protein concentration of wheat and rye. Agron. J. 82:664-666.
- Garcia, L.R., and J.J. Hanway. 1976. Foliar fertilization of soybeans during the seed-filling period. Agron. J. 68:653-657.
- Garrido-Lestache, E., L. Lopez-Billido, and R.J. Lopez-Bellido. Durum wheat quality under Mediterranean conditions as affected by N rate, timing and splitting, N form and S fertilization. 2005. E. J. of Agron. 23:265-278.
- Gary, R.C. 1977. Foliar fertilization with primary nutrients during the reproductive stage of plant growth. Proc. Fert. Soc. London, England. no. 164.
- Gooding, M.J. 1988. Interactions between late-season foliar applications of urea and fungicide on foliar disease, yield and breadmaking quality of winter wheat. PhD thesis. Harper Adams Agric Coll., Salop, UK.
- Gooding, M.J., and W.P. Davies. 1992. Foliar fertilization of cereals: a review. Fert. Res. 32:209-222.
- Goos, R.J., D.G. Westfall, A.E. Ludwick, and J.E. Goris. 1982. Grain protein concentration as an indicator of nitrogen sufficiency for winter wheat. Agron. J. 75:130-133.
- Hanzen, J.L. 2000. Symposium adjuvants-terminology, classification, and chemistry 1. Weed Technol. 14:773-784.
- Hofman, V.L., and E.G. Solseng. 2004. Spray Equipment and Calibration. Ext. Publ. AE73. North Dakota State Univ. Fargo, ND.

- Hurd, C., J.M. Randall, and M. Tu. 2001. Weed Controls Methods Handbook, The Nature Conservancy (online). Available at: https://www.invasive.org/gist/products/ handbook/methods-handbook.pdf (accessed 19 Oct. 2017).
- Jenner, C.F., T.D. Ugalde, and D. Aspinall. 1991. The physiology of starch and protein deposition in the endosperm of wheat. Aust. J. Plant Physiol. 18:221-226.
- Jones, C., and K. Olson-Rutz. 2012. Practices to increase grain protein. Misc. Publ. Montana State Univ., Bozeman, MT.
- Kaiser, D. 2017. Your guide to foliar nutrient applications for Sept. 5, 2017. Misc. Publ. Univ. of Minnesota, St. Paul, MN.
- Knoche, M., G. Noga, and F. Lenz. 1992. Surfactant induced phytotoxicity: evidence for interaction with epicuticular wax fine structure. Crop Prot. 11:51-56.
- Lawlor, D.W., M. Kontturi, and A.T. Young. 1989. Photosynthesis by flag leaves of wheat in relation to protein, Ribulose Bisphosphate Carboxylase activity and nitrogen supply. J. Exp. Bot. 40:43-52.
- Mercer, G.N. 2007. A simple diffusion model of the effect of droplet size and spread area on foliar uptake of hydrophilic compounds. Pestic. Biochem. and Physiol. 88:128-133.
- Merrit, C.R. 1982. The influence of form of deposit on the phytoxicity of difenzoquat applied as individual drops to Avena fauta. Annuals of applied biology. 101:517-525.
- Mullins, G.L., and S.B. Phillips. 2004. Foliar burn and wheat grain yield responses following topdress-applied nitrogen and sulfur fertilizers. J. of Plant Nutr. 27:921-930.
- Neidig, R.E., and R.S Snyder. 1924. The relation of moisture and available nitrogen to the yield and protein content of wheat. Soil Sci. 18:173–179.

- North Dakota Agricultural Weather Network. 2017. Available at https://ndawn.ndsu.nodak.edu/ weather-data-daily.html (accessed 19 Oct. 2017). Fargo, ND.
- North Dakota Wheat Nitrogen Calculator. 2017. Available at https://www.ndsu.edu/pubweb/ soils/wheat/ (accessed 2 Nov. 2017). Fargo, ND.
- NDSU Small Grains Disease Forecasting Model. 2017. Available at https://www.ag.ndsu.edu/ cropdisease/ (accessed 2 Nov. 2017). Fargo, ND.
- Olson, F. 2015. Crop Outlook: Now what?! (online). Available at: https://www.ag.ndsu.edu/ cropeconomics/documents/CropUpdateMhd-7-09-10.pdf. (accessed: 15 Oct. 2017).
- Poulton, P.R., L.V. Vaidyanathan, D.S. Powlson, and D.S. Jenkinson. 1990. Evaluation of the benefit of substituting foliar urea for soil-applied nitrogen for winter wheat. Aspects of Appl. Biol. 25:301-308.
- Ransom, J.K., M. Mergoum, S. Simsek, M. Aceveda, T. Friesen, M. McMullen, S. Zhong, R.
  Olson, E. Eriksmoen, B. Hanson, G. Martin, G. Bradbury, and B. Schatz. 2012. North
  Dakota hard red spring wheat variety trial results for 2012 and selection guide. Ext. Publ.
  A-574-12. North Dakota State Univ. Ext. Serv. Fargo, ND.
- Rehm, G., and D. Franzen. 2005. Fertility management of wheat. P. 60-64. *In*: J.J. Wiersma and J.K. Ransom (eds.) The small grains field guide. North Dakota State Univ. and the Univ. of Minnesota. Fargo, ND, and St. Paul, MN.
- Schatz, B.G. 2012. Influence of 'late' post-anthesis nitrogen on spring wheat performance (online). Available at: https://www.ag.ndsu.edu/carringtonrec/documents/agronomyrd/ docs2012/2012-influence-of-a-late-post-anthesis-nitrogen-on-spring-wheat-performance. (accessed 19 Oct. 2017). Fargo, ND.

- Sharma, L.K., H. Bu, A. Denton, and D.W. Franzen. 2015. Active-optical sensors using red NDVI compared to red edge NDVI for prediction of corn grain yield in North Dakota. Sensors 15:27832-27843.
- Simpson, R.J., H. Lambers, and M.J. Dalling. 1983. Nitrogen redistribution during grain growth in wheat (*Triticum aestivum* L.). Plant Physiol. 71:7-14.
- Terman, G.L. 1979. Yields and protein concentration of wheat grain as affected by cultivar, N, and environmental growth factors. Agron. J. 71:437-440.
- Tewolde, H., C.H. Fernandez, and C.A. Erickson. 2006. Wheat cultivars adapted to post-heading high temperature stress. J. Agron. Crop Sci. 192:111-120.
- USDA-Natural Resources Conservation Service. 2016. Available at http://websoilsurvey.nrcs. usda.gov/ app/HomePage.htm (accessed 19 Oct. 2017). USDA-NRCE, Washington, DC.
- Wyatt, E. 2013. Effect of droplet size and nitrogen rate on protein content of hard red winter wheat (*Triticum Aestivum* L.). Masters Thesis. Oklahoma State Univ. Stillwater, OK.
- Zadoks, J.C., T.T. Chang, and C.F. Konzak. 1974. A decimal code for the growth stages of cereals. Weed Res. 14:415-421
- Zollinger, R. 2010. Ponderings, Pontifications, and Prognostications on Adjuvants. *In*: Proceedings of the Crop Pest Management Shortcourse. St. Paul, MN. 8 Dec. 2010.
- Zollinger, R., M. Christoffers, C. Dalley, G. Endres, G. Gramig, K. Howatt, B. Jenks, C. Keene,
  R. Lym, M. Ostlie, T. Peters, A. Robinson, A. Thostenson, and H. Valenti. 2017. North
  Dakota Weed Control Guide for 2017. Ext. Publ. W-253. North Dakota State Univ. Ext.
  Serv. Fargo, ND.

#### CONCLUSION

Grain quality for HRSW producers can be difficult to consistently predict within and across years. The DSSAT can be a valuable tool to accurately predict HRSW growth by updating the CSM with measured weather data during the growing season, while supplementing historic weather data to forecast weather for the remainder of the growing season. In this study, the best approach to forecast weather was through a style that uses daily weather over multiple years to give a range of possible outcomes (distribution). However, plant growth should not be predicted with forecasted weather before ZGS 45. The accuracy of simulations improves as plant development progresses to ZGS 61. Variability in simulation accuracy existed between locations, therefore improved physical soil and cultivar measurements would be needed to improve the accuracy of predictions. However, overall DSSAT was determined to be an effective tool to predict HRSW yield during the growing season. Since yield was predicted with adequate accuracy, DSSAT could be used as a decision support system to allow producers to make informed management decisions after ZGS 45. A decision producers may determine based on grain yield is whether or not a late season foliar N application would be needed to increase GPC.

A late season foliar N application might be considered to optimize GPC, if anticipated grain yields are likely to exceed the yield considered when pre-plant recommendations were determined (Wuest and Cassman, 1992). In this study, a foliar N application at either ZGS 65 or 69 significantly increased GPC, with the greatest increase occurring with applications at ZGS 69 using either UAN or US. An N solution combined with a fungicide and applied at ZGS 65 will not decrease GPC. However, the timing of this application indicated that a more effective increase in GPC can be realized if the applications are done separately. The addition of different adjuvants or NBPT did not improve the effectiveness of a UAN application to increase GPC.

However, phytotoxicity from the N solution was severely increased with these additions. The severity of leaf burn caused by foliar N applications also depended on the weather conditions at the time of application. Depending on the severity of burning, TW and yield were significantly decreased.

The cost of a late season foliar N application must be equal to or less than the economic return from a premium or reduced discount in GPC from the market threshold of 140 g kg<sup>-1</sup>. Implementing DSSAT can be a management tool for producers to accurately predict grain yield from the ZGS 45 if predicted yields are sufficient for an economic return to be realized. This would allow little time for a management decision to be made, since the optimum timing to increase GPC from this application occurs at ZGS 69. The techniques used in this study suggest the additional cost of an adjuvant or NBPT would not increase GPC high enough to realize an economic return.

## APPENDIX

	inite of				022			000 =0	10.			
	Carri	ngton	Hettinger		Lang	gdon	Mi	not	Will	iston	Com	bined
	Sim.‡	Obs.§	Sim.	Obs.	Sim.	Obs.	Sim.	Obs.	Sim.	Obs.	Sim.	Obs.
Year						-DAP						
2005	73	71	71	65	67	52	70	64	73	67	71	64
2006	69	66	68	61	67	55	67	61	67	58	68	60
2007	69	62	67	62	71	72	67	61	69	63	69	64
2008	79	80	73	69	75	76	72	68	75	70	75	73
2009	69	67	72	68	69	56	71	66	74	71	71	66
2010	69	68	78	78	74	79	69	59	71	70	72	71
2011	69	63	72	68	65	52	68	61	66	59	68	61
2012	73	69	82	81	72	72	72	68	69	82	74	74
2013	63	56	74	70	64	50	63	57	66	58	66	58
2014	69	63	74	74	72	54	69	57	66	60	70	62
2015	77	76	79	79	72	70	71	68	69	66	74	72
2016	73	72	79	78	70	62	68	61	65	59	71	66

Table A.1. Simulated anthesis date using full season weather data and anthesis date determined by the NDAWN wheat GDD calculator† in 2005-2016.

<sup>†</sup> North Dakota Agricultural Weather Network wheat Growing Degree Day calculator (NDAWN, 2017b).

‡ Simulated anthesis date from model output.

§ Observed anthesis date determined by the NDAWN wheat GDD calculator.

¶ Days after planting.

	Carri	arrington Hettinger		Lan	Langdon		Minot		ston	Combined		
	Sim.‡	Obs.§	Sim.	Obs.	Sim.	Obs.	Sim.	Obs.	Sim.	Obs.	Sim.	Obs.
Year						kg ha <sup>-1</sup>						
2005	4128	3945	4438	2623	5551	4052	5066	5044	4813	4830	4799	4099
2006	4281	4104	2536	2408	1744	3904	2426	2170	2109	2358	2594	3014
2007	4396	3922	3425	2650	5250	5631	5237	5367	4275	3687	4362	4406
2008	4968	4708	3563	2585	4860	5631	4531	4484	2841	2548	3957	4187
2009	3432	4032	3967	2246	6041	5955	4064	5203	2698	3582	3696	4548
2010	4456	5341	4807	4350	5087	5373	4462	4754	3660	2892	4403	4633
2011	4250	2746	4545	4486	3949	5160	4418	3014	2855	2715	3992	3636
2012	4318	3778	4078	4713	4854	4792	4301	3330	3581	2235	4353	3643
2013	2032	2014	4779	4258	3647	5105	4685	3635	4682	3765	3861	3860
2014	4926	5564	4797	5392	3859	5437	5124	4225	2408	2340	4342	4473
2015	3580	4207	5373	4491	4565	5408	5162	4757	2922	2783	4144	4506
2016	3506	3975	2507	3308	5716	4540	4761	4720	4267	4240	4312	3996

Table A.2. Simulated grain yield using full season weather data and observed grain yield obtained from North Dakota REC<sup>†</sup> cultivar trials in 2005-2016.

† Research extension center.

Simulated grain yield from model output.§ Observed yield from North Dakota Research Extension Center cultivar trials.

		Experim	ent One	Experin	nent Two
Location, Year		ZGS† 65	ZGS 69	ZGS 65	ZGS 69
	Date‡	18 June 2016	21 June 2016	NA††	18 June 2016
Fargo,	Time§	9:00 am	2:15 PM		10:00 am
2016	Temp.¶	18	28		18
	Wind#	6 NW	14-16 N		6 NW
	Date	28 June 2016	18 June 2016	NA	28 June 2016
Prosper,	Time	11:00 am	11:45 am		12:00 PM
2016	Temp.	21	27		21
	Wind	14 N-NW	3 NW		17 N-NW
	Date	28 June 2016	18 June 2016	NA	28 June 2016
Casselton,	Time	12:30 PM	1:00 PM		1:30 PM
2016	Temp.	22	28		22
	Wind	6-8 N-NW	16-18 NW		6-8 N-NW
	Date	3 July 2017	26 June 2017	26 June 2017	3 July 2017
Ada,	Time	7:00 am	7:30 am	7:30 am	8:30 am
2017	Temp.	20	13	13	20
	Wind	6-10 S	5-8 NW	5-8 NW	6-10 S
	Date	30 June 2017	26 June 2017	26 June 2017	30 June 2017
Prosper,	Time	7:00 am	2:00 PM	1:00 PM	8:00 am
2017	Temp.	18	23	23	18
	Wind	3-13 NW	6-14 NW	6-14 NW	3-13 NW
	Date	5 July 2017	5 July 2017	28 June 2017	10 July 2017
Casselton,	Time	8:45 am	7:00 am	8:30 am	7:40 am
2017	Temp.	24	23	18	18
	Wind	6-11 E-SE	5-10 E-SE	8-10 S	2-5 E

Table A.3. Weather conditions at the time of treatment applications for all environments in experiment one and two in 2016 and 2017.

† Zadoks growth stage.

<sup>‡</sup> Date of treatment application.

§ Time of application.

¶ Air temperature (°C).

# Wind speed and direction. Wind speed measured in km  $h^{-1}$ .

†† Not applicable, no treatment applications at ZGS 65 in 2016

Treatment	Application Timing	Ada	Cass	Pros	Combined
1) Control, no treatment		0.20	0.25	0.31	0.25
2) Fungicide + NIS <sup>†</sup>	ZGS 65	0.33	0.29	0.33	0.32
3) Fungicide + NIS + UAN‡	ZGS 65	0.33	0.30	0.32	0.32
4) Fungicide + UAN + NBPT§	ZGS 65	0.32	0.31	0.33	0.32
5) UAN	ZGS 65	0.30	0.28	0.29	0.29
6) Fungicide + NIS + US¶	ZGS 65	0.31	0.29	0.29	0.30
7) Fungicide + US + NBPT	ZGS 65	0.24	0.26	0.28	0.26
8) US	ZGS 65	0.31	0.27	0.31	0.30
9) Fungicide + NIS	ZGS 65	0.30	0.33	0.31	0.31
UAN	ZGS 69	0.31	0.30	0.32	0.31
10) UAN	ZGS 69	0.29	0.36	0.29	0.31
11) Fungicide + NIS	ZGS 65	0.30	0.28	0.29	0.29
US	ZGS 69	0.25	0.23	0.29	0.26
12) US	ZGS 69	0.30	0.28	0.30	0.29
	Mean	0.29	0.29	0.3	0.29
	LSD <sub>0.05</sub>	0.02	NS	0.01	0.04

Table A.4. Effects of treatment applications on leaf burn measured by normalized difference red edge (NDRE) in 2017 for individual and combined environments in Experiment One.

† Nonionic surfactant.

<sup>‡</sup> Urea ammonium nitrate (28-0-0).

§ N-n-butyl thiophosphoric triamide, urease inhibitor.

¶ Urea (46-0-0) solution.

Timing†	Treatment	TW	Yield
		kg m <sup>-3</sup>	kg ha <sup>-1</sup>
	1) Control, No treatment	763	5153
ZGS 65	2) Fungicide + NIS <sup>‡</sup>	764	5295
ZGS 65	3) Fungicide + NIS + UAN	762	5184
ZGS 65	4) Fungicide + UAN + NBPT#	760	5140
ZGS 65	5) UAN	760	5148
ZGS 65	6) Fungicide + NIS + US	762	5221
ZGS 65	7) Fungicide + US + NBPT	761	5233
ZGS 65	8) US	760	5095
ZGS 65 ZGS 69	9) Fungicide + NIS	759	5179
ZGS 69	10) UAN	755	5139
ZGS 65	11) Fungicide + NIS	765	5294
ZGS 69	US¶		• _ / ·
ZGS 69	12) US	760	5077
	Mean	761	5180
	$LSD_{0.05}$	NS	NS

Table A.5. Test weight and yield for all treatments in experiment one combined across six environments in 2016 and 2017.

† Timing of application.‡ Non Ionic Surfactant.

<sup>•</sup> § Urea Ammonium Nitrate (28-0-0).

¶ Urea (46-0-0) Solution.

# N-n-butyl thiophosphoric triamide, urease inhibitor.

			Test	Weight			Yield				Protein			
Application Timing <sup>+</sup>	Treatment	Ada	Cass	Pros	Comb.	Ada	Cass	Pros	Comb.	Ada	Cass	Pros	Comb.	
			k	g m <sup>-3</sup>			kg ha <sup>-1</sup>				g kg <sup>-1</sup>			
	1) Control, no Fertilizer	779	758	768	769	3854	4185	4543	4194	125	128	129	127	
	2) Control, with Fertilizer	792	769	765	775	6080	5818	4604	5500	127	129	132	129	
ZGS 65	3) Fungicide, no NIS‡	794	772	768	778	6165	5519	4524	5403	132	124	134	130	
ZGS 65	4) Fungicide + NIS	801	774	765	780	6264	5805	4871	5647	128	136	132	132	
ZGS 65	5) Fungicide + NIS + UAN§	791	768	763	774	5803	5726	4426	5318	134	141	136	137	
ZGS 65	6) Fungicide + UAN + NBPT¶	789	766	762	772	5873	5502	4700	5359	135	132	135	134	
ZGS 65	7) UAN	790	772	766	776	6260	5993	4519	5591	139	142	135	139	
ZGS 65	8) Fungicide + NIS + US#	799	775	766	780	6293	5645	4819	5585	137	133	133	134	
ZGS 65	9) Fungicide + US + NBPT	793	771	763	776	6191	6046	4737	5658	137	141	135	138	
ZGS 65	10) US	790	767	765	774	6321	5591	4378	5430	137	139	135	137	
ZGS 65 ZGS 69	11) Fungicide + NIS UAN	793	773	761	776	6186	5869	4523	5526	137	146	138	141	
ZGS 69	12) UAN	793	767	760	774	6053	5766	4517	5445	138	148	139	142	
ZGS 65 ZGS 69	13) Fungicide + NIS US	795	774	764	778	6257	5781	4703	5580	138	145	138	140	
ZGS 69	14) US	791	764	761	772	5831	5295	4628	5251	136	150	135	141	
	LSD <sub>0.05</sub>	9	6	NS	56	337	767	NS	561	3	8	3	6	

Table A.6. Test weight, yield, and grain protein content influenced by all treatments in experiment one for individual and combined environments in 2017.

† Timing of application.

‡ Non Ionic Surfactant.

§ Urea Ammonium Nitrate (28-0-0).

¶ N-n-butyl thiophosphoric triamide, urease inhibitor.

# Urea (46-0-0) Solution.

	Ada	Cass	Pros	Combined
1) Control, no Fertilizer	0.22	0.23	0.29	0.25
2) Control, with Fertilizer	0.31	0.29	0.30	0.30
3) UAN† Medium Droplet	0.28	0.29	0.29	0.29
4) UAN Coarse Droplet	0.30	0.28	0.29	0.29
5) UAN + MSO <sup>‡</sup> Medium Droplet	0.30	0.22	0.28	0.27
6) UAN + MSO Coarse Droplet	0.28	0.24	0.28	0.26
7) UAN + POC§ Medium Droplet	0.29	0.24	0.27	0.27
8) UAN + POC Coarse Droplet	0.29	0.23	0.29	0.27
9) UAN + NIS¶ Medium Droplet	0.28	0.23	0.26	0.26
10) UAN + NIS Coarse Droplet	0.28	0.22	0.27	0.26
11) UAN + MSOOS# Medium Droplet	0.28	0.22	0.28	0.26
12) UAN+ MSOOS Coarse Droplet	0.27	0.23	0.28	0.26
13) UAN 60:40 Dilution Ratio <sup>††</sup>	0.29	0.26	0.28	0.28
14) UAN 75:25 Dilution Ratio	0.29	0.29	0.28	0.28
15) UAN + NBPT <b>; ;</b>	0.28	0.26	0.26	0.27
16) Urea Solution + NBPT	0.31	0.28	0.27	0.29
17) UAN Streamer Bars at ZGS 65	0.31	0.30	0.31	0.31
18) UAN Streamer Bars at ZGS 65	0.29	0.27	0.28	0.28
Mean	0.28	0.25	0.28	0.27
LSD <sub>0.05</sub>	0.03	0.04	0.02	0.03

Table A.7. Effects of treatment applications on leaf burn measured by normalized difference red edge (NDRE) in 2017 for individual and combined environments in Experiment Two.

† Urea ammonium nitrate (28-0-0).

‡ Methylated seed oil.

§Petroleum oil concentrate.

¶ Nonionic surfactant.

# Methylated seed oil organisilicone surfactant.

†† Dilution ratio of nitrogen solution (UAN:Water)

**‡‡** N-n-butyl thiophosphoric triamide, urease inhibitor

		<u>2(</u>	<u>)16</u>	L.		<u>2</u>	017 DVI	
								<u> </u>
	Cass.	Fargo	Pros.	Comb	Ada	Cass.	Pros.	Comb.
		ç	%					
1) Control, No Treatment					0.77	0.72	0.74	0.74
2) UAN‡ Medium Droplet	37 ab	25 a	33 a	32 ab	0.72	0.68	0.70	0.70
3) UAN Coarse Droplet	35 ab	38 bc	28 a	33 ab	0.75	0.69	0.68	0.71
4) UAN + MSO§ Medium Droplet	33 ab	65 d	78 c	58 c	0.73	0.56	0.68	0.65
5) UAN + MSO Coarse Droplet	33 ab	55 cd	70 bc	53 bc	0.69	0.58	0.67	0.65
6) UAN + POC¶ Medium Droplet	75 d	66 d	33 a	58 c	0.69	0.56	0.70	0.65
7) UAN + POC Coarse Droplet	75 d	63 d	30 a	56 bc	0.70	0.55	0.68	0.65
8) UAN + NIS# Medium Droplet	68 cd	60 d	65 bc	64 c	0.68	0.55	0.67	0.63
9) UAN + NIS Coarse Droplet	60 cd	50 cd	78 bc	56 c	0.69	0.53	0.60	0.61
10) UAN + MSOOS <sup>†</sup> <sup>†</sup> Medium Droplet	63 cd	48 cd	65 bc	58 c	0.67	0.55	0.65	0.62
11) UAN+ MSOOS Coarse Droplet	38 bc	48 cd	60 bc	52 bc	0.71	0.56	0.68	0.65
12) UAN 60:40 Dilution Ratio <sup>‡‡</sup>	40 ab	36 bc	28 a	34 ab	0.73	0.64	0.69	0.68
13) UAN 75:25 Dilution Ratio	40 ab	33 bc	35 a	36 ab	0.72	0.68	0.67	0.69
14) UAN + NBPT§§	25 a	¶¶	35 a	27 ab	0.71	0.64	0.65	0.66
15) Urea Solution + NBPT	25 a	99	20 a	23 a	0.75	0.67	0.66	0.69
Means	47	49	45	47	0.71	0.62	0.68	0.67
##LSD <sub>0.05</sub>					0.03	0.08	0.06	0.06

Table A.8. Effects of treatment applications on leaf burn measured by visual percent leaf burn (2016) and red normalized difference vegetative index (NDVI) (2017) for individual environments and combined within years in experiment two.

<sup>†</sup> Means separation based on transformed values of % leaf burn, which were used to calculate the LSD values.

‡ Urea ammonium nitrate (28-0-0).

§ Methylated seed oil.

¶ Petroleum oil concentrate.

# Nonionic surfactant.

†† Methylated seed oil organisilicone surfactant.

**‡‡** Dilution ratio of nitrogen solution (UAN:Water).

§§ N-n-butyl thiophosphoric triamide, urease inhibitor.

¶ Treatments not included at Fargo in 2016.

## Least significant difference not indicated due to means separation available in table.

	TW	Yield
	kg m <sup>-3</sup>	kg ha <sup>-1</sup>
Control, no treatment	766	5397
UAN§	765	5476
$UAN + MSO\P$	760	5304
UAN + POC#	758	5260
$UAN + NIS^{\dagger\dagger}$	760	5133
UAN + MSOOS <sup>‡‡</sup>	763	5283
Medium Droplet	762	5314
Coarse Droplet	761	5268
§§LSD <sub>0.05</sub>	4	198
¶¶ LSD <sub>0.05</sub>	4	NS

Table A.9. Means for test weight and yield for adjuvant<sup>†</sup> and droplet<sup>‡</sup> treatments combined across all environments in 2016 and 2017 in experiment two.

<sup>†</sup> Means derived from all treatments receiving respective adjuvant regardless of droplet size.

‡ Means derived from all treatments receiving respective droplet size regardless of adjuvant.

§ Urea ammonium nitrate (28-0-0).

¶ Methylated seed oil.

# Petroleum oil concentrate.

†† Nonionic surfactant.

**‡‡** Methylated seed oil organisilicone surfactant.

§§ LSD comparing adjuvant treatments to control.

¶ LSD comparing droplet treatments to control.

	TW	Yield
	kg m <sup>-3</sup>	kg ha <sup>-1</sup>
1) Control, No Treatment	766 a	5397 abc
2) UAN† Medium Droplet	766 a	5526 a
3) UAN Coarse Droplet	765 ab	5426 ab
4) UAN + MSO <sup>*</sup> Medium Droplet	760 bcd	5360 abc
5) UAN + MSO Coarse Droplet	761 abcd	5247 bcd
6) UAN + POC§ Medium Droplet	758 d	5339 abc
7) UAN + POC Coarse Droplet	759 cd	5182 cd
8) UAN + NIS¶ Medium Droplet	761 abcd	5093 d
9) UAN + NIS Coarse Droplet	758 d	5173 cd
10) UAN + MSOOS# Medium Droplet	764 abc	5256 bcd
11) UAN+ MSOOS Coarse Droplet	761 abcd	5310 abcd
12) UAN 60:40 Dilution Ratio††	762 abcd	5479 ab
13) UAN 75:25 Dilution Ratio	763 abcd	5446 ab
14) UAN + NBPT‡‡	758 d	5330 abc
15) Urea Solution + NBPT	762 abcd	5498 a
Me	ean 762	5334

Table A.10. Means for test weight and yield influenced by all treatments combined across all environments in 2016 and 2017 in experiment two.

† Urea ammonium nitrate (28-0-0).

‡ Methylated seed oil.

§Petroleum oil concentrate.

¶ Nonionic surfactant.

# Methylated seed oil organisilicone surfactant.
†† Dilution ratio of nitrogen solution (UAN:Water)

**‡‡** N-n-butyl thiophosphoric triamide, urease inhibitor

	Casse	elton	Pro	sper	Fargo	Ada	Combined		
	2016	2017	2016	2017	2016	2017	2016-2017		
	g kg <sup>-1</sup>								
1) Control, No Treatment	132	129	126	134	132	129	130		
2) UAN† Medium Droplet	140	139	133	139	136	136	137		
3) UAN Coarse Droplet	138	138	133	141	130	134	136		
4) UAN + MSO <sup>+</sup> Medium Droplet	140	138	130	138	132	135	136		
5) UAN + MSO Coarse Droplet	140	136	132	138	134	133	136		
6) UAN + POC§ Medium Droplet	140	136	135	138	129	135	135		
7) UAN + POC Coarse Droplet	138	142	136	138	132	134	137		
8) UAN + NIS¶ Medium Droplet	138	136	130	140	134	134	135		
9) UAN + NIS Coarse Droplet	139	137	132	138	134	135	136		
10) UAN + MSOOS# Medium Droplet	138	132	132	139	132	136	135		
11) UAN+ MSOOS Coarse Droplet	138	138	133	138	133	136	136		
12) UAN 60:40 Dilution Ratio <sup>††</sup>	141	135	134	139	135	135	136		
13) UAN 75:25 Dilution Ratio	143	139	135	140	135	136	138		
14) UAN + NBPT <b>* *</b>	141	138	133	139	##	135	137		
15) Urea Solution + NBPT	138	132	131	138	##	136	135		
§§Range	11	13	10	7	7	7	8		
Mean	139	136	132	138	133	135	136		
LSD <sub>0.05</sub>	4	NS	4	NS	4	4	2		

Table A.11. Grain protein content influenced by all treatments for individual and combined environments in experiment two.

† Urea ammonium nitrate (28-0-0).

‡ Methylated seed oil.

§Petroleum oil concentrate.

¶ Nonionic surfactant.

# Methylated seed oil organisilicone surfactant.

†† Dilution ratio of nitrogen solution (UAN:Water).‡‡ N-n-butyl thiophosphoric triamide, urease inhibitor.

§§ Difference between maximum and minimum values across all treatments for that location.

•	Protein			TW				Yield				
	Ada	Cass.	Pros.	Comb.	Ada	Cass.	Pros.	Comb.	Ada	Cass.	Pros.	Comb.
		g kg <sup>-1</sup>		kg m <sup>-3</sup>			kg ha-1					
1) Control, no Fertilizer	127	123	131	127	778	755	765	766	3819	4728	4723	4423
2) Control, with Fertilizer	129	129	134	131	801	758	762	774	6175	5907	4712	5598
3) UAN <sup>†</sup> Medium Droplet	136	139	139	138	797	761	766	775	6458	6225	4684	5789
4) UAN Coarse Droplet	134	138	141	138	800	756	757	771	6235	5788	4225	5416
5) UAN + MSO <sup>‡</sup> Medium Droplet	135	138	138	137	794	754	764	771	6130	5686	4606	5474
6) UAN + MSO Coarse Droplet	133	136	138	136	796	757	761	771	6020	5531	4409	5320
7) UAN + POC§ Medium Droplet	135	136	138	136	788	757	757	767	6129	5755	4604	5496
8) UAN + POC Coarse Droplet	134	142	138	138	793	741	764	766	6028	5156	4652	5278
9) UAN + NIS¶ Medium Droplet	134	136	140	136	794	755	762	770	6045	5556	4074	5225
10) UAN + NIS Coarse Droplet	135	137	138	137	787	753	761	767	6101	5484	4421	5335
11) UAN + MSOOS# Medium Droplet	136	132	139	135	795	754	765	771	5992	5390	4612	5331
12) UAN+ MSOOS Coarse Droplet	136	138	138	137	793	755	763	770	6285	5516	4393	5398
13) UAN 60:40 Dilution Ratio <sup>††</sup>	135	135	139	136	796	759	760	772	6443	5819	4563	5608
14) UAN 75:25 Dilution Ratio	136	139	140	138	793	761	758	771	6257	5973	4428	5553
15) UAN + NBPT <b>* *</b>	136	138	139	138	796	750	759	769	6253	5570	4288	5370
16) Urea Solution + NBPT	136	132	138	135	796	762	759	773	6529	5838	4759	5709
17) UAN Streamer Bars at ZGS 65	135	128	135	133	797	761	767	775	6317	5696	4349	5454
18) UAN Streamer Bars at ZGS 65	138	128	136	134	796	758	765	773	6181	5465	4419	5355
LSD <sub>0.05</sub>	4	9	4	4	10	12	6	7	369	815	506	577

Table A.12. Test weight, yield, and grain protein content influenced by all treatments for individual and combined environments in 2017 in experiment two.

† Urea ammonium nitrate (28-0-0).

‡ Methylated seed oil.

§ Petroleum oil concentrate.

¶ Nonionic surfactant.

# Methylated seed oil organisilicone surfactant.

*††* Ratio of UAN:Water.

tt N-n-butyl thiophosphoric triamide, urease inhibitor.